## **Supporting Information**

## Prion peptides are extremely sensitive to copper induced oxidative stress

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**Figure S1**. Kinetic profiles of DA (3 mM) oxidation with time (DA autoxidation in brown), in 50 mM Hepes buffer solution at pH 7.4 and 20 °C in the presence of  $Cu^{2+}$  (25  $\mu$ M) (orange) and with addition of PrP<sub>84-114</sub> (8  $\mu$ M, yellow; 25  $\mu$ M, green; 50  $\mu$ M, blue; 75  $\mu$ M, light blue). DA oxidation was monitored through the absorption band of dopaminochrome at 475 nm.



**Figure S2.** Selected absorption spectra recorded during the oxidation of DA (3 mM) in 50 mM Hepes buffer solution at pH 7.4 and 20 °C in the presence of: (A) 25  $\mu$ M copper(II), and (B) 25  $\mu$ M copper(II) and 75  $\mu$ M PrP<sub>76-114</sub>. In (A) and (B), the first spectrum (lower trace) was recorded after 5 s, and the other spectra were recorded every 110 s.

**Table S1**. Computed percentage formation relative to  $Cu^{2+}$  of complex species formed in the  $Cu^{2+}$ -PrP<sub>76-114</sub> system in 50 mM Hepes buffer solution at pH 7.4 and different metal to ligand ratios (M/L). Protonation and  $Cu^{2+}$  complexation constants with PrP<sub>76-114</sub> are from ref. 2S.

M_L	free Cu	CuB	CuH5L	CuH4L	CuH3L	CuH2L	CuHL	CuL	Cu2HL	Cu2L	Cu2H-1L
1_1	0.01	0.59	0.03	3.91	24.70	50.44	16.70	0.25	0.39	0.54	0.61
1_2	0.00	0.01	0.04	4.07	25.70	52.48	17.37	0.26	0.01	0.01	0.02
1_3	0.00	0.01	0.04	4.08	25.71	52.48	17.38	0.26	0.00	0.01	0.01





**Figure S3**. Computed species distribution of the complexes formed in the presence of  $Cu^{2+}$  (25  $\mu$ M) and (A) 25  $\mu$ M PrP<sub>76-114</sub>, (B) 50  $\mu$ M PrP<sub>76-114</sub> and (C) 75  $\mu$ M PrP<sub>76-114</sub>, in 50 mM Hepes buffer solution at pH 7.4. Protonation and  $Cu^{2+}$  complexation constants with PrP<sub>76-114</sub> are from ref. 2S.

**Table S2.** Computed percentage formation relative to  $Cu^{2+}$  of complex species formed in the  $Cu^{2+}$ -PrP<sub>84-114</sub> system in 50 mM Hepes buffer solution at pH 7.4 and different metal to ligand ratios (M/L). Protonation and  $Cu^{2+}$ -complexation constants with PrP<sub>84-114</sub> are from ref. 1S.

	free		CuH	CuH4	CuH3	CuH2			Cu2H	Cu2H2	Cu2H	Cu2	Cu2H	Cu2H	Cu2H
	Cu	CuB	5L	L	L	L	CuHL	CuL	3L	L	L	L	-1L	-2L	-3L
1_1	0.16	6.79	0.02	1.44	9.99	51.20	13.16	0.20	0.01	0.12	1.10	4.17	2.63	0.44	0.01
1_2	0.03	1.27	0.02	1.80	12.44	63.80	16.40	0.25	0.00	0.03	0.26	0.97	0.61	0.10	0.00
1_3	0.02	0.67	0.02	1.85	12.76	65.48	16.83	0.26	0.00	0.02	0.14	0.52	0.33	0.05	0.00





**Figure S4**. Computed species distribution of the complexes formed in the presence of  $Cu^{2+}$  (25  $\mu$ M) and (A) 25  $\mu$ M PrP<sub>84-114</sub>, (B) 50  $\mu$ M PrP<sub>84-114</sub> and (C) 75  $\mu$ M PrP<sub>84-114</sub>, in 50 mM Hepes buffer solution at pH 7.4. Protonation and Cu<sup>2+</sup> complexation constants with PrP<sub>84-114</sub> are from ref. 1S.

**Table S3**. Computed percentage formation relative to  $Cu^{2+}$  of complex species formed in the  $Cu^{2+}$ -PrP<sub>106-114</sub> system in 50 mM Hepes buffer solution at pH 7.4 and different metal to ligand ratios (M/L). Protonation and  $Cu^{2+}$  complexation constants with PrP<sub>106-114</sub> are from ref. 4S.

M/L	free Cu	CuB	CuH2L	CuHL	CuL	CuH-1L	CuH-2L	CuH-3L
1_1	0.89	37.69	0.01	0.29	44.23	16.43	0.04	0.00
1_2	0.40	16.92	0.01	0.40	59.80	22.22	0.05	0.00
1_3	0.24	10.23	0.01	0.43	64.84	24.08	0.06	0.00







**Figure S5**. Computed species distribution of the complexes formed in the presence of  $Cu^{2+}$  (25  $\mu$ M) and (A) 25  $\mu$ M PrP<sub>106-114</sub>, (B) 50  $\mu$ M PrP<sub>106-114</sub> and (C) 75  $\mu$ M PrP<sub>106-114</sub>, in 50 mM Hepes buffer solution at pH 7.4. Protonation and Cu<sup>2+</sup> complexation constants with PrP<sub>106-114</sub> are from ref. 4S.



**Figure S6**. Kinetic profiles of MC (3 mM) oxidation with time, in 50 mM Hepes buffer solution at pH 7.4 and 20 °C in the presence of  $Cu^{2+}$  (25  $\mu$ M) (brown) and with addition of PrP<sub>84-114</sub> (25  $\mu$ M, orange; 50  $\mu$ M, yellow; 75  $\mu$ M, green).



**Figure S7**. Kinetic profiles of MC (3 mM) oxidation with time, in 50 mM Hepes buffer solution at pH 7.4 and 20 °C in the presence of  $Cu^{2+}$  (25  $\mu$ M) (brown) and with addition of PrP<sub>106-114</sub> (15  $\mu$ M, orange; 25  $\mu$ M, light green; 35  $\mu$ M, yellow; 50  $\mu$ M, green; 75  $\mu$ M, light blue).



**Figure S8**. Kinetic traces of absorbance at 401 nm *vs*. time for the oxidation of MC (3 mM) at pH 7.4 and 20 °C in the presence of free Cu<sup>2+</sup> (25  $\mu$ M) (orange trace), PrP<sub>76-114</sub> (25  $\mu$ M) (green trace), Cu<sup>2+</sup> (25  $\mu$ M) and PrP<sub>76-114</sub> (25  $\mu$ M) (light blue trace) in Hepes buffer (5 mM). MC autoxidation is shown by the yellow trace.



**Figure S9**. Selected absorption spectra recorded during the oxidation of MC (3 mM) in 5 mM Hepes buffer solution at pH 7.4 and 20 °C in the presence of : (A) 25  $\mu$ M copper(II), and (B) 25  $\mu$ M copper(II) and 25  $\mu$ M PrP<sub>76-114</sub>. In (A) and (B), the first spectrum (lower trace) was recorded after 5 s, and the other spectra were recorded every 360 s.



**Figure S10**. Kinetic traces of MC (3 mM) oxidation with time, in 50 mM Hepes buffer solution at pH 7.4 and 20 °C in the presence of  $Cu^{2+}$  (25  $\mu$ M, light blue trace) and with addition of PrP<sub>76-114</sub> (50  $\mu$ M, yellow trace). A similar experiment was carried out with the solvent saturated with pure oxygen with  $Cu^{2+}$  (25  $\mu$ M, orange trace) and with addition of PrP<sub>76-114</sub> (50  $\mu$ M, green trace).



**Figure S11.** Dependence of the reaction rates of 4-methylquinone formation on the concentration of MC. The reactions were performed in Hepes buffer (50 mM) pH 7.4, at 20°C, in the presence of free copper(II) (white circles),  $[Cu^{2+}-PrP_{106-114}]$  complex (25  $\mu$ M) (orange circles),  $[Cu^{2+}-PrP_{84-114}]$  complex (25  $\mu$ M) (light blue circles), and  $[Cu^{2+}-PrP_{76-114}]$  complex (25  $\mu$ M) (green circles). Solid lines correspond to fitting of experimental data with Michaelis-Menten equation.



**Figure S12.** Kinetic traces of MC (3 mM) oxidation with time, in 50 mM Hepes buffer solution at pH 7.4 and 20 °C in the presence of  $Cu^{2+}$  (25  $\mu$ M, light blue trace) and with addition of PrP<sub>76-114</sub> (25  $\mu$ M, yellow trace). Similar experiments were carried out with 1 mM H<sub>2</sub>O<sub>2</sub> with both  $Cu^{2+}$  (25  $\mu$ M, green trace) and  $Cu^{2+}$  (25  $\mu$ M) with addition of PrP<sub>76-114</sub> (25  $\mu$ M, orange trace). Blank experiment in the presence of MC and H<sub>2</sub>O<sub>2</sub> is shown by the blue trace.



Scheme S1. Reaction of histidine residues of PrP peptides with MQ and subsequent oxidation of the resulting adduct to quinone.



**Figure S13**. HPLC-MS elution profiles of peptides resulting from oxidation of MC (3 mM) by Cu-PrP<sub>84-114</sub> (25  $\mu$ M) at 60 min reaction time in 50 mM Hepes buffer at pH 7.4 and 20 °C. The assignment of the peaks is shown (MQ indicates a mass increment of 120 Da corresponding to the formation of a covalent adduct with 4-methylquinone).

$25 \mu\text{M}$ copper(II) nitrate and 3 mM MC in Hepes buffer (50 mM) pH 7.4 at 20 °C.						
Time (min)	PrP <sub>84-114</sub>	+	16	+120	+136	+152
	(%)	(%	6)	(%)	(%)	(%)
	$(R_t 31')$	(R <sub>t</sub> 32')	(R <sub>t</sub> 29')	(R <sub>t</sub> 35')	(R <sub>t</sub> 36')	(R <sub>t</sub> 37')
1	95.6	1.2	0.4	1.8	0.4	0.6
10	80.0	3.4	3.5	5.8	5.4	1.9
15	72.7	8.9	2.4	7.6	6.2	2.2
60	59.0	7.7	8.3	6.9	11.7	6.4
90	44.9	8.9	1.7	9.0	21.7	13.8
100	11.0	14.5	4.1	13.7	28.0	28.7

**Table S4**. Modifications of  $PrP_{84-114}$  peptide (25  $\mu$ M) detected by LC-MS analysis, in the presence of 25  $\mu$ M copper(II) nitrate and 3 mM MC in Hepes buffer (50 mM) pH 7.4 at 20 °C.



**Figure S14**. HPLC-MS elution profiles of peptides resulting from the oxidation of MC (3 mM) by Cu-PrP<sub>106-114</sub> (25  $\mu$ M) at 100 min reaction time in 50 mM Hepes buffer at pH 7.4 and 20 °C. The assignment of the peaks is shown (MC and MQ indicate mass increments of 122 and 120 Da, corresponding to the formation of covalent adducts with 4-methylcatechol and 4-methylquinone, respectively).

25 µM c	opper(II) nit	rate and 3 r	nM MC in F	lepes buffer	(50 mM) pH 7	7.4 at 20 °C.	
Time	PrP <sub>106-114</sub>	+16	+32	+120	+122	+136	+138
(min)	(%)	(%)	(%)	(%)	(%)	(%)	(%)
	$(R_t 11')$	$(R_t 21')$	(R <sub>t</sub> 19')	(Rt 28')	(R <sub>t</sub> 29')	(Rt 23')	(R <sub>t</sub> 25')
10	82.7	2.8	0.2	6.5	6.4	0.5	0.9
20	67.7	9.0	1.7	10.2	3.9	1.5	6.0
30	60.6	9.5	0.3	13.7	11.8	2.7	1.4
55	47.0	23.2	3.6	7.4	8.1	7.4	3.3
100	35.1	34.9	5.1	4.8	7.8	7.9	4.4
120	21.1	36.8	8.3	4.9	7.5	14.2	7.2

**Table S5**. Modifications of  $PrP_{106-114}$  peptide (25 µM) detected by LC-MS analysis, in the presence of 25 µM copper(II) nitrate and 3 mM MC in Hepes buffer (50 mM) pH 7.4 at 20 °C.



**Figure S15**. Proton NMR spectrum of Hepes buffer (5 mM) pH 7.4 and MC (3 mM) in  $D_2O$  at 20 °C. The Hepes buffer signal of the  $-CH_2$ - group bound to sulfonic acid group at 3.7 ppm was used as internal standard.



**Figure S16.** Enlargement of the aromatic region of the proton NMR spectra of MC (3 mM) and Hepes buffer (5 mM) pH 7.4 in D<sub>2</sub>O at 20 °C (gray trace), and after 2 h of reaction in the presence of copper(II) nitrate (25  $\mu$ M) (red trace) and Cu<sup>2+</sup>-PrP<sub>76-114</sub> (25  $\mu$ M) (blue trace).

Time (min)	[MC] (mM)					
	$Cu^{2+}$	$Cu^{2+}-PrP_{76-114}$				
0	3.00	3.00				
5	$2.94\pm0.08$	$2.55 \pm 0.02$				
30	$2.82\pm0.09$	$2.36\pm0.06$				
60	$2.68\pm0.10$	2.17 ± 0. 10				
90	$2.53\pm0.06$	$1.88\pm0.04$				
120	$2.44\pm0.02$	$1.55\pm0.07$				
150	$2.34\pm0.04$	$1.32 \pm 0.04$				

**Table S6**. Determination of residual MC concentration by NMR upon oxidation of MC (3 mM) by  $Cu^{2+}$  and  $Cu^{2+}$ -PrP<sub>76-114</sub> (25  $\mu$ M) in 5 mM Hepes deuterated buffer at pH 7.4 and 20 °C.





**Figure S17.** Plots of UV-Vis absorbance at 560 nm for the NBT reduction to  $MF^+$  by  $O_2^-$  in the presence of  $Cu^{2+}$  (black circles) and  $Cu^{2+}$  complexes (white circles) with  $PrP_{106-114}$  (A),  $PrP_{84-114}$  (B) and  $PrP_{76-114}$  (C). Spectra were taken at 20 °C, in 50 mM phosphate buffer at pH 7.4.