

Presentation and outcome with second line treatment in AL amyloidosis previously sensitive to non-transplant therapies

Giovanni Palladini¹, Paolo Milani¹, Andrea Foli¹, Marco Basset¹, Francesca Russo¹, Stefano Perlini^{1,2}, Giampaolo Merlini¹

¹Amyloidosis Research and Treatment Center, Fondazione Istituto di Ricovero e Cura a Carattere Scientifico (IRCCS) Policlinico San Matteo, and Department of Molecular Medicine, University of Pavia, Pavia, Italy

²Department of Internal Medicine, Fondazione IRCCS Policlinico San Matteo and University of Pavia, Pavia, Italy

Corresponding author:

Giampaolo Merlini, MD

Amyloidosis Research and Treatment Center

Fondazione IRCCS Policlinico San Matteo

Viale Golgi, 19 – 27100 Pavia, Italy

telephone: +39-0382-502994

fax: +39-0382-502990

e-mail: gmerlini@unipv.it

Text word count: 3918/4000

Abstract word count: 229

Figures/Tables count: 7/7

References: 48/100

Scientific Category: Clinical Trials and Observations

Key points

- Exposure to melphalan and bortezomib and quality of response to upfront treatment prolong time to second-line therapy in AL amyloidosis.
- Patients who need second-line therapy after initial response have a good outcome if they are rescued before cardiac progression.

Abstract

The management of light chain (AL) amyloidosis has improved in recent years thanks to accurate biomarker-based staging systems and response criteria and availability of novel effective therapies. However, previous studies have focused on newly diagnosed patients, and little is known on relapsed patients, despite trials of new agents are often performed in this setting. In the present study we report the outcome of 259 patients who responded to upfront therapy. Ninety-two patients (35%) needed second-line therapy after a median of 49 months. Cardiac and renal progression were observed in 22% and 12% of patients who received second-line therapy, respectively. Complete response after upfront treatment and frontline therapy with combined bortezomib, melphalan and dexamethasone independently prolonged time to second-line therapy. Median survival of relapsing patients was 59 months. Patients who had a “high-risk dFLC progression,” which we defined as a difference between involved and uninvolved FLC (dFLC) of >20 mg/L, a level >20% of baseline value, and a >50% increase from the value reached at best response, had a shorter survival after initiation of second-line therapy on univariate, but not on multivariate analysis, where cardiac progression was the only independent predictor of survival after starting rescue treatment. Patients with AL amyloidosis who need second-line therapy after response to upfront treatment generally have a good outcome. A “high-risk dFLC progression” should trigger rescue treatment while cardiac progression should not be awaited.

Introduction

The introduction of biomarkers of clonal and organ disease has greatly improved the management of patients with light-chain (AL) amyloidosis, allowing accurate treatment selection based on risk stratification.¹ Our understanding of the biology of the amyloidogenic plasma cell clone is also improving,²⁻⁴ leading to better ability to predict patients' outcome based on multiparametric flow cytometry,⁵ revealing differential susceptibility to melphalan and bortezomib based on cytogenetic abnormalities,⁶⁻⁹ and indicating features that can be exploited for the design of targeted therapies,^{10,11} moving towards a more personalized treatment approach. These advancements, coupled with the availability of new effective drugs, resulted in a significant improvement of patients outcome over the years.^{12,13} Moreover, the availability of validated criteria for early assessment of hematologic and organ response, that can reliably predict overall survival and progression to dialysis^{14,15} and have been proposed as surrogate endpoints for clinical trials,¹⁶ are accelerating the development of novel therapeutic agents. Currently, upfront treatment is based on autologous stem cell transplant (ASCT) and bortezomib combinations,¹⁷⁻¹⁹ while an increasing number of novel agents, such as novel proteasome inhibitors, third-generation immunomodulatory drugs or anti plasma cell antibodies, are being tested in relapsed and refractory patients.²⁰⁻²⁵ Moreover, different therapeutic approaches, targeting the amyloid deposits are being developed.²⁶

However, most of currently available data on the natural history and outcome of AL amyloidosis refer to newly-diagnosed, treatment-naïve patients, while little is known on clinical presentation and prognostic factors at the time of relapse, although three studies recently reported patterns of relapse after autologous stem cell transplantation.²⁷⁻²⁹ This is particularly relevant because AL amyloidosis is still characterized by a high rate of early deaths, while long-term outcome is superior to that of multiple myeloma,³⁰ and relapsed and refractory patients are selected for a relatively good outcome. This lack of knowledge results in the absence of validated criteria of progression, in lack of uniformity in the timing of re-treatment in relapsing patients across referral centers,³¹ and in varying, non-comparable reporting of progression free survival in clinical trials. This is acutely relevant, because most of clinical trials of new drugs take place in the relapsed and refractory setting.

In the present study, we report the outcome, variables leading to second-line therapy initiation, and variables predicting survival after rescue treatment in 259 consecutive patients with AL amyloidosis who responded to upfront non-transplant chemotherapy.

Methods

The prospectively-maintained database of the Pavia Amyloidosis Research and Treatment Center was systematically searched for patients with AL amyloidosis who responded to upfront chemotherapy. Response to upfront chemotherapy was deemed satisfactory in case any of the following was reached: 1) complete response (CR), 2) very good partial response (VGPR), 3) partial response (PR) plus organ response.^{16,32-34} Patients who received ASCT upfront were excluded. All patients gave written informed consent for the use of their clinical data for research purposes according to the institutional review board guidelines. The diagnosis of AL amyloidosis was biopsy-proven and the deposits were characterized as AL type by immuno-electron microscopy³⁵ or mass spectrometry³⁶ in all cases.

The patients were stratified according to the modified 2004 Mayo Clinic staging system,^{37,38} based on N-terminal pro-natriuretic peptide type-B (NT-proBNP, cutoff 332 ng/L) and cardiac troponin I (cTnI, cutoff 0.1 ng/mL), with stage I, II, and III patients having none, one or both markers above the cutoff. Stage III patients were classified as stage IIIa or IIIb based on whether their NT-proBNP was below or above 8500. Staging of renal involvement was based on estimated glomerular filtration rate (eGFR, cutoff 50 mL/min per 1.73 m²) and proteinuria (cutoff 5 g/24h), with stage I, II, and III patients having non, one, or both unfavorable markers.¹⁵

Hematologic, cardiac, and renal responses were assessed according to current validated criteria.^{14,15} Complete response required negative serum and urine immunofixation and normal free light chain (FLC) ratio, VGPR was defined as a difference between involved (amyloidogenic) and uninvolved FLC (dFLC) <40 mg/L, and PR as a >50% decrease in dFLC. A pre-treatment dFLC >50 mg/L was required to assess VGPR and PR. Cardiac response was defined as a decrease both >30% and >300 ng/L of NT-proBNP in patients with a pre-treatment NT-proBNP of at least 650 ng/L. Renal response required a >30% decrease in 24 hour proteinuria or a reduction of proteinuria below 0.5 g/24 hours in the absence of a >25% decrease in eGFR in subjects whose pre-treatment proteinuria was >0.5 g/24h.

For the purpose of the present study, relapse was defined as initiation of second-line therapy. The decision to start rescue treatment was made after review of case by at least 2 physicians of the Pavia Amyloidosis Research and Treatment Center. The variables considered to decide re-treatment were organ progression, absolute dFLC level before

initiation of upfront therapy, and change in dFLC compared to the nadir reached after upfront therapy. Time from discontinuation of upfront therapy to initiation of second-line therapy was defined treatment free survival (TFS). Clonal (bone marrow plasma cell infiltrate, dFLC, involved FLC concentration, ratio of involved and uninvolved FLC, and depth of hematologic response) and organ markers (stage, concentration of NT-proBNP, cTnI, eGFR, proteinuria, ventricular wall thickness and ejection fraction at echocardiography), as well as type of frontline therapy were tested for their ability to predict TFS. Receiver Operator Characteristic (ROC) analyses based on initiation of second-line therapy at 2 years were used to identify cutoffs of tested variables best predicting this endpoint. A different set of ROC analyses based on death within 2 years from initiation of second-line therapy was used to identify cutoffs of tested variables best predicting survival after initiation of second-line therapy. Overall survival (OS) was measured from diagnosis in the whole cohort and from the time of second-line therapy initiation in a separate analysis of patients who needed second-line therapy. The analysis of factors affecting the outcome after initiation of second line therapy was based on OS after second-line therapy initiation. Survival curves were plotted according to Kaplan Meier, and differences in survival were tested for significance with the log-rank test. Cox multivariate models were fitted including variables that predicted TFS and OS at univariate analysis. Continuous variables are presented as median and interquartile range (IQR). MedCalc Statistical Software version 14.12.0 (MedCalc Software bvba, Ostend, Belgium; <http://www.medcalc.org>; 2014) was used for computation.

Results

Between 2006 and 2015, 865 newly-diagnosed treatment-naïve patients were seen at the Pavia Amyloidosis Research and Treatment Center. Twenty-two patients were excluded because they received ASCT upfront. Of the remaining patients, 208 died before the evaluation of response and 180 were refractory to upfront therapy. In 196 cases best response to upfront therapy was deemed unsatisfactory, and these patients immediately proceeded to second-line therapy. Levels of FLC and markers of organ involvement in the 196 patients who required second line therapy immediately after discontinuation of upfront chemotherapy due to unsatisfactory response are reported in Supplemental Table 1. The remaining 259 patients discontinued therapy after attainment of response to upfront therapy and were included in the study. Their median age was 66 years (range 39-84 years), and 138 subjects (53%) were males. The heart was involved in 184 patients (71%), the kidney in 179 (69%), the soft tissues in 46 (18%), the peripheral nervous system in 34 (13%), and the liver in 29 (11%). At the time of diagnosis, cardiac stage was I in 75 subjects (29%), II in 119 (46%), IIIa in 47 (18%), and IIIb in 18 (7%). Renal stage was I in 124 patients (48%), II in 101 (39%), and III in 34 (13%). Upfront treatment was melphalan and dexamethasone (MDex) in 129 patients (50%), cyclophosphamide, bortezomib and dexamethasone (CyBorD) in 71 (27%), bortezomib plus MDex (BMDex) in 46 (18%), bortezomib and dexamethasone (BDex) in 10 (4%), and rituximab plus BDex in 3 subjects (1%) with IgM-AL amyloidosis. The criteria guiding the choice of upfront therapy at our center have been reviewed in ¹. Briefly, MDex was preferred in patients with neuropathy or other contraindications to bortezomib, and cyclophosphamide was preferred over melphalan in patients with renal failure and in those with potentially reversible contraindications to ASCT. Best hematologic response after upfront therapy was CR in 82 patients (32%), VGPR in 134 (52%), and PR in 43 (16%). Cardiac response was achieved in 38% of patients and renal response in 27%. All patients in whom treatment was discontinued after achievement of PR had also achieved organ response.

Patterns leading to initiation of rescue therapy

After a median follow-up of living patients of 41 months, 92 subjects (35%) needed second-line therapy. Values of dFLC and markers of cardiac and renal involvement in patients who required second line therapy are reported in Table 1.

At the time of rescue therapy initiation, median dFLC was 55 mg/L (IQR 26-108 mg/L, >20 mg/L in 80% of patients), corresponding to 41% of baseline value (IQR 19-84%, >20% in 74% of patients) and to a 237% increase from the value reached at best response (IQR 54-538%, >50% in 76% of patients). Importantly, dFLC at the time of second-line therapy initiation remained lower than the threshold of measurable disease (50 mg/L) in 44 patients (48%). However, when considered the new threshold proposed for assessing “low-dFLC response” (20 mg/L),^{39,40} 75 patients (81%) had measurable disease.

Progression of NT-proBNP was observed in 20 patients (22%). Response to upfront therapy in these patients was CR in 5 (25%), VGPR in 11 (55%), and PR in 4 (20%). At the time of cardiac progression dFLC had reached a median of 50% of baseline value (IQR 20-83%), had increased by a median of 402% from the value reached at best response (IQR 121-1201%), and was >20 mg/L in 17 patients (85%). In 4 patients dFLC increase and cardiac progression were noted simultaneously, at the same evaluation; whereas, in 15 subject dFLC increase preceded cardiac progression by a median of 6 months (range 2-8 months, Figure 1). However, in one patient NT-proBNP progression occurred with stable dFLC. In this subject, NT-proBNP increased from 3034 ng/L (achieved at best response) to 6544 ng/L 16 months after upfront therapy discontinuation with stable dFLC (baseline 90 mg/L, best response 0 mg/L with positive serum and urine immunofixation, time of second-line therapy 0 mg/L with positive serum and urine immunofixation). In this patient, baseline NT-proBNP was 7049 ng/L, cardiac response was reached after 8 cycles of CyBorD, eGFR was stable, and no other causes of NT-proBNP increase were identified. The patient died due to heart failure 5 months after NT-proBNP progression. There was no other known cause of heart disease.

In 11 patients (12%) estimated glomerular filtration rate (eGFR) decreased by >25%. Renal progression was associated with a dFLC increase to greater than 20 mg/L in all cases [median 40% of baseline value (IQR 22-76%), median 225% increase from the value reached at best response (IQR 60-1030%)]. A >50% increase in proteinuria to >0.5 g/24 was observed in 17 patients (18%). In all of them dFLC also increased to >20 mg/L, reaching a median of 41% of the baseline value (IQR 24-71%) and with a median increase of 118% compared to the value reached at best response (IQR 24-201%). Notably, amongst the 167 patients in whom rescue treatment was not deemed necessary, 8 (5%) had renal progression defined as a >25% decrease in eGFR, with an increase in dFLC from the value reached at best response that was <20% in all cases.

Treatment-free survival

Median TFS was 49 months (Figure 2). The only baseline clonal marker predicting earlier initiation of second-line therapy was high involved/uninvolved FLC ratio (iuFLCR). The cutoff best predicting TFS was 6 (56% vs. 72% at 3 years, $P=0.027$, Figure 3A). Markers of severity of organ involvement at diagnosis did not predict TFS in responders to upfront therapy. Treatment with BMDex granted a longer TFS (Figure 3B) compared to MDex (80% vs. 56% at 3 years, $P=0.022$) and CyBorD (80% vs. 57% at 3 years, $P=0.027$). The quality of hematologic response after upfront therapy significantly affected TFS, with 25%, 39%, and 62% of patients who achieved CR, VGPR, and PR after upfront treatment requiring second line therapy at 3 years, respectively (Figure 3C). At multivariate analysis, achievement of CR after upfront therapy and frontline treatment with BMDex remained independent predictors of prolonged TFS (Table 2).

Second line therapy

Second-line therapy was BDex in 22 patients (24%) 13 of whom (59%) reached hematologic response, CyBorD in 22 (24%) with 16 (73%) responders, MDex in 19 (21%) with 14 (74%) responders, lenalidomide-based in 11 (12%) with 6 (54%) responders, thalidomide-based in 10 (11%) with 6 (60%) responders, high-dose dexamethasone in 5 (5%) with 3 (50%) responders, BMDex in 2 (2%) both responders, and ASCT in 1 (1%) in whom hematologic response was restored. Patients who had become transplant-eligible were offered ASCT. Patients who received upfront MDex were considered for bortezomib-based combinations unless contraindications to bortezomib persisted. In other cases, we repeated upfront therapy if possible. If this was not possible, patients received lenalidomide, or thalidomide in case of nephrotic syndrome or renal failure.

Survival after initiation of second-line therapy

Overall, 32 patients died (12% of the whole cohort, 35% of patients who underwent rescue therapy). All deaths occurred in patients who needed second line treatment. Median survival of the whole cohort was 99 months (Figure 4A); whereas, median survival from the time of second-line therapy initiation of the 92 patients who underwent rescue therapy was 59 months (Figure 4B). Complete response, upfront treatment with BMDex, and iuFLCR <6 were

not associated with survival after second-line therapy initiation. Moreover, re-challenge with the same therapy used upfront (54 patients, 59%) and treatment with a different therapy (38 patients, 41%) did not affect survival from the time of second-line therapy initiation (median 59 vs. 52 months, $P=0.709$), in agreement with the recent observation by Tandon and coworkers.⁴¹

The ROC analyses showed that dFLC cutoffs best predicting death after second-line therapy initiation were an absolute value of >20 mg/L, a level $>20\%$ of baseline value, and an increase from the value reached at best response of $>50\%$. Based on these findings, we defined as “high-risk dFLC progression” having reached all of these cutoffs. The dFLC level in the 60 patients with “high-risk dFLC progression” was above the standard threshold of measurable disease (>50 mg/L) in 70% of cases, while, by definition, the updated threshold for patients with low dFLC burden (>20 mg/L)^{39,40} was reached in all cases. Amongst patients who required rescue therapy, “high-risk progression” occurred in 56% of those who had achieved CR after upfront therapy, in 65% of patients in VGPR ($P=0.402$ compared to CR), and in 81% of patients in PR ($P=0.089$ compared to CR, $P=0.231$ compared to VGPR). As reported above, “high-risk dFLC progression” was associated with cardiac progression in 85% of cases (Figure 1). Patients with NT-proBNP progression at the time of second-line treatment had a significantly shorter survival after rescue treatment was started (median 17 vs. 62 months, $P=0.002$, Figure 5A), as well as those with “high-risk dFLC progression” (median 46 months vs. not reached, $P=0.004$, Figure 5B).

We assessed the impact of type and severity of organ involvement at diagnosis on survival after second-line therapy initiation. Renal involvement and renal stage at presentation did not affect survival after rescue treatment. There was a non-significant trend for a shorter survival for patients who presented with heart involvement (median 47 months vs. not reached, $P=0.085$). Advanced (stage III) heart involvement at diagnosis did not predict survival after rescue therapy initiation (median 45 vs. 58 months, $P=0.537$). The number of organs involved at the time of second-line therapy initiation did not predict survival.

At multivariate analysis only NT-proBNP progression (HR 4.15, 95%CI 1.72-10.00, $P=0.002$), and not “high-risk dFLC progression” (HR 2.19, 95%CI 0.69-7.18, $P=0.198$) and heart involvement at diagnosis (HR 1.14, 95%CI 0.30-4.34) remained an independent predictor of survival in patients who required rescue therapy.

Discussion

In the present study we elucidated the clinical characteristics and outcome of a large series of patients with AL amyloidosis who required second-line therapy after initial response to chemotherapy. We identified factors predicting shorter time to second-line therapy, and we identified variables associated with a poor outcome after second-line therapy initiation. The relatively long follow-up (median 3.4 years for living patients) allowed a reliable assessment of outcome. The median TFS of patients who achieved a satisfactory response (CR, VGPR, or PR plus organ response) to upfront therapy by Kaplan-Meier analysis was 4.1 years in the overall population and 6.2 years in patients who achieved CR after upfront therapy. In our study, the patients who needed further treatment received second-line therapy after a median of 2.7 years (range 1.2-9.1 years, IQR 1.6-3.9 years) in the overall series and of 3.9 years (range 1.2-10.1 years, IQR 2.1-5.5 years) in patients who achieved CR. This is in agreement with the study from the Boston University group, reporting a median time to hematologic relapse of 4.3 years (range 1.5-21.6 years) in patients who achieved CR after ASCT,²⁸ and with the study from the Mayo Clinic group, reporting that the median time from ASCT to initiating a second-line therapy was 2.1 years (IQR 0.9-4.1 years).²⁹

Outcome of patients who respond to upfront therapy: implications for clinical trial design

In the present study, the OS of patients who achieved PR plus organ response, VGPR or CR after frontline therapy, was very good, with a median exceeding 8 years from diagnosis. Also patients who needed rescue treatment after an initial response to upfront treatment maintain quite a good outcome, with a 5 year median OS after initiation of rescue treatment. This is in agreement with the results of a study by Warsame and Colleagues, reporting a 4.3 year median OS after relapse in 146 patients who received upfront ASCT.²⁷ In the study by Sanchorawala, et al. focusing on patients relapsing after an initial CR induced by ASCT, the OS from the time of relapse was 8.5 years.²⁸ Overall, these data compare favorably with refractory patients. In two small series of homogeneously treated subjects with refractory AL amyloidosis who received lenalidomide- or pomalidomide-based rescue treatment, median OS was 1.2 and 2.2 years, respectively.^{22,42} Overall, these findings indicate that relapsing patients are selected for favorable prognostic factors: survival in the first months after diagnosis, when most patients with advanced heart involvement succumb

to the disease, and sensitivity of the plasma cell clone to chemotherapy. This should be considered when designing clinical trials of new drugs in previously treated patients, that usually enroll both relapsed and refractory patients. These two groups have very different outcomes and appropriate stratification should be planned to avoid biased results.

Characteristics of patients who receive second-line therapy

Despite the lack of validated criteria for disease progression in AL amyloidosis, the present study revealed some uniformity in the changes triggering rescue treatment at our center. Importantly, no deaths were observed in subjects who were believed to have no need of second-line therapy.

We were able to identify factors predicting time to second-line therapy initiation. It was reassuring that the depth of hematologic response as assessed by current criteria also predict TFS, and this is the first validation of the ability of current response criteria to predict progression, further corroborating their utility as endpoints in AL amyloidosis. However, novel prognostic factors for progression also emerged. They were high iuFLCR and exposure to both melphalan and bortezomib upfront. High iuFLCR reflects suppression of non-clonal plasma cells and its relevance is in agreement with the observation that immunoparesis is a marker of poor prognosis in AL amyloidosis.⁴³ Treatment with BMDex conferred a prolonged TFS in this series that was able to overcome the impact of high iuFLCR and was independent of the quality of hematologic response. It is possible that this combination can remain highly effective independently of the cytogenetics abnormalities gain 1q21 and t(11:14) that reduce the efficacy of melphalan- and bortezomib-based therapy, respectively.^{6-8,44} However, data on immunoparesis as defined by Muchtar, et al.⁴³ and results of fluorescence in situ hybridization studies were not available in the present series.

Outcome after initiation of second-line therapy

In the present study, patients who required second-line therapy had a relatively low dFLC level (median 55 mg/L). We and the Heidelberg group have recently shown that approximately 20% of patients with AL amyloidosis have a low dFLC value (<50 mg/L) at diagnosis.^{39,40} This is associated with a longer survival compared to other patients. Nevertheless, reduction to very low levels (below 10 mg/L in subjects who have at least 20 mg/L at baseline, defined as low-dFLC response) results in even better survival and improved

renal outcome. Moreover, in a pilot study of evaluation of minimal residual disease (MRD) by flow cytometry in AL amyloidosis, we observed that the persistence of MRD can prevent organ improvement in patients otherwise in CR.⁴⁵ In the present study, relatively small increases in the absolute dFLC value preceded by several months cardiac progression. Taken together, these observations indicate that even small amounts of circulating amyloidogenic free light chains can be able to foster organ progression in AL amyloidosis, and should not be underestimated. Novel sensitive methods to identify and measure monoclonal amyloidogenic light chains will improve our ability to monitor this small clones in the future.⁴⁶⁻⁴⁸ Almost two thirds of patients who started rescue therapy had at least a 50% increase in dFLC from the value reached after upfront therapy to an absolute value of ≥ 20 mg/L that corresponded to at least 20% of the baseline value observed at diagnosis. This was defined “high-risk dFLC progression”. “High-risk dFLC progression” preceded cardiac progression by a median of 6 months in 85% of cases and was associated with a significantly shorter survival after second-line therapy initiation. Moreover, initiation of rescue therapy after cardiac progression was associated with a median survival of only 17 months. This emphasizes the prognostic relevance of NT-proBNP progression also in the relapsing setting. Importantly, multivariate analysis showed that the impact of dFLC increase on survival was not independent of cardiac progression. These data indicate that cardiac progression should not be awaited to start rescue therapy. A “high-risk dFLC progression” defined as in the present study could probably be considered a trigger to start second-line therapy, also taking into account that the novel hematologic response criteria for patients with low dFLC level, requiring a dFLC above 20 mg/L to have “measurable disease”, can be applied to all subjects with “high-risk dFLC progression”.^{39,40} However, validation in independent series is warranted.

In the present study, a total of 19 patients had renal progression during follow-up. Rescue treatment was started in 11 of them with more pronounced dFLC increases, all fulfilling the criteria of “high-risk dFLC progression”. It should be kept in mind that in AL amyloidosis renal progressions can occur also in the absence of hematologic progression, particularly in patients with advanced renal failure. Thus, renal progression should not always be considered a trigger for rescue treatment initiation.

Conclusion

Patients with AL amyloidosis who need rescue treatment after response to upfront therapy generally have a very good outcome, that is better than that reported for refractory patients. This indicates that clinical trials in relapsed/refractory patients with AL amyloidosis require appropriate stratification based on response to previous therapy. Depth of hematologic response and exposure to melphalan and bortezomib upfront delay progression. A “high-risk dFLC progression” could be considered a trigger for rescue therapy initiation before cardiac progression which is associated with poor survival. However, the generalizability of the results of the present study in independent populations treated upfront with different approaches, including ASCT, is warranted through large, international validation studies of “high-risk dFLC progression” as a possible hematologic progression criterion in AL amyloidosis.

Acknowledgements

This study was supported in part by grant from “Associazione Italiana per la Ricerca sul Cancro - Special Program Molecular Clinical Oncology 5 per mille n. 9965”, from CARIPLO “Structure-function relation of amyloid: understanding the molecular bases of protein misfolding diseases to design new treatments n. 2013-0964”, and from CARIPLO "Molecular mechanisms of Ig toxicity in age-related plasma cell dyscrasias n. 2015-0591."

GP is supported in part by the Bart Barlogie Young Investigator Award from the International Myeloma Society (IMS).

We acknowledge the study coordinator and data manager Anna Carnevale Baraglia.

Contributors

GP and PM designed the study, evaluated patients, collected data, analyzed data, wrote the manuscript and gave final approval.

GM designed the study, evaluated patients, critically reviewed the manuscript, and gave final approval.

MB, FR, AF, PB and SP evaluated patients, collected data, critically reviewed the manuscript, and gave final approval.

Conflict of interest

GP received honoraria from Janssen-Cilag, honoraria and travel support from Prothena and travel support from Celgene.

GM is consultant for Millennium Pharmaceuticals, Inc., Pfizer, Janssen, Prothena, and IONIS.

References

1. Palladini G, Merlini G. What is new in diagnosis and management of light chain amyloidosis? *Blood*. 2016;128(2):159-168.
2. Paiva B, Martinez-Lopez J, Corchete LA, et al. Phenotypic, transcriptomic, and genomic features of clonal plasma cells in light-chain amyloidosis. *Blood*. 2016;127(24):3035-3039.
3. da Silva Filho MI, Försti A, Weinhold N, et al. Genome-wide association study of immunoglobulin light chain amyloidosis in three patient cohorts: comparison with myeloma. *Leukemia*. 2017;31(8):1735-1742.
4. Meziane I, Huhn S, da Silva Filho MI, et al. Genome-wide association study of clinical parameters in immunoglobulin light chain amyloidosis in three patient cohorts. [published online ahead of print 4 July 2017]. *Haematologica*. doi: 10.3324/haematol.2017.171108.
5. Muchtar E, Jevremovic D, Dispenzieri A, et al. The prognostic value of multiparametric flow cytometry in AL amyloidosis at diagnosis and at the end of first-line treatment. *Blood*. 2017;129(1):82-87.
6. Bochtler T, Hegenbart U, Kunz C, et al. Gain of chromosome 1q21 is an independent adverse prognostic factor in light chain amyloidosis patients treated with melphalan/dexamethasone. *Amyloid*. 2014;21(1):9-17.
7. Bochtler T, Hegenbart U, Kunz C, et al. Translocation t(11;14) is associated with adverse outcome in patients with newly diagnosed AL amyloidosis when treated with bortezomib-based regimens. *J Clin Oncol*. 2015;33(12):1371-1378.
8. Bochtler T, Hegenbart U, Kunz C, et al. Prognostic impact of cytogenetic aberrations in AL amyloidosis patients after high-dose melphalan: a long-term follow-up study. *Blood*. 2016;128(4):594-602.
9. Muchtar E, Dispenzieri A, Kumar SK, et al. Interphase fluorescence in situ hybridization in untreated AL amyloidosis has an independent prognostic impact by abnormality type and treatment category. *Leukemia*. 2017;31(7):1562-1569.
10. Oliva L, Orfanelli U, Resnati M, et al. The amyloidogenic light chain is a stressor that sensitizes plasma cells to proteasome inhibitor toxicity. *Blood*. 2017;129(15):2132-2142.
11. Dispenzieri A. The yin and yang of autophagy in AL amyloidosis. *Blood*. 2017;129(15):2044-2045.

12. Muchtar E, Gertz MA, Kumar SK, et al. Improved outcomes for newly diagnosed AL amyloidosis between 2000 and 2014: cracking the glass ceiling of early death. *Blood*. 2017;129(15):2111-2119.
13. Reece DE. Breaking bad...proteins. *Blood*. 2017;129(15):2041-2042.
14. Palladini G, Dispenzieri A, Gertz MA, et al. New criteria for response to treatment in immunoglobulin light chain amyloidosis based on free light chain measurement and cardiac biomarkers: impact on survival outcomes. *J Clin Oncol*. 2012;30(36):4541-4549.
15. Palladini G, Hegenbart U, Milani P, et al. A staging system for renal outcome and early markers of renal response to chemotherapy in AL amyloidosis. *Blood*. 2014;124(15):2325-2332.
16. Merlini G, Lousada I, Ando Y, et al. Rationale, application and clinical qualification for NT-proBNP as a surrogate end point in pivotal clinical trials in patients with AL amyloidosis. *Leukemia*. 2016;30(10):1979-1986.
17. Sanchorawala V, Sun F, Quillen K, Sloan JM, Berk JL, Seldin DC. Long-term outcome of patients with AL amyloidosis treated with high-dose melphalan and stem cell transplantation: 20-year experience. *Blood*. 2015;126(20):2345-2347.
18. Palladini G, Sachchithanatham S, Milani P, et al. A European collaborative study of cyclophosphamide, bortezomib, and dexamethasone in upfront treatment of systemic AL amyloidosis. *Blood*. 2015;126(5):612-615.
19. Cibeira MT, Bladé J. Upfront CyBorD in AL amyloidosis. *Blood*. 2015;126(5):564-566.
20. Dispenzieri A, Buadi F, Laumann K, et al. Activity of pomalidomide in patients with immunoglobulin light-chain amyloidosis. *Blood*. 2012;119(23):5397-5404.
21. Sanchorawala V, Shelton AC, Lo S, Varga C, Sloan JM, Seldin DC. Pomalidomide and dexamethasone in the treatment of AL amyloidosis: results of a phase 1 and 2 trial. *Blood*. 2016;128(8):1059-1062.
22. Palladini G, Milani P, Foli A, et al. A phase 2 trial of pomalidomide and dexamethasone rescue treatment in patients with AL amyloidosis. *Blood*. 2017;129(15):2120-2123.
23. Sher T, Fenton B, Akhtar A, Gertz MA. First report of safety and efficacy of daratumumab in 2 cases of advanced immunoglobulin light chain amyloidosis. *Blood*. 2016;128(15):1987-1989.

24. Kaufman GP, Schrier SL, Lafayette RA, Arai S, Witteles RM, Liedtke M. Daratumumab yields rapid and deep hematologic responses in patients with heavily pretreated AL amyloidosis. *Blood*. 2017;130(5):900-902.
25. Sanchorawala V, Palladini G, Kukreti V, et al. A phase 1/2 study of the oral proteasome inhibitor ixazomib in relapsed or refractory AL amyloidosis. *Blood*. 2017;130(5):597-605.
26. Weiss BM, Wong SW, Comenzo RL. Beyond the plasma cell: emerging therapies for immunoglobulin light chain amyloidosis. *Blood*. 2016;127(19):2275-2280.
27. Warsame R, Bang SM, Kumar SK, et al. Outcomes and treatments of patients with immunoglobulin light chain amyloidosis who progress or relapse postautologous stem cell transplant. *Eur J Haematol*. 2014;92(6):485-490.
28. Browning S, Quillen K, Sloan JM, Doros G, Sarosiek S, Sanchorawala V. Hematologic relapse in AL amyloidosis after high-dose melphalan and stem cell transplantation. *Blood*. 2017;130(11):1383-1386.
29. Hwa YL, Warsame R, Gertz MA, et al. Delineation of the timing of second-line therapy post-autologous stem cell transplant in patients with AL amyloidosis. *Blood*. 2017;130(13):1578-1584.
30. Dispenzieri A, Seenithamby K, Lacy MQ, et al. Patients with immunoglobulin light chain amyloidosis undergoing autologous stem cell transplantation have superior outcomes compared with patients with multiple myeloma: a retrospective review from a tertiary referral center. *Bone Marrow Transplant*. 2013;48(10):1302-1307.
31. Milani P, Gertz MA, Merlini G, Dispenzieri A. Attitudes about when and how to treat patients with AL amyloidosis: an international survey. *Amyloid*. 2017:1-4.
32. Merlini G, Wechalekar AD, Palladini G. Systemic light chain amyloidosis: an update for treating physicians. *Blood*. 2013;121(26):5124-5130.
33. Dispenzieri A, Buadi F, Kumar SK, et al. Treatment of Immunoglobulin Light Chain Amyloidosis: Mayo Stratification of Myeloma and Risk-Adapted Therapy (mSMART) Consensus Statement. *Mayo Clin Proc*. 2015;90(8):1054-1081.
34. Wechalekar AD, Gillmore JD, Bird J, et al. Guidelines on the management of AL amyloidosis. *Br J Haematol*. 2015;168(2):186-206.
35. Fernández de Larrea C, Verga L, Morbini P, et al. A practical approach to the diagnosis of systemic amyloidoses. *Blood*. 2015;125(14):2239-2244.

36. Brambilla F, Lavatelli F, Di Silvestre D, et al. Shotgun protein profile of human adipose tissue and its changes in relation to systemic amyloidoses. *J Proteome Res.* 2013;12(12):5642-5655.
37. Dispenzieri A, Gertz M, Kyle R, et al. Serum cardiac troponins and N-terminal pro-brain natriuretic peptide: a staging system for primary systemic amyloidosis. *J Clin Oncol.* 2004;22(18):3751-3757.
38. Wechalekar AD, Schonland SO, Kastiris E, et al. A European collaborative study of treatment outcomes in 346 patients with cardiac stage III AL amyloidosis. *Blood.* 2013;121(17):3420-3427.
39. Milani P, Basset M, Russo F, Foli A, Merlini G, Palladini G. Patients with light-chain amyloidosis and low free light-chain burden have distinct clinical features and outcome. *Blood.* 2017;130(5):625-631.
40. Dittrich T, Bochtler T, Kimmich C, et al. AL amyloidosis patients with low amyloidogenic free light chain levels at first diagnosis have an excellent prognosis. *Blood.* 2017;130(5):632-642.
41. Tandon N, Sidana S, Gertz MA, et al. Treatment Patterns and Outcome Following Initial Relapse or Refractory Disease in Patients with Systemic Light Chain Amyloidosis. *Am J Hematol.* 2017;92(6):549-554.
42. Palladini G, Russo P, Foli A, et al. Salvage therapy with lenalidomide and dexamethasone in patients with advanced AL amyloidosis refractory to melphalan, bortezomib, and thalidomide. *Ann Hematol.* 2012;91(1):89-92.
43. Muchtar E, Dispenzieri A, Kumar SK, et al. Immunoparesis status in immunoglobulin light chain amyloidosis at diagnosis affects response and survival by regimen type. *Haematologica.* 2016;101(9):1102-1109.
44. Muchtar E, Dispenzieri A, Kumar SK, et al. Interphase fluorescence in situ hybridization in untreated AL amyloidosis has an independent prognostic impact by abnormality type and treatment category. *Leukemia.* 2017;31(7):1562-1569.
45. Palladini G, Massa M, Basset M, et al. Persistence of Minimal Residual Disease By Multiparameter Flow Cytometry Can Hinder Recovery of Organ Damage in Patients with AL Amyloidosis Otherwise in Complete Response. *Blood.* 2016;128(22).
46. Barnidge DR, Dispenzieri A, Merlini G, Katzmann JA, Murray DL. Monitoring free light chains in serum using mass spectrometry. *Clin Chem Lab Med.* 2016;54(6):1073-1083.

47. Milani P, Murray DL, Barnidge DR, et al. The utility of MASS-FIX to detect and monitor monoclonal proteins in the clinic. *Am J Hematol.* 2017;92(8):772-779.
48. Russo F, Valentini V, Basset M, et al. Identification and quantification of urinary monoclonal proteins by capillary electrophoresis in AL amyloidosis. *Amyloid.* 2017;24(sup1):66-67.

Table 1. Levels of dFLC and markers of organ involvement in patients who required second line therapy

Variables	Baseline median (IQR)	Best response median (IQR)	Second-line therapy initiation median (IQR)
dFLC, mg/L (92 patients)	140 (59-324)	12 (5-31)	55 (26-108)
NT-proBNP, ng/L (56 patients with heart involvement)	2913 (1329-5577)	1419 (724-4107)	1602 (958-5443)
Proteinuria, g/24h (61 patients with renal involvement)	5.2 (2.5-7.5)	2.7 (1.3-4.9)	2.4 (0.7-4.5)
eGFR, mL/min per 1.73 m ² (61 patients with renal involvement)	72 (49-87)	79 (42->90)	46 (32-75)

dFLC, difference between involved (amyloidogenic) and uninvolved free light chains; eGFR, estimated glomerular filtration rate; IQR, interquartile range; NT-proBNP, N-terminal pro-natriuretic peptide type-B.

Baseline values were measured at the time of diagnosis. Best response was achieved after a median of 5.8 months from upfront therapy initiation.

Table 2. Cox univariate and multivariate analyses of baseline variables predicting treatment-free survival

Univariate analysis		
Variables	HR (95% CI)	P
iuFLCR >6 at diagnosis	1.69 (1.06-2.69)	0.017
BMDex as upfront therapy	0.36 (0.13-0.82)	0.009
CR after upfront therapy	0.36 (0.18-0.64)	<0.001
Age, years	1.00 (0.99-1.02)	0.877
BMPC, %	1.01 (0.99-1.04)	0.404
BMPC >10%	1.13 (0.67-1.92)	0.644
dFLC >50 mg/L	1.44 (0.87-2.37)	0.160
dFLC >180 mg/L	1.04 (0.69-1.59)	0.835
Heart involvement	0.94 (0.61-1.45)	0.770
Cardiac stage III	0.85 (0.50-1.47)	0.570
Renal stage III	0.70 (0.30-1.60)	0.398
Multivariate analysis		
Variables	HR (95% CI)	P
iuFLCR >6 at diagnosis	1.41 (0.87-2.27)	0.161
BMDex as upfront therapy	0.40 (0.20-0.81)	0.011
CR after upfront therapy	0.40 (0.24-0.65)	<0.001

BMDex, bortezomib, melphalan, and dexamethasone; BMPC, bone marrow plasma cell infiltrate; CR, complete response; dFLC, difference between involved and uninvolved free light chain; HR, hazard ratio; iuFLCR, involved / uninvolved free light chain ratio.

Cardiac stage III is defined by N-terminal pro-natriuretic peptide type-B >332 ng/L and cardiac troponin I >0.1 ng/L.

Renal stage III is defined by estimated glomerular filtration rate <50 mL/min per 1,73 m² and proteinuria >5 g/24h.

Figure 1. Increase of dFLC from the nadir reached after upfront therapy in 20 patients with cardiac progression

Cardiac progression is defined by an increase in NT-proBNP that is both >30% and >300 ng/L.

Black bars: >10% increase in dFLC

Grey bars: “high-risk dFLC progression”, defined as an increase in dFLC that is >20 mg/L, >20% of baseline value observed at diagnosis, and >50% of the value reached at best response.

dFLC, difference between involved and uninvolved free light chain; NT-proBNP, N-terminal pro-natriuretic peptide type-B.

Figure 2. Time to second line therapy in 259 patients with AL amyloidosis who achieved hematologic response after upfront therapy

Median 49 months

Figure 3. Variables affecting time to second line therapy in 259 patients with AL amyloidosis who achieved hematologic response after upfront therapy

A. Impact of involved / uninvolved free light chain ratio (iuFLCR) at baseline on time to second line therapy.

Solid line, iuFLCR <6, 94 patients; dotted line iuFLCR ≥6, 165 patients. P=0.027.

B. Impact of upfront treatment type on time to second line therapy.

Solid line, bortezomib, melphalan and dexamethasone (BMDex, 46 patients); dashed line cyclophosphamide, bortezomib and dexamethasone (CyBorD, 71 patients, P=0.027 compared to BMDex); dotted line melphalan / dexamethasone (MDex, 129 patients, P=0.022 compared to BMDex).

C. Impact of quality of hematologic response after upfront therapy on time to second line therapy.

Solid line, complete response (CR, 82 patients, median 74 months); dashed line very good partial response (VGPR, 134 patients, median 44 months, P=0.006 compared to CR); dotted line partial response (PR, 43 patients, median 24 months, P=0.036 compared to VGPR). All the patients in PR had also achieved organ response.

Figure 4. Overall survival

- A. Survival from diagnosis of the whole cohort** (259 patients, median 99 months)
- B. Survival from the time of initiation of second-line therapy** (92 patients, median 59 months)

Figure 5. Variables affecting survival after second-line therapy initiation

- A. Impact of NT-proBNP progression on survival after second-line therapy initiation (P=0.002)**

Solid line, no NT-proBNP progression (40 patients, median survival 62 months), dotted line NT-proBNP progression (20 patients, median survival 17 months).

- B. Impact of dFLC progression on survival after second-line therapy initiation (P=0.004)**

Dotted line patients with “high-risk dFLC progression” (60 patients, median survival 46 months); all of the following are required to define “high-risk dFLC progression”:

- dFLC >20 mg/L,
- dFLC >20% of baseline value,
- dFLC increase by >50% of value reached at best response.

Solid line all other patients (32 patients, median survival not reached).

Figure 1.

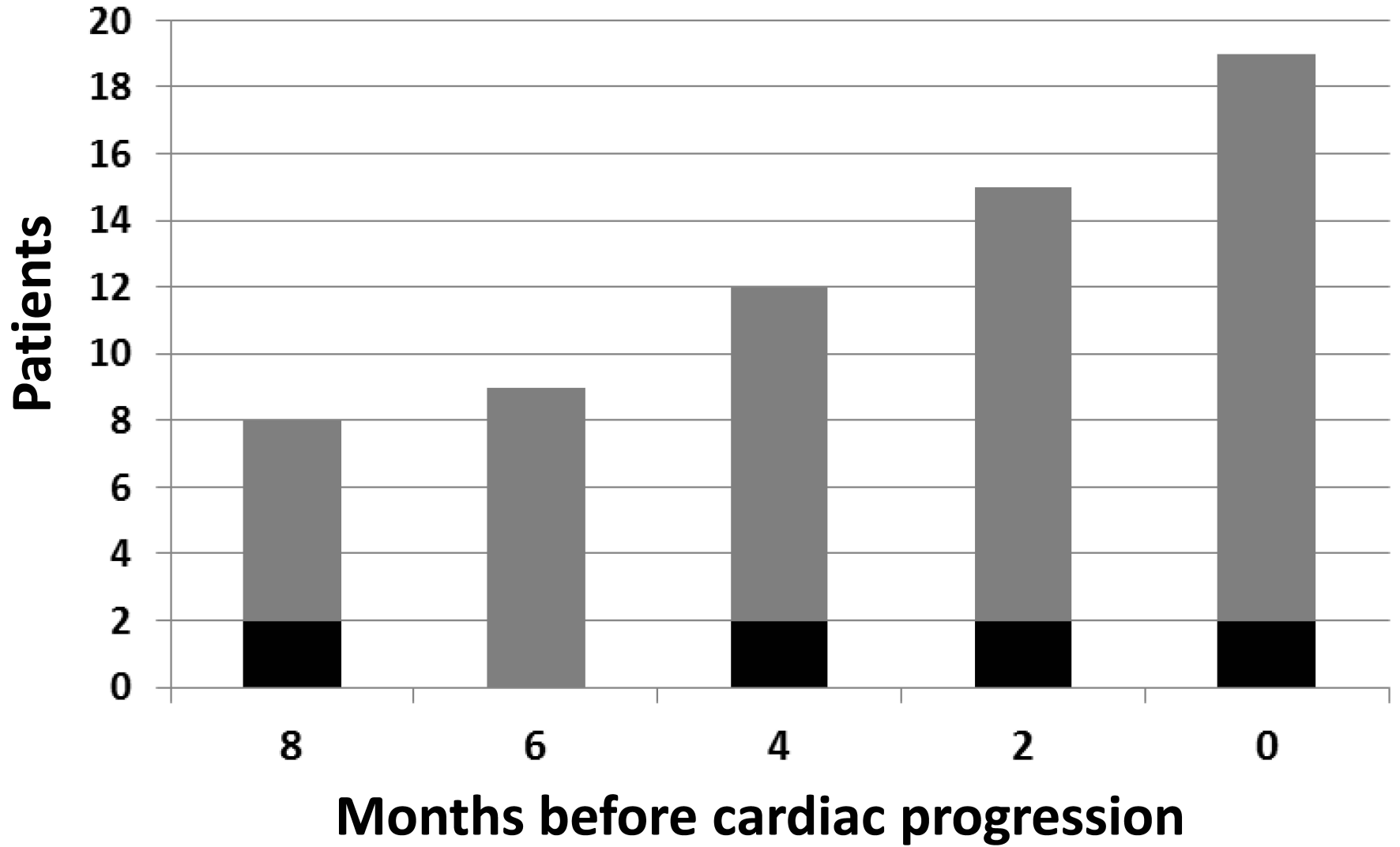


Figure 2.

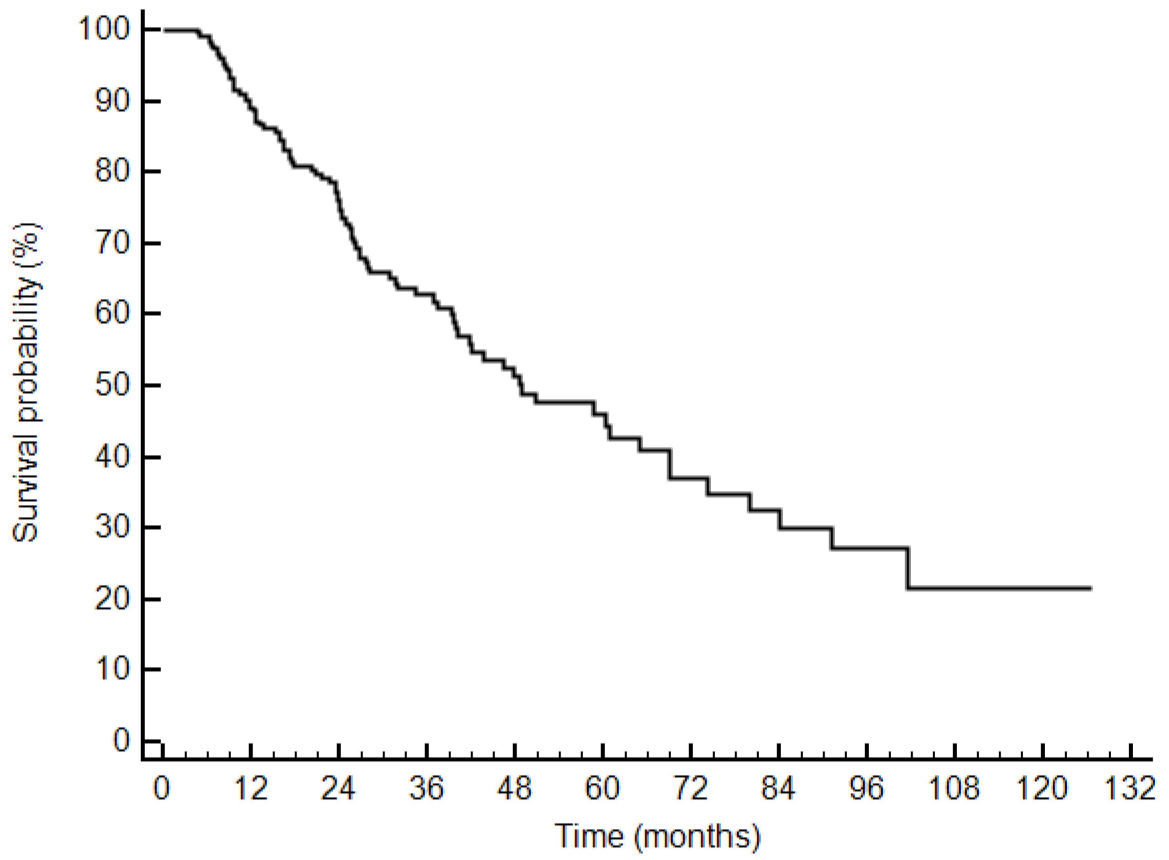


Figure 3.

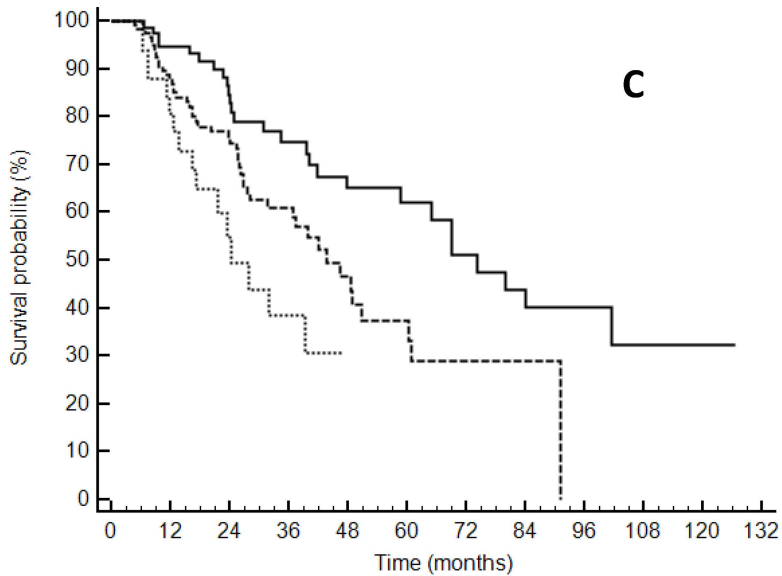
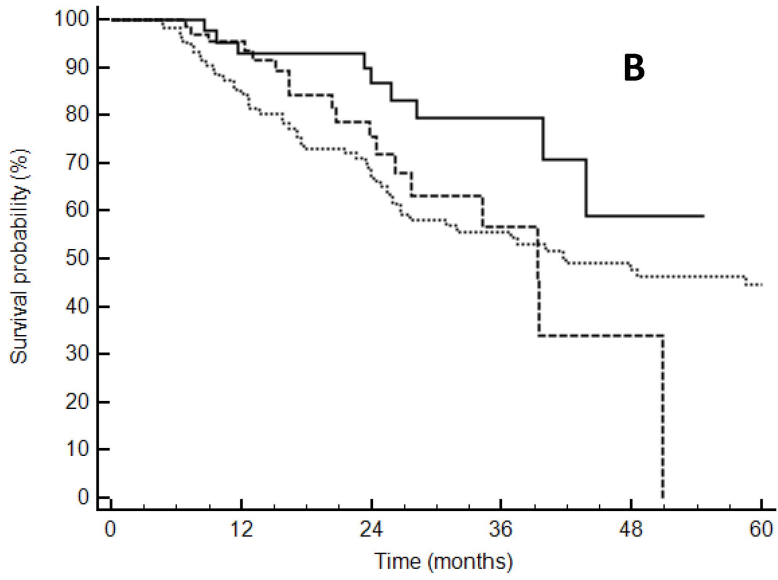
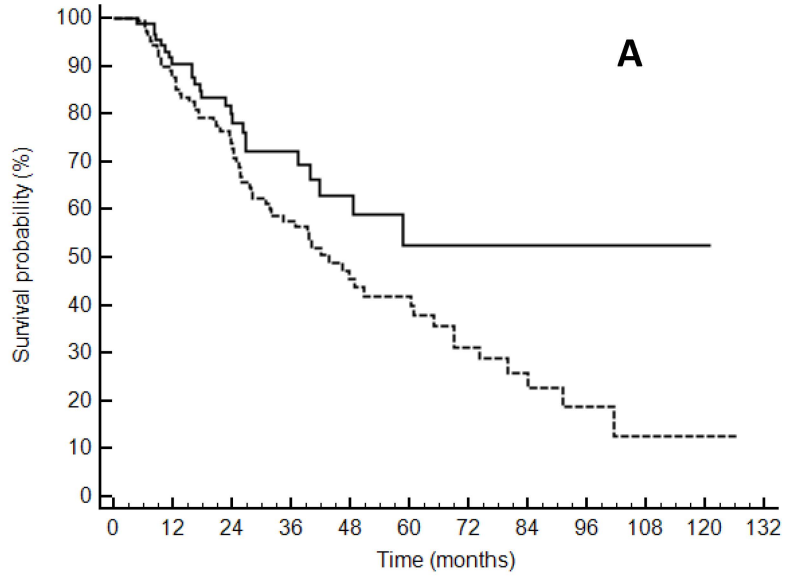


Figure 4.

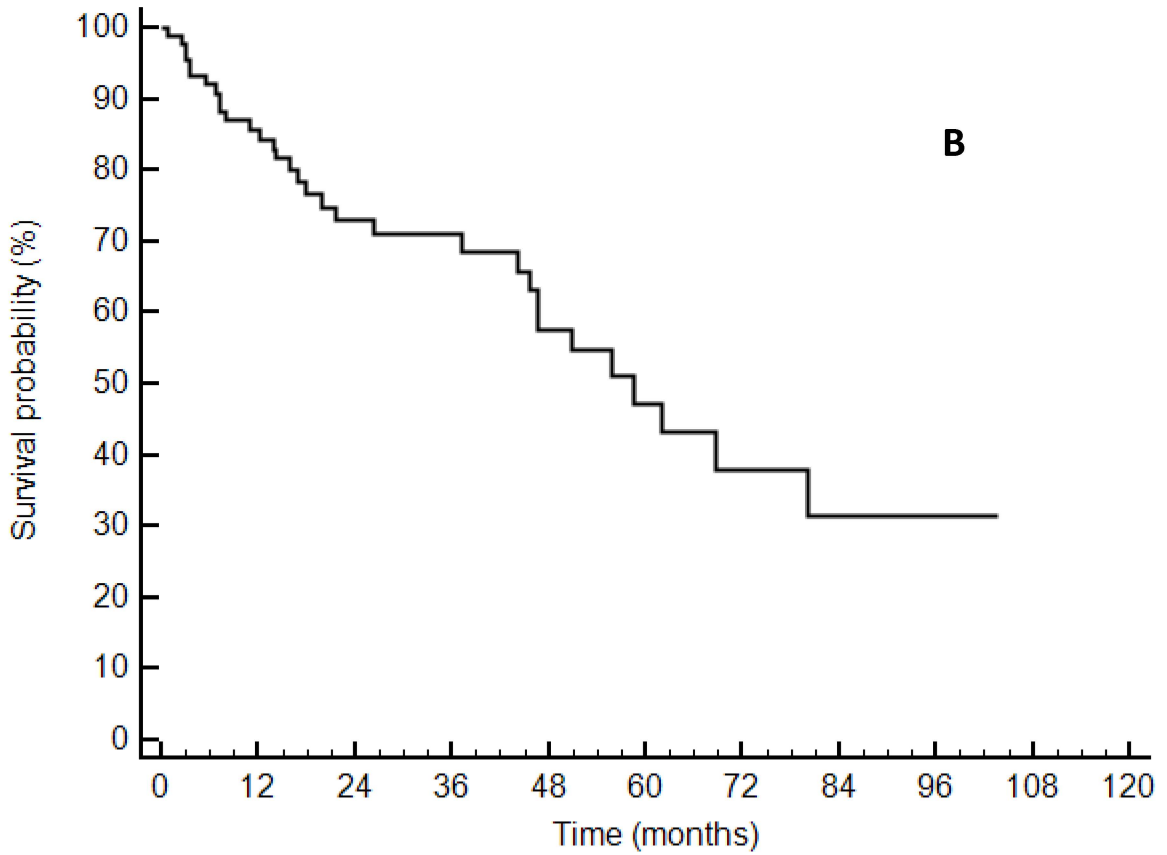
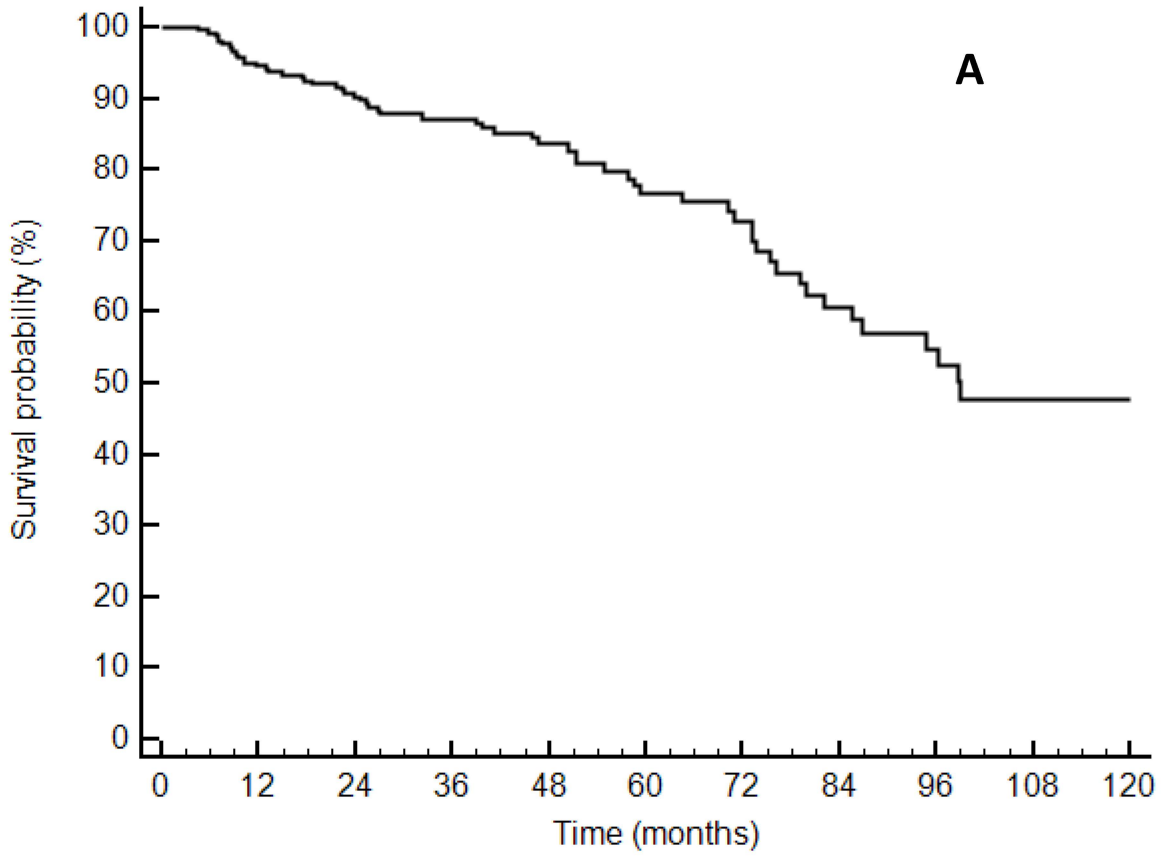


Figure 5.

