Microchimica Acta

Silica-supported carbon from Kraft lignin pyrolysis for solid-phase extraction of fluoroquinolone probes pollutants from environmental waters --Manuscript Draft--

Manuscript Number:					
Full Title:	Silica-supported carbon from Kraft lignin pyrolysis for solid-phase extraction of fluoroquinolone probes pollutants from environmental waters				
Article Type:	Original Paper				
Keywords:	Emerging pollutants; Fluoroquinolones; Lignin; Pyrolysis; Solid-phase extraction; Surface water				
Corresponding Author:	Andrea Speltini, Ph.D University of Pavia, Italy Pavia, PV ITALY				
Corresponding Author Secondary Information:					
Corresponding Author's Institution:	University of Pavia, Italy				
Corresponding Author's Secondary Institution:					
First Author:	Andrea Speltini, Ph.D				
First Author Secondary Information:					
Order of Authors:	Andrea Speltini, Ph.D				
	Michela Sturini				
	Federica Maraschi				
	Elettra Mandelli				
	Dhanalakshmi Vadivel				
	Daniele Dondi				
	Antonella Profumo, Prof				
Order of Authors Secondary Information:					
Funding Information:					
Abstract:	This work presents a novel carbon-based material obtained from a low cost biomass, and its original application for solid-phase extraction (SPE) of organic pollutants from natural waters. Marbofloxacin (MAR) and Enrofloxacin (ENR), two widely used fluoroquinolones (FQs), were chosen as model emerging contaminants. The material was easily prepared by adsorption of lignin (LG) onto silica microparticles followed by pyrolityc treatment at 1200°C for 2 h. Silica-supported lignin (LG-silica) was characterized by scanning electron microscopy (SEM), surface area measurement (BET), thermogravimetric analysis (TGA) and electron paramagnetic resonance (EPR). LG-silica (200mg) was tested as column-packed SPE sorbent for pre-concentration of tap and river water spiked with MAR and ENR in the range 10-1000 ng L-1 (n=3). Quantitative adsorption was observed at the sample native pH (~7.5) for both drugs, which were eluted from the cartridge with 2 x 2 mL aqueous tetrabutyl ammonium hydroxide-acetonitrile (70:30), and quantified by high performance liquid chromatography coupled to fluorescence detection. Recoveries in the range 70-116% and suitable inter-day precision (RSDs 7-16%) were observed for 20-1000 ng L-1 spikes, also in raw river water. Sample volumes up to 500 mL provided enrichment factors up to 125, useful for determination at environmentally significant levels (few ng L-1) and, at the same time, attractive compared to the commercial sorbents. The batch-to-batch reproducibility was verified on 3 LG-silica preparations. The novel sorbent phase was successfully applied to the determination of these drugs in actual environmental waters.				

1	Silica-supported carbon from Kraft lignin pyrolysis for solid-phase extraction of
2	fluoroquinolone probes pollutants from environmental waters
3	Andrea Speltini ^{a,*} , Michela Sturini ^a , Federica Maraschi ^a , Elettra Mandelli ^a , Dhanalakshmi Vadivel ^{a,b} , Daniele Dondi ^a ,
4	Antonella Profumo ^a
5	
6	^a Department of Chemistry, University of Pavia, via Taramelli 12, 27100 Pavia, Italy
7	^b Department of Chemistry, Bharathiar University, 641046 Coimbatore, India
8	*Corresponding author. Tel.: +39 0382-987349; fax: +39 0382-528544.
9	E-mail address: andrea.speltini@unipv.it
10	
11	
12	
13	
14	
15	
16	
17	
18	
19	
20	
21	
22	
23	
24	
25	
26	
27	
28	
29	

30 Abstract

This work presents a novel carbon-based material obtained from a low cost biomass, and its original application for solid-phase extraction (SPE) of organic pollutants from natural waters. Marbofloxacin (MAR) and Enrofloxacin (ENR), two widely used fluoroquinolones (FQs), were chosen as model emerging contaminants. The material was easily prepared by adsorption of lignin (LG) onto silica microparticles followed by pyrolityc treatment at 1200°C for 2 h. Silica-supported lignin (LG-silica) was characterized by scanning electron microscopy (SEM), surface area measurement (BET), thermogravimetric analysis (TGA) and electron paramagnetic resonance (EPR). LG-silica (200mg) was tested as column-packed SPE sorbent for pre-concentration of tap and river water spiked with MAR and ENR in the range 10-1000 ng L⁻¹ (n=3). Quantitative adsorption was observed at the sample native pH (~7.5) for both drugs, which were eluted from the cartridge with 2×2 mL aqueous tetrabutyl ammonium hydroxide-acetonitrile (70:30), and quantified by high performance liquid chromatography coupled to fluorescence detection. Recoveries in the range 70-116% and suitable inter-day precision (RSDs 7-16%) were observed for 20-1000 ng L^{-1} spikes, also in raw river water. Sample volumes up to 500 mL provided enrichment factors up to 125, useful for determination at environmentally significant levels (few ng L⁻¹) and, at the same time, attractive compared to the commercial sorbents. The batch-to-batch reproducibility was verified on 3 LG-silica preparations. The novel sorbent phase was successfully applied to the determination of these drugs in actual environmental waters.

47 Keywords : Emerging pollutants; Fluoroquinolones; Lignin; Pyrolysis; Solid-phase extraction; Surfa	flace water
--	-------------

60 Introduction

61 Reversed-phase materials have been commonly used in separation science for pre-concentration of a variety of organic compounds from aqueous solution [1]. Beside the conventional C18-silica, in the recent years novel materials have been 62 prepared and applied for analytical purposes, exploiting the exceptional properties of new forms of sp^2 carbon, such as 63 64 carbon nanotubes and more recently graphene [2-4]. The latter, made of single-layer or few-layer thickness of sp²-65 hybridized carbon atoms arranged in a honeycomb-like structure, showed great potentiality to prepare efficient sorbents 66 for solid-phase extraction (SPE) of pollutants [5, 6]. Differently from C18-silica, an aspecific sorbent merely based on 67 hydrophobic effects, graphene exerts several interplays with target species, viz. π - π stacking, cation- π bonding, electron 68 donor-acceptor and hydrophobic interactions, hydrogen bond, etc. [5]. Comparable or even better performance was 69 observed in comparison with carbon nanotubes, graphitic carbon, and commercial sorbents for pre-concentration of 70 various contaminants, e.g. sulfonamides [7] and fluoroquinolones (FQs) [8]. These, antibiotics for human and veterinary 71 medicine endowed with broad activity spectrum and good oral absorption, are important emerging contaminants. In the 72 recent years their occurrence has been assessed both in aquatic [9] and soil [10] compartments at concentrations in the ranges ng-µg L-1 and µg-mg kg-1, respectively. Various reasons stand for the environmental widespread, primarily the 73 74 partial metabolization and not quantitative abatement in wastewater treatment plants (WWTPs) [11]. This involves the 75 release of variable quantities of pharmaceutically active drugs into water basins, affecting transformation/removal rates. 76 The common practice of recycling manure from livestock farming and sewage sludge from WWTPs as fertilizers, and 77 recycling of sludge for the production of compost, widely employed as soil conditioner/fertilizer, are other remarkable 78 contamination routes. These biosolids can contain appreciable amounts of residual drugs [12, 13]. Despite FQs 79 photosensitivity, both in water [14, 15] and soil systems [16, 17], the presence of these antibiotics in the environment 80 implicates serious threats to the ecosystem and human health. Mainly, bacterial resistance stimulation [18, 19] and 81 formation of pharmacologically active photoproducts that contribute to the overall impact [19, 20]. For these reasons, 82 monitoring FQs in the environment is an important task, and needs accurate and sensitive analytical methods. The low 83 concentrations usually detected in natural waters (from tens ng L^{-1} to few $\mu g L^{-1}$) require a SPE pre-concentration step. 84 In this work we prepared a novel SPE sorbent from pyrolysis of Kraft lignin (LG) onto silica. LG was chosen as the 85 carbon precursor due to its base aromatic structure, large availability and low cost. LG is indeed one of the three main 86 constituents of biomass together with cellulose and hemicelluloses, and also the major waste of the pulping process of 87 the paper industry, with increasing amounts from the emerging cellulosic ethanol industry [21, 22]. As apparent from

the new insights in LG pyrolysis [23, 24], growing attention is being paid to its conversion into renewable energy

sources by pyrolytic treatment, that yields valuable byproducts, *viz.* char, bio-oil, syngas, and fine chemicals such as
oxygenated aromatic compounds [25].

91 In particular, Kraft LG is obtained as a byproduct of the paper pulping process. In the Kraft process the insoluble LG 92 polymer present in wood is decomposed into smaller units under high temperatures (170-180°C) in the presence of 93 strong alkali solutions and sulfides. During the process the LG polymer backbone is broken and sulfides are included in 94 the structure, making it water-soluble.

95 LG pyrolysis, i.e. the treatment at high temperature in oxygen-free atmosphere, leads to the elimination of sulfur and 96 oxygen present in the polymer, thus leaving carbon as a residue [23, 25]. Indeed, from the chemical viewpoint LG is 97 one of the most interesting natural polymers for production of graphenes. In fact, LG backbone consists of 98 hydroxyphenylpropanoic units linked through ether and C-C condensed types of linkages, with p-coumaryl alcohol, 99 coniferyl alcohol, and sinapyl alcohol as the main monomers [25], that shows a polymerization degree of the order of 100 several hundred [26]. This complex base-structure has in principle great potentiality to be rearranged in extended 101 graphene-like aromatic layers, as demonstrated under microwave plasma irradiation [27] and also by pyrolitic treatment 102 [28]. It is documented that carbonaceous residues from LG pyrolysis contain appreciable amounts of polycyclic 103 aromatic hydrocarbons [25, 29], and that polymeric carbon with fine sorption properties can be obtained by pyrolysis of 104 olive stones LG biomass [26]. It is also reported that softwood pyrolysis yields graphene sheets rearranged in 105 turbostratic carbon [28], and recently it was seen that pyrolysis of LG deposited on SiO₂/silicon substrates produces 106 graphene-like carbons [30]. Furthermore, it is well known that pyrolysis of Kraft LG proceeds in two steps according to 107 the pyrolysis temperature. Between 200 and 400°C CO₂ is produced together with volatile sulfur compounds [23, 31], 108 and in the range 500-600°C CO is evolved from phenols decomposition. H₂ production begins at 500°C [32, 33], 109 indicating that, starting from this temperature, carbon defects are cured. In fact, bulk pyrolysis of LG at high 110 temperature (> 3000 K) is known to yield high-ordered carbon [34].

111 Herein we report the preparation of a carbon-based material consisting of silica-supported pyrolyzed LG, to be used as a 112 novel SPE sorbent. The preparation of the material, simple and costless, consists of two steps only, viz. adsorption of aqueous LG onto micrometric silica followed by pyrolytic treatment to convert lignin into sp² carbon phase. LG-silica 113 114 has been characterized by thermogravimetric analysis (TGA), scanning electron microscopy (SEM), surface area BET 115 measurements and electron paramagnetic resonance (EPR), and tested as SPE sorbent for pre-concentration of 116 Marbofloxacin (MAR) and Enrofloxacin (ENR) from tap and not tampered river water. These two FQ drugs, widely 117 diffused in surface waters of Northern Italy due to the large use for veterinary purposes [35], were chosen as probe 118 contaminants to test this novel sorbent as extractant. After SPE on LG-silica, separation and quantification were

- 119 performed by high performance liquid chromatography coupled with fluorescence detection (HPLC-FD). The batch-to-
- batch reproducibility was assessed, the main analytical aspects were evaluated and, finally, the sorbent phase was

applied to the determination of FQs in actual environmental waters.

122

123 Experimental

- 124 Chemicals
- 125 All chemicals were reagent grade or higher in quality (the complete list is in Supplementary material).
- 126

127 Preparation and characterization of LG-silica

LG-silica was prepared by adding 1 g silica and known amounts (20-50-100 mg, nominal loadings 2-5-10 wt%) of LG in 50 mL distilled water. The suspension was mixed to promote LG adsorption onto silica microparticles and then water was removed under vacuum overnight. For the pyrolysis (high-temperature annealing under vacuum), 200 mg sample was put in an alumina combustion boat, located inside a quartz tube inserted into a cylindrical oven (Watlow ceramic fiber heater, i.d. 38 mm, length 305 mm, electrical power 600 W). A double stage diffusion and rotary vacuum pump was used to work in vacuum (10⁻³ Pa), mandatory to get anoxic atmosphere. The heating rate of the oven was 10°C min⁻ and the final temperature of 1200°C was kept for 20 min.

135 The final material was characterized by SEM, BET, TGA and EPR (experimental details in Supplementary material).

136

137 SPE procedure

138 The cartridge (6-mL polypropylene tubes) was prepared by slurry packing 200 mg LG-silica, dispersed in MeOH, 139 between two polyethylene frits. To remove impurities and minimize void/channeling effect the cartridge was washed 140 under vacuum (Resprep manifold, Restek Corporation, Bellefonte, USA) with 40 mL MeOH, 40 mL distilled water, 10 141 mL 50 mM TBAH-ACN (70:30) and finally 40 mL distilled water. Before the extraction procedure, performed by a 142 multi-channel peristaltic pump (Gilson, Italy), the cartridge was conditioned with 5 mL MeOH and 5 mL ultrapure 143 water; the water sample (50-500 mL) was fed to the column at a flow rate of ~ 3.5 mL min⁻¹, and then the cartridge was 144 dried for 5 min. The analytes were eluted under vacuum with 2×2 mL ACN-50 mM TBAH (30:70), at a flow rate of 1 145 mL min⁻¹. The extract was acidified with 1 M HCl ($pH \sim 2.5$) before HPLC injection. Details about water samples and 146 HPLC are in Supplementary material.

147

148 Analytical evaluation

149 Since the aim was applying LG-silica to real samples, the analytical evaluation of the whole procedure was carried out.

As no CRMs are available, trueness was evaluated by recovery tests [36, 37], on tap water fortified with different

amounts of FQs, in the range 20-1000 ng L^{-1} (*n*=3), and on blank river water – previously analyzed by a published

152 procedure [35] – spiked at 10-100 ng L⁻¹ (*n*=3).

- 153 The intra-day precision (repeatability) was evaluated on independently fortified tap water samples (50 ng L^{-1} , n=3). The
- inter-day precision was assessed on tap water (20-1000 ng L⁻¹, n=3) and also on river water spiked at 20-100 ng L⁻¹ (n=3).
- 156 The batch-to-batch reproducibility was studied by recovery tests on tap water (50 mL) spiked with 5 μ g L⁻¹ of MAR and 157 ENR, using three LG-silica samples independently synthesized following the entire preparation.
- Selectivity was evaluated from the HPLC-FD chromatograms of the SPE extracts obtained by pre-concentration of nottampered blank river water [38].
- 160 Linearity was determined by three independent six-point calibration curves generated for each analyte in the range 1-10
- 161 $\mu g L^{-1}$ using the weighted linear least squares (wLLSR) regression.
- Theoretical method detection and quantification limits (MDLs, MQLs) were obtained on the basis of the instrumental detection and quantification limits (IDLs, IQLs) calculated using the residual standard deviation (Sy/x) of the wLLSR regression as $(3 \times Sy/x)/s$ lope and $(10 \times Sy/x)/s$ lope, respectively. Experimental method detection and quantification limits (MDL, MQL) were determined as the minimum concentration that provided a FD response significantly different from zero, and the lowest concentration that provided acceptable recovery (\geq 70%) and precision (RSD<20%), respectively [38].
- 168

169 Results and Discussion

170 Characterization of LG-silica

171 Fig. 1 shows the SEM images of pristine silica (a), LG-free silica after pyrolysis (b) and 2wt% LG@silica after 172 pyrolysis (c), together with the related pictures (a1,b1,c1). LG-silica appears as a blackish powder (c1), while the other 173 two samples appear white colored (a1,b1). SEM characterization indicates that in the pristine material (a) the particles 174 surface is smoother and the particle size is slightly bigger compared to the treated samples (b,c). Apart from these little 175 differences, no substantial changes in shape and dimension occurred after thermal treatment (b). Moreover, no 176 significant features can be found in LG-silica (c) compared to the samples not containing LG (a,b), reasonably due to 177 the small amount of immobilized carbonaceous material. Indeed, results from TGA showed that silica was effectively 178 derivatized with pyrolytic carbon, 0.72, 1.12 and 1.23wt% for the samples prepared with nominal LG loadings of 2, 5 and 10wt%, respectively. The residual carbon content resulted lower with respect to the initial loading due to the weight
loss occurring during pyrolysis (loss of oxygenated and sulfur moieties present in the Kraft lignin [23]).

BET analysis indicated surface area values of $370 \text{ m}^2 \text{ g}^{-1}$ and $430 \text{ m}^2 \text{ g}^{-1}$ for LG-silica and for the *blank* silica submitted to the same pyrolytic treatment, respectively. Compared to pristine silica (450 m² g⁻¹), the value found for LG-silica accounted for the silica surface modification induced by the deposition of LG-derived carbons.

184 The LG-silica samples were analyzed by EPR spectroscopy at ambient temperature (see Supplementary material). It is 185 evident that the spectrum shape is as function of pyrolysis temperature. The sample pyrolized at lower temperature (Fig. 186 S1) showed an asymmetric peak due to the presence of both carbon and oxygen centred radicals. Moreover, the 187 spectrum linewidth decreased at higher temperature due to the curing of the material with the formation of a less 188 defective graphitized carbon. Another information deriving from the EPR measurement is the estimation of the 189 extension of the delocalization of the free electrons on carbon. In fact, by increasing the applied microwave power, the 190 EPR active sites can undergo to a saturation due to the relaxation mechanism. More effective is the relaxation (thus 191 more extended is the delocalization) the lower is the saturation. From the analysis of the curves (Fig. S2) appears how 192 an higher temperature gives a less defective carbon (the number of spins per gram is lower), but with a formation of 193 smaller domains (increased deviation from the linearity at high power).

194

195 Solid-phase extraction on LG-silica

196 Preliminary tests were performed to investigate the affinity of the two drugs for LG-silica. The amount of material (200 197 mg) was chosen basing on the most used SPE sorbents commercially available for enrichment of trace contaminants 198 from surface waters, FQs included [9]. The first recovery tests were carried out on tap water (50 mL) spiked with 5 µg 199 L^{-1} of each FQ (*n*=3), working at the native sample pH (7.7±0.1). The percolated samples after SPE were analyzed by 200 HPLC-FD to verify the antibiotics adsorption. MAR and ENR were quantitatively adsorbed on the 2-5wt% LG-silica 201 $(MAR \le IDL \text{ and } ENR < IQL \text{ in the percolated samples});$ in the case of 10wt% LG-silica MAR concentration was < 202 IQL, while just 8% of the ENR initial amount was not adsorbed (for IDL and IQL see Analytical evaluation). These 203 findings attest the good affinity of the zwitterionic form of the two drugs for LG-silica, accordingly to their dissociation 204 constants (MAR pK_a1 5.51-5.69, pK_a2 8.02-8.58; ENR pK_a1 6.27, pK_a2 8.3 [9]), and in agreement with the results 205 previously obtained on graphene-derivatized silica [8]. Therefore no pH adjustment before SPE was necessary for 206 molecular adsorption.

The role of LG-derived pyrolytic carbon in the adsorption of the two drugs was proved by testing silica *not* containing
 LG (*blank* control sample) that was submitted to the same pyrolytic process. It was verified that the bare material does

not show any adsorption affinity for FQs, clearly highlighting the key role of LG immobilized onto the silica support bypyrolysis, even at low wt%.

211 Considering the preliminary findings and TGA results, it seemed convenient to select 2wt% LG-silica (real carbon 212 content 0.72wt%), that was then tested for further extractions (50 mL tap water, native pH, spike 5 μ g L⁻¹) to investigate 213 the elution step. Basing on previous papers [8, 9] and on the material characteristics (i.e. type and amount of carbon), 214 25mM H₃PO₄-ACN (85:15), 1% NH₃-MeOH (70:30) and 50 mM TBAH-ACN (70:30) were selected as the potential 215 eluting solutions. After pre-concentration, the analytes were eluted by 2×2 mL of the first solution obtaining very poor 216 recovery (<2%), while the second one allowed greater but not quantitative desorption (65-70% for MAR, 42-63% for 217 ENR); using 2×2 mL 50 mM TBAH-ACN (70:30) the recovery was 79(3)% and 75(1)% for MAR and ENR, 218 respectively.

The batch-to-batch reproducibility was assessed by recovery tests on three batches of LG-silica (nominal 2wt%) independently prepared starting from the pristine microparticles; SPEs of 50 mL tap water samples spiked with 5 μ g L⁻¹ gave RSD values of 8% and 7% for MAR and ENR, respectively, using 50 mM TBAH-ACN (70:30).

222 This solution was then tested for recovery tests performed at lower concentrations, from 20 to 1000 ng L^{-1} (sample 223 volumes up to 500 mL). Results showed that analytes were quantitatively eluted by 2×2 mL, obtaining satisfactory 224 recovery and precision (Table 1), and at the same time pre-concentration factors up to 125. These results obtained on 225 LG-derived carbon supported on silica (200 mg, measured carbon 0.72wt%) were really satisfactory compared to those 226 obtained using 200 mg of bulk HLB polymer [35, 39]. In view of the good performance of LG-silica, we tested its 227 extraction ability in a more complex matrix, that is not tampered river water. Recovery trials were undertaken at 228 concentrations in the range 10-100 ng L⁻¹, amounts commonly detected in natural-contaminated surface waters [9, 35]. 229 As shown in Table 1, recovery was quantitative for concentrations between 20 and 100 ng L^{-1} , with values higher than 230 70% and RSDs below 14%. This indicates that LG-silica is a suitable sorbent for SPE of FQs in actual samples, and a 231 typical chromatogram is shown in Fig. 2. At the lowest spike (10 ng L⁻¹) recovery was anyway semiquantitative (see 232 Table 1).

From the results, it appears that derivatization of silica with LG-derived pyrolytic carbon is a valid new route to obtain a composite SPE material with good adsorption properties. This is more clearly evident considering that the amount of carbon in the final material is around 1 wt%. Therefore, 200 mg LG-silica containing ca. 2 mg pyrolytic carbon allowed processing sample volumes up to 500 mL, providing pre-concentration factors suitable for environmental determination at the low ng L⁻¹. These experimental evidences make this novel material certainly appealing compared to the commercial bulk HLB polymer, but also to other silica-supported carbons such as C18-silica [35] and graphenederivatized silica [8]. The results here presented give new evidences about the great potentiality of low-cost materials
(i.e. waste lignin biomass) for analytical application.

241

242 Analytical evaluation

243 Before application of LG-silica to real samples, the main analytical aspects of the entire procedure were evaluated, as 244 hereafter summarized. Due to the absence of CRMs, trueness was evaluated by recovery tests. All the mean recoveries 245 (%) obtained in water samples spiked at environmentally significant concentrations (10-1000 ng L^{-1}) are gathered in 246 Table 1. Recovery was calculated as the ratio between the concentration determined in the SPE extract and that 247 expected after pre-concentration on LG-silica, calculated considering the initial amount of analyte and the enrichment factor. As apparent, recovery was generally between 71% and 105% in tap water; quantitative extraction, with 248 249 recoveries higher than 70% (n=3), was also obtained in untreated river water spiked with 20-100 ng L⁻¹, and around 250 55% for the lowest concentration of 10 ng L^{-1} . Precision was evaluated by calculating the RSDs associated to the mean 251 recovery obtained for each concentration (see Table 1). The intra-day precision (repeatability), calculated for 50 ng L^{-1} 252 spike, showed RSDs <8% (*n*=3). As shown in Table 1, the inter-day precision showed RSDs <15% (*n*=3) in tap water, 253 for spikes in the range 10-1000 ng L^{-1} ; suitable reproducibility was also observed in untreated river water spiked with 254 10-100 ng L⁻¹ FQs, obtaining RSDs lower than 16% (n=3).

Selectivity was checked from the chromatograms of blank water extracts. Fig. 2 shows the profile obtained for a blank river water SPE extract and the one obtained from the same river water sample spiked with 50 ng L⁻¹ of each FQ (enrichment factor 125). No peaks were noticed at the retention times of the two analytes, thus interference of matrix substances accountable for false-positive signals can be excluded.

259 No matrix interference was observed at the FD [17] from the analysis of FQs standard solutions prepared in the SPE 260 eluting solution and in the SPE extracts from pre-concentration of blank river water (matrix-matched calibration), in 261 accordance with previous work [8]. The wLLSR regression equations obtained in 50 mM TBAH-ACN (70:30), mean of 262 three independent calibration lines, showed good linearity for both drugs, as reported in Table 2 together with 263 instrumental and method sensitivity. Theoretical MDL and MQL were in the range of few ng L⁻¹, while the MDL and MQL in raw river water experimentally determined were respectively 8 ng L⁻¹ and 20 ng L⁻¹, for both compounds, 264 265 thanks to the enrichment factor gained using LG-silica. These values are comparable with those reported for FQs in 266 river water by SPE on HLB followed by UPLC-MS [40]. Considering the typical FQs concentrations in surface waters

267 [8, 9], the method sensitivity is suitable for quantitative determination in actual samples, but also for qualitative 268 "sensing" at the low ng L^{-1} levels.

269

270 *Application of LG-silica to actual samples*

271 LG-silica was finally applied for the determination of MAR and ENR in ditch waters collected in the Pavia county 272 downstream pigs and cattle farms regularly using these veterinary drugs. Variable amounts of antibiotics were 273 determined (confirmed also by duplicate spike on the SPE extract), ranging from few to some tens ng L⁻¹. In particular, 274 the highest concentrations of FQs were found in Ditch-1 sampled directly in the farm effluents, wherein 72 ng L⁻¹ of 275 MAR and 76 ng L⁻¹ of ENR were found (RSDs<7%, *n*=3). The chromatographic profile obtained for Ditch-1 after SPE 276 on LG-silica is shown in Fig. 3. FQs were detected also in Ditch-2, although their concentrations were below MQLs 277 (chromatogram not shown).

278 These findings evidence that livestock farms are major source for surface water contamination, further supporting 279 previous data [8, 35], and call for more care in controlling the diffusion of such emerging pollutants in rural areas 280 devoted to livestock breeding.

281

282 Conclusions

283 This is the first-time reporting on the application of pyrolytic carbon from LG as SPE sorbent phase. A novel sp² 284 carbonaceous material obtained from a low-cost biomass and supported on silica microparticles has been prepared, 285 characterized and tested as SPE cartridge for pre-concentration of FQ antibiotics from water, chosen as probes organic 286 contaminants to test the performance of the material. LG-silica, easily prepared in laboratory by pyrolysis of silica-287 adsorbed LG, was packed in conventional SPE columns and proved to be useful for analytical application. In fact, pre-288 concentration of sample volumes up to 500 mL provided enrichment factors up to 125, suitable for determination at 289 nanograms per liter levels. Quantitative recovery and suitable reproducibility were observed both in tap and untreated 290 river water. LG-silica was successfully employed for the analysis of actual surface water samples.

In conclusion, the preparation of this material is environmentally benign, simple, inexpensive and LG-silica is efficient
 for pre-concentration of trace contaminants from aqueous complex matrices.

293

294 Acknowledgements

295 The Authors want to thank Prof. Chiara Milanese and Dr. Alberto Zeffiro (Department of Chemistry, University of296 Pavia) for their support in material characterization.

297	The Authors acknowledge the European Union for an ERASMUS MUNDUS Action 2 scholarship through the
298	PANACEA project Agreement Number 2012 - 2647 / 001 - 001 - EMA2 (European Commission) (D. Vadivel
299	fellowship).
300	
301	The Authors declare that they have no conflict of interest.
302	
303	References
304	1. Liška I (2000) Fifty years of solid-phase extraction in water analysis - historical development and overview. J
305	Chromatogr A 885:3–16.
306	
307	2. Speltini A, Merli D, Profumo A (2013) Analytical application of carbon nanotubes, fullerenes and nanodiamonds in
308	nanomaterials-based chromatographic stationary phases: A review. Anal Chim Acta 783:1-16.
309	
310	3. Speltini A, Maiocchi M, Cucca L, Merli D, Profumo A (2014) Solid-phase extraction of PFOA and PFOS from
311	surface waters on functionalized multiwalled carbon nanotubes followed by UPLC-ESI-MS. Anal Bioanal Chem
312	406:3657–3665.
313	
314	4. Socas-Rodríguez B, Herrera-Herrera AV, Asensio-Ramos M, Hernández-Borges J (2014) Recent applications of
315	carbon nanotube sorbents in analytical chemistry. J Chromatogr A 1357:110-146.
316	
317	5. Sitko R, Zawisza B, Malicka E (2013) Graphene as a new sorbent in analytical chemistry. Trends Anal Chem 51:33-
318	43.
319	
320	6. Pytlakowska K (2015) Dispersive micro solid-phase extraction of heavy metals as their complexes with 2-(5-bromo-
321	2-pyridylazo)-5-diethylaminophenol using graphene oxide nanoparticles. Microchim Acta. doi:10.1007/s00604-015-
322	1596-3
323	
324	7. Sun N, Han Y, Yan H, Song Y (2014) A self-assembly pipette tip graphene solid-phase extraction coupled with
325	liquid chromatography for the determination of three sulfonamides in environmental water. Anal Chim Acta 810:25-31.
326	

- Speltini A, Sturini M, Maraschi F, Consoli L, Zeffiro A, Profumo A (2015) Graphene-derivatized silica as an efficient solid-phase extraction sorbent for pre-concentration of fluoroquinolones from water followed by liquid-chromatography fluorescence detection. J Chromatogr A 1379:9–15.
 Speltini A, Sturini M, Maraschi F, Profumo A (2010) Fluoroquinolone antibiotics in environmental waters: sample preparation and determination. J Sep Sci 33:1115–1131.
- 334 10. Speltini A, Sturini M, Maraschi F, Profumo A, Albini A (2011) Analytical methods for the determination of
 335 Fluoroquinolones in solid environmental matrices. Trends Anal Chem 30:1337–1350.
- 336

11. Reemtsma T, Jekel M (2006) Organic Pollutant in the Water Cycle. Wiley-VCH, Weinheim.

338

12. Lillenberg M, Yurchenko S, Kipper K, Herodes K, Pihl V, Sepp K, Lõhmus R, Nei L (2009) Simultaneous
determination of fluoroquinolones, sulfonamides and tetracyclines in sewage sludge by pressurized liquid extraction
and liquid chromatography electrospray ionization-mass spectrometry. J Chromatogr A 1216:5949–5954.

342

343 13. Speltini A, Sturini M, Maraschi F, Viti S, Sbarbada D, Profumo A (2015) Fluoroquinolone residues in compost by
344 green enhanced microwave-assisted extraction followed by ultra performance liquid chromatography tandem mass
345 spectrometry. J Chromatogr A 1410:44–50.

346

347 14. Sturini M, Speltini A, Maraschi F, Profumo A, Pretali L, Fasani E, Albini A (2010) Photochemical degradation of
348 Marbofloxacin and Enrofloxacin in natural waters. Environ Sci Technol 44:4564–4569.

349

350 15. Babíc S, Periša M, Škoríc I (2013) Photolytic degradation of norfloxacin, enrofloxacin and ciprofloxacin in various
351 aqueous media. Chemosphere 91:1635–1642.

352

353 16. Sturini M, Speltini A, Maraschi F, Profumo A, Pretali L, Fasani E, Albini A (2012) Sunlight-induced degradation of
354 soil-adsorbed veterinary antimicrobials Marbofloxacin and Enrofloxacin. Chemosphere 86:130–137.

355

356	17. Speltini A, Sturini M, Maraschi F, Profumo A, Albini A (2012) Microwave-assisted extraction and determination of
357	enrofloxacin and danofloxacin photo-transformation products in soil. Anal Bioanal Chem 404:1565–1569.
358	
359	18. Kusari S, Prabhakaran D, Lamshöft M, Spiteller M (2009) In vitro residual anti-bacterial activity of difloxacin,
360	sarafloxacin and their photoproducts after photolysis in water. Environ Pollut 157:2722-2730.
361	
362	19. Sturini M, Speltini A, Maraschi F, Pretali L, Profumo A, Fasani E, Albini A, Migliavacca R, Nucleo E (2012)
363	Photodegradation of fluoroquinolones in surface water and antimicrobial activity of the photoproducts. Water Res
364	46:5575–5582.
365	
366	20. Sukul P, Lamshöft M, Kusari S, Zühlke S, Spiteller M (2009) Metabolism and excretion kinetics of ¹⁴ C-labeled and
367	non-labeled difloxacin in pigs after oral administration, and antimicrobial activity of manure containing difloxacin and
368	its metabolites. Environ Res 109:225–231.
369	
370	21. Azadi P, Inderwildi OR, Farnood R, King DA (2013) Liquid fuels, hydrogen and chemicals from lignin: a critical
371	review. Renewable Sustainable Energy Rev. 21:506–523.
372	
373	22. Cotana F, Cavalaglio G, Nicolini A, Gelosia M, Coccia V, Petrozzi A, Brinchi L (2014) Lignin as co-product of
374	second generation bioethanol production from ligno-cellulosic biomass. Energy Procedia 45:52-60.
375	
376	23. Dondi D, Zeffiro A, Speltini A, Tomasi C, Vadivel D, Buttafava A (2014) The role of inorganic sulfur compounds
377	in the pyrolysis of Kraft lignin. J Anal Appl Pyrol 107:53–58.
378	
379	24. Zhou S, Brown RC, Bai X (2015) The use of calcium hydroxide pretreatment to overcome agglomeration of
380	technical lignin during fast pyrolysis. Green Chem 17:4748-4759.
381	
382	25. Zhou H, Wu C, Onwudili JA, Meng A, Zhang Y, Williams PT (2014) Polycyclic aromatic hydrocarbon formation
383	from the pyrolysis/gasification of lignin at different reaction conditions. Energy Fuels 28:6371-6379.
384	

385 26. Simitzis J, Sfyrakis J (1993) Pyrolysis of lignin biomass-novolac resin for the production of polymeric carbon
386 adsorbents. J Anal Appl Pyro. 26:31-52.

387

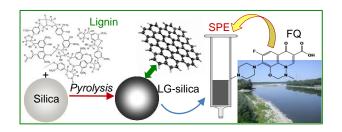
- 388 27. Wang Z et al. (2015) Nanocarbons from rice husk by microwave plasma irradiation: from graphene and carbon
 389 nanotubes to graphenated carbon nanotubes hybrids Carbon 94:479–484.
- 390
- 391 28. Paris O, Zollfrank C, Zickler GA (2005) Decomposition and carbonisation of wood biopolymers–a microstructural
 392 study of softwood pyrolysis. Carbon 43:53–66.
- 393
- 29. Custodis VBF, Bährle C, Vogel F, van Bokhoven JA (2015) Phenols and aromatics from fast pyrolysis of variously
 prepared lignins from hard- and softwoods. J Anal Appl Pyrol 115:214–223.
- 396
- 30. Zeffiro A, Dondi D, Speltini A, Pezzini S, Rossella F, Bellani V, Piazza V, Pingue P, Buttafava A (2013) Lignin
 pyrolysis as a novel method for graphene production: synthesis and characterization, FISMAT-The Italian National
 Conference on Condensed Matter Physics, Milan (Italy).
- 400
- 401 31. Liu Q, Wang S, Zheng Y, Luo Z, Cen K (2008) Mechanism study of wood lignin pyrolysis by using TG–FTIR
 402 analysis. J Anal Appl Pyrol 82:170-177.
- 403
- 404 32. Fushimi C, Araki K, Yamaguchi Y, Tsutsumi A (2003) Effect of heating rate on steam gasification of biomass. 2.
 405 Thermogravimetric-mass spectrometric (TG-MS) analysis of gas evolution. Ind Eng Chem Res 42:3929-3936.
- 406
- 33. Baumlin S, Broust F, Bazer-Bachi F, Bourdeaux T, Herbinet O, Ndiaye FT, Ferrer M, Lédé J (2006) Production of
 hydrogen by lignins fast pyrolysis. Int J Hydrogen Energ 31:2179-2192.
- 409
- 410 34. Rodríguez-Mirasol J, Cordero T, Rodríguez JJ (1996) High-temperature carbons from Kraft lignin. Carbon 34:43411 52.
- 412
- 413 35. Sturini M, Speltini A, Pretali L, Fasani E, Profumo A (2009) Solid-phase extraction and HPLC determination of
- 414 fluoroquinolones in surface waters. J Sep Sci 32:3020–3028.

415	
416	36. Dorival-García N, Zafra-Gómez A, Camino-Sánchez FJ, Navalón A, Vílchez JL (2013) Analysis of quinolone
417	antibiotic derivatives in sewage sludge samples by liquid chromatography-tandem mass spectrometry: comparison of
418	the efficiency of three extraction techniques. Talanta 106:104–118.
419	
420	37. Speltini A, Sturini M, Maraschi F, Porta A, Profumo A (2016) Fast low-pressurized microwave-assisted extraction
421	of benzotriazole, benzothiazole and benezenesulfonamide compounds from soil samples. Talanta 147:322-327.
422	
423	38. Vazquez-Roig P, Segarra R, Blasco C, Andreu V, Picó Y (2010) Determination of pharmaceuticals in soils and
424	sediments by pressurized liquid extraction and liquid chromatography tandem mass spectrometry, J Chromatogr A
425	1217:2471–2483.
426	
427	39. Pena A, Chmielova D, Lino CM, Solich P (2007) Determination of fluoroquinolone antibiotics in surface waters
428	from Mondego River by high performance liquid chromatography using a monolithic column. J Sep Sci 30:2924-2928.
429	
430	40. Tamtam F, Mercier F, Eurin J, Chevreuil M, Le Bot B (2009) Ultra performance liquid chromatography tandem
431	mass spectrometry performance evaluation for analysis of antibiotics in natural waters. Anal Bioanal Chem 393:1709-
432	1718.
433	
434	
435	
436	
437	
438	
439	
440	
441	
442	
443	
444	

445	Figure captions
446	Fig. 1 SEM images acquired on pristine silica (a), pyrolized bare silica (b), pyrolized LG@silica (c), and their related
447	pictures (a1,b1,c1)
448	
449	Fig. 2 HPLC-FD chromatograms overlay of a blank river water (500 mL) SPE extract (a) and the SPE extract of the
450	same river water sample spiked with 50 ng L ⁻¹ FQs prior pre-concentration (b)
451	
452	Fig. 3 HPLC-FD chromatogram obtained from pre-concentration of the Ditch 1 sample collected in a livestock farming
453	area near Pavia (Lombardy, Italy)
454	
455	
456	Table captions
457	Table 1 Mean recoveries (%) and inter-day precision RSDs (%) for LG-silica SPE of tap and river water samples
458	spiked with FQs
459	

460 Table 2 Calibration curves, correlation coefficients, detection and quantification limits

Graphical Abstract





Dr. Andrea Speltini Department of Chemistry, University of Pavia via Taramelli 12, 27100 Pavia, Italy

e-mail: andrea.speltini@unipv.it phone: +39 0382 987349 fax: +39 0382 528544

To the Editor of Microchimica Acta, Prof. Wolfbeis

Dear Editor,

we are sending a paper on the preparation, characterization and analytical application of silica microparticles supporting carbons from Kraft lignin pyrolysis. The material was tested as solid-phase extraction sorbent for pre-concentration of Fluoroquinolones, chosen as model emerging contaminants, from natural waters. Please find enclosed the manuscript we wish to submit for publication in **MCA** as an **Original Paper**.

Manuscript title

Silica-supported carbon from Kraft lignin pyrolysis for solid-phase extraction of fluoroquinolone probes pollutants from environmental waters

Manuscript significance

This work presents the application of a novel carbon-based material, made of silica-supported pyrolytic carbon deriving from lignin, for solid-phase extraction (SPE) of emerging contaminants from water, specifically Fluoroquinolone antibiotics (FQs), chosen as test compounds.

A hybrid material consisting of silica microparticles supporting pyrolytic sp² carbon, easily obtained from Kraft lignin as the carbon precursor, was prepared, characterized by various techniques (SEM, TGA, BET, EPR) and evaluated for pre-concentration of Marbofloxacin (MAR) and Enrofloxacin (ENR), two drugs largely used in veterinary medicine, chosen as the probe pollutants.

To the best of our knowledge, this is the first report on the application of a carbon-based material obtained by lignin pyrolysis for SPE purposes.

The material (LG-silica) was tested on tap and river water samples spiked with MAR and ENR at concentrations in the range 10-1000 ng L⁻¹. Quantitative adsorption was observed at the sample native pH for both drugs, which were eluted from the cartridge with 2×2 mL aqueous tetrabutyl ammonium hydroxide-acetonitrile (70:30), and quantified by HPLC-FD. Quantitative recoveries (70-116%, *n*=3), and suitable inter-day precision (RSDs 8-16%, *n*=3) were observed for 20-1000 ng L⁻¹ spikes, also in raw river water. Sample volumes up to 500 mL provided enrichment factors up to 125, useful for FQs determination at environmentally significant levels (few ng L⁻¹), and at the same time attractive compared to the commercial sorbents. The batch-to-batch reproducibility was verified, and the material was successfully applied to the determination of these drugs in actual environmental waters.

In conclusion, the new sorbent phase is easy to be prepared, costless, efficient and applicable for quantitative determinations in real contaminated water samples. Results suggest that use of LG-silica could be extended also to other classes of contaminants.

The paper is unpublished and not submitted for publication elsewhere. The authors declare no conflict of interest.

Best regards Andrea Speltini and co-workers Audrea Speltin'

Pavia, January 3, 2016

Electronic Supplementary Material

Click here to access/download Electronic Supplementary Material Supplementary Material.doc

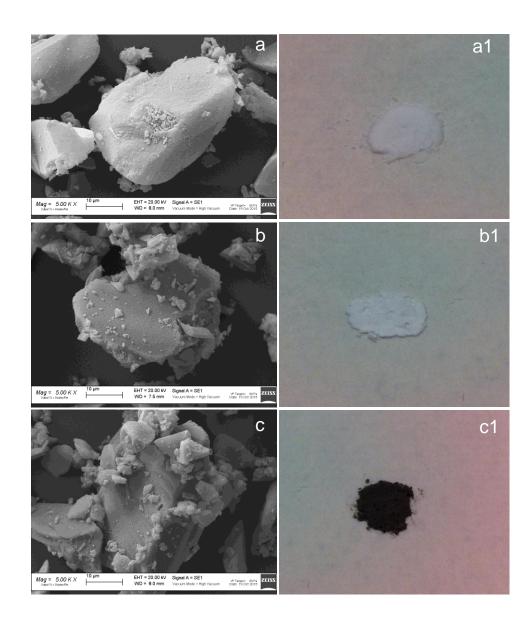


Figure 2

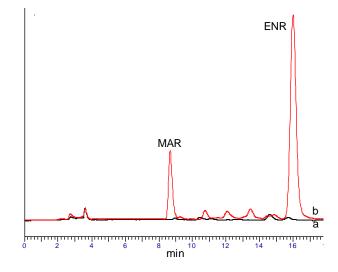
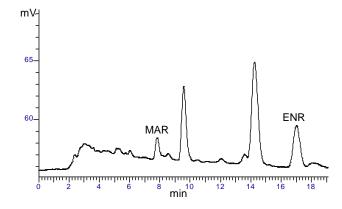


Figure 3



Mean absolute recovery (%) and inter-day precision (RSD%)							
Tap water (<i>n</i> =3)			River water (<i>n</i> =3)				
25	50	500	1000	10	20	50	100
73(16)	81(8)	71(9)	105(11)	51(15)	115(11)	116(10)	75(9)
92(12)	90(7)	93(10)	90(13)	56(7)	72(14)	70(9)	77(9)
	Tap wate 25 73(16)	Tap water $(n=3)$ 25 50 73(16) 81(8)	Tap water (n=3) 25 50 500 73(16) 81(8) 71(9)	Tap water (n=3) 25 50 500 1000 73(16) 81(8) 71(9) 105(11)	Tap water (n=3) River water wat	Tap water $(n=3)$ River water $(n=3)$ 25505001000102073(16)81(8)71(9)105(11)51(15)115(11)	Tap water (n=3)River water (n=3) 25 50 500 1000 10 20 50 $73(16)$ $81(8)$ $71(9)$ $105(11)$ $51(15)$ $115(11)$ $116(10)$

	Equation	Linearity (r^2)	IDL ^b	IQL ^b	MDL ^b	MQL ^b
	Equation ^a		(µg L ⁻¹)	(µg L ⁻¹)	(ng L ⁻¹)	(ng L ⁻¹)
MAR	y=18580(297) x - 342(425)	0.9975	0.14	0.42	1.1	3.4
ENR	y=171802(2307) x + 2073(3243)	0.9982	0.11	0.34	0.9	2.7

^aCalculated as peak area (y) vs. analyte concentration (x); in parentheses slope and intercept uncertainties obtained by wLLSR (weighted linear least squares regression). ^b Calculated from wLLSR parameters.