1	Oligonucleotide-based interventions targeting VEGF in eye
2	neovascularization: potential synergy between present and future molecules
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14	Abstract
15	Roughly ten years ago the FDA approved most of the presently used anti-VEGF drugs for the
16	treatment of neovascular AMD and other eye pathologies characterized by ocular neoangiogenesis.
17	However, the recent findings on the physiologic activities of VEGF isoforms impose to reconsider
18	the inhibitory effects of pan-VEGF antagonists and the concept that to face pathological alterations
19	at ocular level is possible only through the full block of all VEGF isoforms. In fact, although pan-
20	VEGF agents rapidly and effectively contrast ocular neovascularization, vascular leakage, and other
21	pathological changes, in the long-term the inhibition of all VEGF isoforms likely may result in the
22	loss of the physiologic effects exerted by $VEGF_{121}$ and the anti-angiogenic $VEGF_{165}b$. Notably,
23	selective inhibitors of VEGF ₁₆₅ a, such as pegaptanib, spare these targets. Moreover, preclinical and
24	clinical evidence suggest that also systemic side effects, secondary to intraocular treatment with

non-selective anti-VEGF drugs, may be reinterpreted in light of these recent findings, which may be
useful to clinicians for the choice of the most appropriate anti-VEGF agent.

Another aspect that should be considered is the involvement of VEGF-independent pathways in ocular neovascularization, therefore a combined therapy can represent a more effective pharmacological approach that might help also to counteract tachyphylaxis, an important issue in anti-VEGF treatment.

This complex picture and the recent findings on current anti-VEGF drugs should be therefore taken into account to guide the development of novel agents targeting VEGF and/or other key factors involved in the pathogenesis of neovascular ocular diseases along the signaling pathways stimulated by the various isoforms. Accordingly, this review also reports on novel pharmacological molecules targeting VEGF at ocular level and currently under development, with a special attention to oligonucleotide-based interventions.

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Keywords VEGF, ocular angiogenesis, neovascular AMD, pegaptanib, ranibizumab, bevacizumab
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40 List of abbreviations

- 41 AMD = Age-related Macular Degeneration
- 42 DR = Diabetic Retinopathy
- 43 EC = Endothelial Cell
- 44 VEGF = Vascular Endothelial Growth Factor
- 45 PLGF = Placental Growth Factor
- 46 DME = Diabetic Macular Edema
- 47 RVO = Retinal Vein Occlusion
- 48 PRN = pro re nata
- 49 VEGFR = VEGF Receptor
- 50 BRVO = Branch Retinal Vein Occlusion

- 51 NRP1 = Neuropilin-1
- 52 RPE = Retinal Pigment Epithelium
- 53 I/R = Ischemic/Reperfusion
- 54 iNOS = inducible Nitric Oxide Synthase
- 55 NO = Nitric Oxide
- 56 RGC = Retinal Ganglion Cell
- 57 OIR = Oxygen-Induced Retinopathy
- 58 SELEX = Systematic Evolution of Ligands by Exponential Enrichment
- 59 HUVEC = Human Umbilical Vein Endothelial Cell
- 60 HBD = Heparin-Binding Domain
- 61 PDGF = Platelet-Derived Growth Factor
- 62 FGF = Fibroblast Growth Factor
- 63 PEG = Polyethylene Glycol
- 64 HMVEC = Human Microvascular Endothelial Cell
- 65 RBD = Receptor-Binding Domain
- 66 CNV = Choroidal Neovascularization
- 67 VISION = VEGF Inhibition Study in Ocular Neovascularization
- 68 GA = Geographic Atrophy
- 69 PED = Pigment Epithelium Detachment
- 70 BFV = blood flow velocities (s) in the
- 71 CRA = Central Retinal Artery
- 72 TPCA = Temporal Posterior Ciliary Artery
- 73 OA = Ophthalmic Artery
- 74 ADR = Adverse Drug Reaction
- 75 PDGFR = PDGF Receptor
- $rac{siRNA} = small interfering RNA$

- 77 DDIT4 = DNA-Damage-Inducible Transcript 4
- 78 KSP = Kinesin Spindle Protein
- 79
- 80

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109 **1. Introduction**

110 Age-related macular degeneration (AMD) and diabetic retinopathy (DR) are the most common ocular diseases dramatically affecting the quality of life of patients and causing an enormous burden 111 to the healthcare system in Europe and USA [1; 2]. AMD, whose pathogenesis is multifactorial and 112 not yet fully elucidated, is classified in early and late stages, atrophic dry (85%) and exudative wet 113 neovascular (15%) degeneration categories [3]. Quite similarly, DR, a sight-threating complication 114 115 of diabetes developed by more than one-third of diabetic individuals, is classified in two stages: non proliferative and proliferative [4; 5]. In some cases, both non proliferative AMD and DR may 116 progress and convert to proliferative neovascular forms, in which the formation of new vessels from 117 118 the existing ones, a process named "ocular angiogenesis", represents a major cause of visual loss. Neovascular AMD and DR are the leading causes of blindness in elderly and working age people, 119 respectively [6; 7]. These pathologies are associated with neovascularization in the posterior 120 121 segment of the eye; in particular, the hallmark of wet AMD is the formation of new blood vessels arising from the choroidal microvascular bed and invading the sub-retinal space, while DR presents 122 123 alterations preferentially at retinal level [8; 9]. Although with some differences, both neovascular AMD and DR are characterized by endothelial cell (EC) proliferation and migration, increase in 124 vascular permeability and inflammation, all processes in which Vascular Endothelial Growth 125 Factor-A (VEGF-A) plays a key role. In mammals, VEGF-A belongs to a family that also includes 126 VEGF-B, -C, -D, and placental growth factor (PLGF). Among these closely-related growth factors, 127 VEGF-A is the most potent mediator of both retinal and choroidal angiogenesis, and its inhibition 128 via intraocular anti-VEGF treatments currently represents the cornerstone of therapies for both 129 AMD and DR [10; 11]. The outcomes of anti-VEGF treatments are to counteract pathological 130 neovascularization and disease progression, to arrest visual impairment and, in the best case, to gain 131 the recovery of vision. Some molecules targeting VEGF-A pathway and acting at multiple levels 132 are currently used in ophthalmology, and much more are under investigation in clinical trials for 133 either AMD, DR, or other eye diseases characterized by neovascularization. These anti-VEGF-A 134

drugs can be divided in three main pharmacological classes: 1) molecules targeting VEGF isoforms, 2) molecules inhibiting VEGF receptors, and 3) molecules inhibiting VEGF downstream signaling [12]. This review will focus on current therapies and novel substances under development belonging to the first pharmacological class, with a special attention to oligonucleotide-based interventions targeting VEGF at ocular level.

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141 2. Pharmacodynamic classification and clinical application of drugs acting upon VEGF

Several agents have been developed to interfere with the VEGF system and various molecules are already used especially in cancer therapy. A few of them are approved as ophthalmic therapies, such as ranibizumab, pegaptanib and aflibercept, others are under investigation and/or currently used off-label like bevacizumab. They mainly act on VEGF itself or on its gene expression and comprise some promising molecules, under preclinical or clinical investigation, that will be briefly illustrated later on in this review.

A fundamental class of therapeutics acting as angiogenesis inhibitors is represented by monoclonal 148 149 antibodies characterized by a high specificity for a given target, which they bind and neutralize. In this category, ranibizumab is an engineered recombinant humanized Fab fragment of 48 kDa 150 designed from the full-length monoclonal antibody bevacizumab to optimize retinal penetration. 151 Ranibizumab binds with high affinity a site present in all VEGF-A isoforms (see below) and their 152 bioactive proteolytic fragments [13]. On the market since 2006, ranibizumab is approved by FDA 153 for the treatment of all lesion types in neovascular AMD, diabetic macular edema (DME), and 154 macular edema secondary to retinal vein occlusion (RVO) [14]. Its intraocular administration is 155 recommended at a dosage of 0.5 mg/month, and treatment protocols usually advice an initial 156 loading dose of three monthly injections followed by administration pro re nata (PRN) based on the 157 disease activity. However, as for the other anti-VEGF treatments, the optimal regimen has yet to be 158 established, although recently the ophthalmic community has made available guidelines on the 159

treatment of patients with advanced AMD also to help clinicians to prevent over- and/or undertreatment with anti-VEGF therapy [11; 15].

Bevacizumab is a full-length humanized monoclonal IgG antibody of 149 kDa targeting the same site of ranibizumab, and thus inhibiting all VEGF-A isoforms. Approved in 2004 by FDA for systemic use in the treatment of certain metastatic cancers, it is widely used off-label as intravitreal therapy in proliferative eye diseases, especially neovascular AMD and DME, although with various limitations of medical, financial and ethical nature [16; 17; 18]. The recommended dose for intraocular treatment is 1.25 mg/month.

Aflibercept, or VEGF Trap-Eye, is a fully human recombinant protein of 115 kDa consisting of key 168 169 binding domains from VEGF receptor (VEGFR) -1 and VEGFR-2 fused to an IgG Fc fragment [19]. It acts as a soluble decoy receptor recognizing and neutralizing all VEGF-A isoforms, with the 170 establishment of a tighter binding than the native receptors. Unlike the anti-VEGF monoclonal 171 172 antibodies currently in use, aflibercept is purposely designed to inhibit also VEGF-B and PLGF-1 and -2 [20]. It has been approved by FDA for the treatment of neovascular AMD at the end of 2011, 173 174 and for RVO in 2012 [21]. The same molecular structure has been approved in 2012 by FDA also for systemic use within the combination therapy for patients with metastatic colorectal cancer with 175 the name of ziv-aflibercept. The two products present substantial differences in both the preparation 176 177 of the purified aflibercept and the drug formulation, indeed aflibercept/VEGF Trap-Eye undergoes more purification steps during manufacturing than ziv-aflibercept, and it is formulated with proper 178 buffers to minimize the risk of ocular irritation. The recommended regimen is an intravitreal dose of 179 2 mg/month for three consecutive treatments, followed by one injection every two months; a 180 possible variant is represented by bimonthly injections from the beginning of the therapy [22]. 181

Besides the just mentioned drugs, which are all proteins targeting pan-VEGF-A isoforms, there is another drug somehow unique in its own way: pegaptanib, a 28-nucleotide RNA aptamer of \sim 50 kDa with a high selectivity for the VEGF-A₁₆₅ isoform. Pegaptanib sodium was the first drug officially registered for the treatment of neovascular AMD in 2004, and also the first aptamer to

enter in therapy. Aptamers can be envisioned as "chemical antibodies" since they offer the advantages of antibodies - high specificity and affinity - in a relatively small, chemically synthesized molecule without cell-culture-derived contaminants. Moreover, aptamers are highly versatile and their commercial synthesis by large-scale manufacturing is fairly uncomplicated and cost-effective [23]. Pegaptanib is used off-label also for DME and branch RVO (BRVO). The intravitreal injection is at a dose of 0.3 mg once every six weeks.

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3. The main target of anti-VEGF agents: VEGF₁₆₅ among pro-angiogenic and anti-angiogenic function - from one main actor to two opponents

The human *VEGF-A* gene is organized in 8 exons separated by 7 introns [24; 25] resulting in the generation of at least 7 isoforms of VEGF-A_{xxx}, where _{xxx} is the number of amino acids encoded. The various isoforms contain between 121 and 206 amino acids, where VEGF-A₁₆₅ is the predominant member, followed by VEGF-A₁₂₁ [26].

In recent years, great interest raised the discovery by Bates et al. of the VEGF-A₁₆₅b isoform, 199 200 formed by an alternative splicing at exon 8 of VEGF-A gene [27]. Now we know that the VEGF-A family is composed by two subfamilies of splice variants which differ for an alternative C-terminal 201 sequence: the "conventional" VEGF-Axxx (or VEGF-Axxxa), formed by proximal splice-site 202 selection, and the "novel" VEGF-A_{xxx}b, generated by a splicing 66 bases downstream. The 203 alternative splicing of VEGF-A (since now referred as VEGF) thus results in proteins of the same 204 length but with different amino acids at the C-terminus: Cys-Asp-Lys-Pro-Arg-Arg in VEGF_{xxx}a 205 isoforms, and Ser-Leu-Thr-Arg-Lys-Asp in VEGF_{xxx}b subfamily, respectively [27]. More 206 importantly, the alternative splicing of exon 8 is the key determinant of isoform switching from 207 208 "pro-angiogenic" VEGF_{xxx}a to "anti-angiogenic" VEGF_{xxx}b.

All VEGF-A isoforms present the VEGFR-binding site and activate both Flt-1/VEGFR-1 and Kdr/VEGFR-2, although with different affinity and potency. VEGFR-1 mediates VEGF-induced chemotaxis and inflammation, while VEGFR-2 is the main mediator of the mitogenic, angiogenic

and permeability-enhancing effects of VEGF [28; 29]. In endothelial cells VEGFR-2 can be co-212 expressed with the neuropilin-1 (NRP1) co-receptor, which enhances VEGF binding to VEGFR-2 213 and its signal transduction up to 6-fold [30] (see also Figure 1). NRP1 is an isoform-specific 214 receptor for VEGF₁₆₅a [31]; VEGF₁₆₅a binding to VEGFR-2 and NRP1 results in receptor 215 dimerization, rotation of its intracellular domain, and its autophosphorylation [32; 33]. Conversely, 216 this full rotation likely does not occur when VEGF₁₆₅b binds to VEGFR-2, resulting in an 217 inefficient autophosphorylation of the receptor [31; 32]. In respect to its angiogenic counterpart, 218 VEGF₁₆₅b isoform is indeed a weaker agonist, since it binds and poorly activates VEGFR-2, 219 resulting in differential activation of intracellular pathways and, most relevantly, not inducing 220 vasculogenesis [32]. This feature seems to be due to the difference in the C-terminal end of 221 VEGF₁₆₅b isoform. In fact, VEGF₁₆₅b does not bind to NRP1 [30; 32], probably because its 222 adjacent C-terminal sequence lacks Cys-160, which instead is present in VEGF₁₆₅a, affecting its 223 224 secondary structure and folding, and likely VEGFR-2 signaling [27]. It is possible that also the neutral charge conferred by the two terminal residues (Lys-Asp) of VEGF₁₆₅b contributes to the 225 226 lack of interaction with NRP1 and the differential VEGFR-2 downstream signaling [31].



Figure 1. VEGF₁₆₅ interacts with VEGFR-2 and NRP1 in endothelial cells. VEGF₁₆₅ functions as a dimer and its binding to VEGFR-2 promotes the receptor dimerization with subsequent activation of tyrosine kinase domains and downstream signaling pathways responsible of the effects reported. In VEGF₁₆₅ is present a disulfide bond between Cysteines 146 and 160 (in orange color), necessary for VEGF activity and its interaction with NRP1 (for more details see text, paragraph 3).

4. Heterogeneous activities of the different VEGF isoforms: a hot topic to understand the effects of selective and non-selective VEGF inhibition

Since the beginning the predominant concept was that blocking all the VEGF-A isoforms, instead of only VEGF₁₆₅, was the winning strategy to face neovascularization, therefore the most commonly used anti-VEGF molecules injected for neovascular ocular diseases have been ranibizumab and bevacizumab. More recent studies on the role of the various VEGF isoforms and their biological effects, as well as relevant clinical evidence raised in pharmacovigilance, impose to
reconsider previous findings on the molecular mechanisms of anti-VEGF drugs in light of the
advanced knowledge.

VEGF is secreted by several cell types in the retina, such as vascular ECs, retinal pigment epithelium (RPE), pericytes, retinal neurons, and astrocytes, indicating that VEGF plays important functions in ocular homeostasis [34]. In particular, VEGF is a critical factor for the homeostasis and plasticity of both blood vessels and neurons.

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248 4.1. The "conventional" VEGF isoforms and the vasculature

VEGF is essential for EC survival in normal conditions, and adult blood vessels require autocrine 249 VEGF for maintenance of homeostasis [35]. In vivo treatment with a soluble receptor VEGF-Trap 250 in adult mice leads to early regression of normal blood capillaries, in a sequence of events 251 252 comprising cessation of blood flow, EC apoptosis, pericytes migration from regressing vessels to surviving ones, and formation of acellular capillaries, also called basement membrane ghosts [36]. 253 254 The features of the various VEGF isoforms and proteolytic products, their tissue expression and roles during development and in adult, have been recently reviewed [37]. The specific functions of 255 VEGF isoforms in vascular patterning have been elucidated by transgenic mouse models. Mice 256 selectively expressing only VEGF₁₆₄ present normal retinal vascular development, while mice 257 expressing only VEGF₁₂₀ have severely impaired outgrowth and patterning of developing retinal 258 vessels [38; 39], indicating that VEGF₁₆₄ plays the main role in vasculogenesis. However, 259 VEGF₁₆₄-deficient mice expressing both VEGF₁₂₀ and VEGF₁₈₈ showed no difference in 260 physiological neovascularization when compared with wild-type control animals, indicating that 261 other VEGF isoforms, in combination, may compensate VEGF₁₆₄ lack and be sufficient to promote 262 normal physiological neovascularization [40]. From the development through all the life, the VEGF 263 isoforms act in concert to assure the optimal formation and maintenance of an adequately branched 264 vessel network, guiding specific ECs in growing and shaping the vascular tree [41-44]. Importantly, 265

of the total amount of VEGF secreted by RPE cells, 75% is represented by VEGF₁₆₅ and 24% by 266 VEGF₁₂₁ [45]. The various isoforms differ not only, as previously mentioned, for alternative 267 splicing but also for their distribution: for instance, VEGF₁₂₁ is freely diffusible; VEGF₁₈₈, which 268 contains two heparin sulfate binding sites, is sequestered by cell membrane or extracellular matrix; 269 VEGF₁₆₅ is in both soluble and bound status. Soluble isoforms are essential for maintenance of RPE 270 and choriocapillaries in the adult, indeed the absence of both VEGF₁₂₀ and VEGF₁₆₄ in mice leads 271 272 to an age-dependent degenerative phenotype characterized by RPE dysfunction, loss of barrier properties, insoluble drusen-like deposits resembling atrophic AMD [46]. 273

VEGF₁₆₅ appears to be a critical isoform for retinal angiogenesis not only under development but 274 275 also in pathological conditions; among the VEGF isoforms, VEGF₁₆₅a is indeed the principal mediator of inflammation and cellular immunity occurring in pathological retinal 276 neovascularization, acting as a proinflammatory cytokine targeting monocytes, macrophages and 277 leukocytes, in a positive feedback loop involving primarily ECs and sustaining the 278 neovascularization process [40; 47]. The expression pattern of VEGF isoforms, which is strictly 279 regulated in normal conditions, is disrupted in diseases; for example, in vivo it has been shown that 280 in pathological retinal neovascularization of rodents, the VEGF₁₆₄/VEGF₁₂₀ ratio undergoes a ten-281 fold increase (~25.5 versus ~2.2 in the physiologically developing retina), and this likely 282 283 contributes to an angiogenic switch and to the appearance of inflammation-associated vessel invasion within the vitreous [40]. VEGF₁₂₁ has less affinity for VEGFRs than VEGF₁₆₅, explaining 284 the lower mitogenic, proinflammatory potency of VEGF₁₂₁ relative to VEGF₁₆₅ [48; 49]. 285

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4.2. The "conventional" VEGF isoforms and the neurons

VEGF plays a fundamental role also in neurogenesis and neuroprotection, and VEGF abnormal expression has been linked to several neurodegenerative disorders [50-52]. *In vitro* studies demonstrated that VEGF, in particular the 165 isoform, increases the survival of various types of neurons under different stresses via VEGFR-2 through multiple molecular mechanisms including

phospholipase C, PI3K, p38/MAPK and MEK1/2 activation, caspase-3 inhibition, and modulation 292 293 of ion channel currents [53-57]. Protective effects are also exerted on supporting cells, such as astrocytes and microglia, through VEGFR-1 [58]. In vivo, intraocular injection of either VEGF₁₂₀ or 294 VEGF₁₆₄ prevents retinal neuron apoptosis resulting from ischemic/reperfusion (I/R) injury in the 295 rat, acting via VEGFR-2 signaling; however, the retina injected with VEGF₁₆₄ develops 296 hemorrhages and edema that were not detected after $VEGF_{121}$ injection [59], suggesting that 297 VEGF₁₂₁ is more implicated in the homeostasis of neurons and vessels than in pathologic contexts. 298 It is well known that downstream the VEGF signaling is induction of Nitric Oxide Synthase 299 (iNOS), leading to a potent vasodilation [60], and that increasing volumetric blood flow enhances 300 301 neuroprotection in ischemic tissues [61]; interestingly, the authors showed that the $VEGF_{121}$ mediated neuroprotection in the I/R retina is only partially dependent by an increase of iNOS and 302 blood flow, and that VEGF₁₂₁ exerts a pro-survival action also on retinal ganglion cells (RGCs) in 303 304 vitro, strongly suggesting a direct neuroprotective effect of VEGF₁₂₁ [59]. In the same in vivo model, a brief ischemic preconditioning increases both VEGF₁₂₀ and VEGF₁₆₄ expression as a 305 306 neuroprotective response, and this benefic pro-survival effect on RGCs is suppressed by either VEGFR-1/Fc fusion protein or an anti-VEGF antibody; conversely and of great interest, pegaptanib 307 does not impair RGCs viability, further suggesting that not fully abrogating VEGF responses, and 308 especially sparing VEGF_{120/121}, represent a key strategy to preserve retinal neurons [59]. More 309 recently the same group confirmed these findings in other in vitro and in vivo models on RGCs 310 death, showing that the pro-survival effects of VEGF₁₂₁ are mediated by VEGFR-2 and PI3K/Akt 311 312 signalings, and that total VEGF blockade significantly exacerbates neuronal cell death [62].

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4.3. The emerging role of VEGF₁₆₅b in blood vessels and neurons

As mentioned, it has been discovered that VEGF family is actually constituted of two subfamilies, whose differential functions still need to be fully investigated. It has been reported that in normal conditions the "anti-angiogenic" VEGF_{xxx}b isoforms represent the predominant proportion of the

total VEGF in human eye tissues and fluids; in particular, VEGF₁₆₅b is highly expressed in the 318 319 normal eye (retina, lens, sclera, iris, vitreous) and it is down-regulated in the vitreous fluid of DR patients, where a switch in VEGF splicing from anti- to pro-angiogenic isoforms likely occurs [63]. 320 This alteration in VEGF balance between the two subfamilies seems to be also present in other 321 angiogenic-associated conditions, as shown in oxygen-induced retinopathy (OIR) mouse model and 322 in laser-induced choroidal neovascularization of AMD mouse model [64; 65]. In humans, beside in 323 DR, significantly lower levels of VEGF₁₆₅b have been also revealed in the vitreous fluid from 324 patients with RVO [66] but not with AMD, although a trend towards a reduction (-20% VEGF₁₆₅b 325 median vs control subjects) was detected [67]. Interestingly, in animal models of OIR and AMD, 326 327 intraocular injections of recombinant human VEGF₁₆₅b inhibit retinal neovascularization [64; 68; 69]. To this regard, in vitro and in vivo studies have demonstrated that VEGF₁₆₅b inhibits several 328 VEGF₁₆₅a-induced processes, such as EC migration and proliferation, vasodilatation [27] and 329 pathological angiogenesis in many tumor types [70; 71]. And as, VEGF₁₆₅b seems to be more than 330 simply an anti-angiogenic factor, since in a dose dependent manner it is able not only to inhibit the 331 pathologic pre-retinal proliferation of new vessels, but also to reduce the ischemic area in OIR 332 animal model [68; 69]. Specifically, these studies show that VEGF₁₆₅b favors physiological 333 revascularization in vivo and acts as a survival factor for both retinal endothelial and epithelial cells 334 in vitro, these latter effects being observed also with its sister isoform VEGF₁₆₅a [69]. VEGF₁₆₅b 335 inhibits VEGF₁₆₅a-induced EC migration and proliferation but does not interfere with regrowth of 336 blood vessels within previously vascularized areas, a process named "revascularization" [69; 72], 337 suggesting that VEGF₁₆₅b contrasts the invasive phenotype and promotes physiological 338 angiogenesis, in agreement with its weak agonist activity. Consistently, in a rat model of 339 proliferative retinopathy, VEGF₁₆₄ blockade by EYE001/pegaptanib inhibits pathological 340 neovascularization but not physiological revascularization, resembling the VEGF₁₆₅b-mediated 341 effects; in contrast, a VEGFR-1/Fc chimera blocking all VEGF isoforms suppresses both 342 pathological and physiological retinal neovascularization [40]. 343

In vitro and *in vivo* evidence also demonstrate that VEGF₁₆₅b is neurotrophic and exerts neuroprotective effects in response to multiple insults in various types of neurons; in particular, *in vivo* VEGF₁₆₅b pretreatment protects RGCs and the inner nuclear layer cells in rat retinal I/R injury model [73]. As for VEGF₁₆₅a, VEGF₁₆₅b-mediated neuroprotection is through VEGFR-2, p44/42 MAPK activation, and caspase-3 inhibition, but, in contrast to VEGF₁₆₅a, it does not involve either PI3K or p38 MAPK [73].

On the whole, these findings implicate that blocking all VEGF isoforms leads rapidly to an effective inhibition of ocular neovascularization, but in the long term this may result in detrimental effects at both vascular and neuronal retinal level, mainly due to the loss of the physiologic effects mediated by $VEGF_{121}$ and $VEGF_{165}$ b. Local and systemic effects secondary to intraocular treatment with pan-VEGF drugs may be also reinterpreted in light of these evidence, as reported below.

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5. Developing the aptamer strategy to interfere with VEGF

The story of pegaptanib discovery and development started in the '90s, when precursors of 357 pegaptanib were selected among huge libraries of oligonucleotides, about 10¹⁴ RNA molecules 358 containing 30 randomized positions [74]. By using the SELEX (Systematic Evolution of Ligands by 359 Exponential Enrichment) technology developed by Larry Gold's group at the Colorado University 360 in the early 1990s [75], specific inhibitors of VEGF-A₁₆₅ (from now referred as VEGF₁₆₅) were 361 searched. The pathogenic role of VEGF₁₆₅ in tumor vascularization and growth was evidenced few 362 years earlier [76-78]. The identification of VEGF₁₆₅ as the main actor of pathological angiogenesis 363 on one side and, on the other, the concurrent research on aptamers as novel therapeutics, converged 364 to the development of some promising anti-VEGF RNA ligands. For the initial selection of the anti-365 VEGF aptamers the investigators used as target the recombinant human VEGF₁₆₅ [74]. 366

The first work on anti-VEGF aptamers, published in 1994, reported an initial set of high affinity RNA ligands selected for their ability to inhibit the binding of [¹²⁵I]VEGF to its receptors in a concentration-dependent manner in human umbilical vein endothelial cells (HUVECs) [74].

Competition experiments on candidate anti-VEGF aptamers revealed that they all bound to a similar 370 371 site within the heparin-binding domain (HBD) of VEGF₁₆₅, and indeed they were displaced by heparin [74]. To address the question of specificity of these high-affinity ligands towards VEGF₁₆₅, 372 the scientists performed binding experiments with various heparin-binding proteins, such as PDGF 373 (Platelet-Derived Growth Factor) and FGF (Fibroblast Growth Factor). In those years, specific 374 inhibitors of VEGF, basically pan-VEGFs, were limited to monoclonal antibodies [79] and soluble 375 VEGF receptor [80], thus the isolation of RNA molecules with unexpected binding selectivity to 376 VEGF₁₆₅ aroused a great interest. The lead compounds were further modified and optimized [81; 377 82], t44-OMe having high binding affinity for VEGF₁₆₅ and the best activity in the Miles assay, an 378 379 in vivo test to evaluate the capacity of a given substance to inhibit vascular leakage following VEGF intradermal injection in guinea pigs [82]. Interestingly, the addition of a 40 kDa polyethylene glycol 380 (PEG) to the 5'-end of the 27 nucleotides long t44-OMe slightly decreased (about 4 fold) the 381 382 binding affinity to VEGF₁₆₅, but markedly improved the inhibitory activity in the Miles assay (83%) with PEG-conjugated t44-OMe; 48% with t44-OMe) [82]. The more efficient inhibition of VEGF-383 384 induced permeability displayed by PEG-t44-OMe might be the consequence of a prolonged tissue permanence of the aptamer due to its conjugation with PEG [83]. Starting from PEG-t44-OMe, and 385 few intermediates such as NX1838 and EYE001, pegaptanib was finally generated [84; 85]. 386 Pegaptanib sodium is a covalent conjugate of 28 nucleotides in length that terminates in a 387 pentylamino linker, to which two 20 kDa monomethoxy PEG units are attached via the two amino 388 groups on a lysine residue. This drug represented the first pharmacologic approach joining the laser 389 photocoagulation and verteporfin photodynamic therapy, considered at that time the sole therapeutic 390 interventions for neovascular ocular diseases. 391

The literature on precursors of pegaptanib provides information on their binding to VEGF, and it is thus useful to better understand the molecular mechanism of inhibition of this aptamer and to clarify its biological activity. Since the beginning it was evident that the candidate aptamers under development were displaced from VEGF₁₆₅ by heparin, suggesting that the HBD of VEGF was

involved in the ligand-protein binding [74]. Accordingly, it was then shown that the amino acid 396 sequence of VEGF that remained photo-cross-linked to the aptamer after digestion corresponded to 397 a specific site within the HBD of VEGF₁₆₅; more precisely, an uridine residue within the minimal 398 RNA sequence capable of high affinity binding to VEGF formed a cross-link with the residue 399 cysteine-137 within the exon 7-encoded domain of VEGF₁₆₅, mediating much of the heparin 400 binding activity of VEGF [82]. These findings in vivo were confirmed, demonstrating that the 401 402 uridine-14 of the therapeutic aptamer forms a cross-link with cysteine-137 and that the HBD is the primary determinant for the affinity and specificity in the complex formed by the aptamer and 403 VEGF₁₆₅ [86]. Notably, the HBD is completely lacking in the VEGF₁₂₁ isoform. 404

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406 6. Sparing VEGF₁₆₅b while targeting VEGF

The previously described differences in the C-terminal structures of VEGF may also explain the fact that pegaptanib selectively binds the angiogenic VEGF₁₆₅a, likely sparing the anti-angiogenic counterpart, as better elucidated below. In other words, the C-terminus of VEGF₁₆₅a may be the key determinant for both the interaction between HBD and NRP1, and the binding between pegaptanib and HBD. In agreement, NMR data and three dimensional solution structures of aptamer-ligand complexes reveal that specificity and affinity for a given binding site are profoundly influenced by the near residues, which affect the adaptive recognition by the aptamer [87].

Interestingly, as later reported by the same group which first identified [27] and characterized 414 VEGF₁₆₅b in terms of mechanism of action and expression [72], pegaptanib does not bind to 415 VEGF₁₆₅b [69]. Specifically, to evaluate whether a direct interaction between pegaptanib and 416 VEGF₁₆₅b exists, the authors incubated the RNA aptamer with VEGF₁₆₅a or VEGF₁₆₅b protein, 417 418 separated the samples on acrylamide gel under non-denaturating conditions, and probed the membranes with either an anti-VEGF₁₆₅ antibody detecting both VEGF isoforms, or a specific anti-419 VEGF₁₆₅b antibody directed to the nine C-terminal amino acids of VEGF₁₆₅b [69], which is 420 currently being used to detect this specific isoform in human tissues [88]. The blot showed a band 421

shift of VEGF₁₆₅a, but not VEGF₁₆₅b, toward higher molecular weight when the aptamer was 422 423 added, suggesting that pegaptanib does not physically interact with VEGF₁₆₅b [69]. The same authors reported that pegaptanib and VEGF₁₆₅b given separately to human microvascular 424 endothelial cells (HMVECs) inhibit the VEGF₁₆₅-induced cell migration; however, the concomitant 425 treatment with the two inhibitors removes the benefit of each agent in the same assay, suggesting 426 that, although there is not a direct interaction between pegaptanib and VEGF₁₆₅b, the combination 427 of the two molecules does not add benefit [69]. This may be explained considering that VEGF 428 functions as a dimer and, although till now it has been solely proven the existence of $VEGF_{165/110}$ 429 heterodimers [48], an alternative is that VEGF₁₆₅b may pair with VEGF₁₆₅a reducing the angiogenic 430 431 potential of the latter through a differential VEGFR signaling, and that pegaptanib may interfere with the formation of this heterodimer. However, further studies are needed to clarify this point. 432

In contrast to pegaptanib, it has been shown that bevacizumab binds to both VEGF₁₆₅a and VEGF₁₆₅b with equal affinity [89], and likely it is the same for ranibizumab, since it was designed on the parent molecule bevacizumab. *In vivo* studies in a cancer model report that VEGF₁₆₅b strongly impairs the efficacy of bevacizumab and this fact implicates that patients expressing high levels of VEGF₁₆₅b may be no-responders to bevacizumab and other pan-VEGF drugs [89]. These findings may also explain some of the undesired side effects of pan-VEGF agents.

439

440 7. Gaining insight into pegaptanib mechanism of action and its biological target

441 The differential biological effects, and related therapeutic profile, exerted by pegaptanib in 442 comparison with pan-VEGF drugs mainly reside in the aptamer inhibitory activity selectively 443 targeted toward VEGF₁₆₅ – or we may even say the pro-angiogenic VEGF₁₆₅a isoform.

It has been suggested that pegaptanib, by binding to a site within HBD, contrasts HBD interaction with NRP1 co-receptor and thus only exerts an inhibitory effect on the NRP1-mediated amplification of VEGFR signaling, instead of an efficient inhibition of the VEGFR signaling itself [90]. In support of this hypothesis there is the observations that VEGF₁₂₁, which contains the

receptor-binding domain (RBD) but lacks the HBD and does not interact with NRP1, activates both 448 449 VEGFR-1 and VEGFR-2, although with much lower potency than VEGF₁₆₅ [48]. However, from the beginning, experimental evidence showed that the pegaptanib precursor t44-OMe efficiently 450 blocked the binding of VEGF₁₆₅ to both Flt-1/VEGFR-1 and Kdr/VEGFR-2 [82]. Accordingly, it 451 was then clearly shown that also the pegaptanib precursor NX1838 bound Kdr/VEGFR-2; in 452 contrast, unambiguous data on VEGFR-1 were not produced because of the lack of specific anti-Flt-453 454 1/VEGFR-1 antibodies at that time [84]. Moreover, internal data Bausch & Lomb [referred by S. Giuffrida] support the assertion that pegaptanib can effectively inhibit VEGF₁₆₅ binding to its 455 receptors, VEGFR-1 (IC₅₀=0.47 nM), VEGFR-2 (IC₅₀=1.10 nM), and NRP1 (IC₅₀=0.23 nM). This 456 report is based on cell-free receptor plate binding studies and shows that the maximal inhibition 457 exerted by pegaptanib on VEGF₁₆₅ binding to the different VEGF receptors is 75-90% (the lowest 458 for VEGFR-1; the highest for VEGFR-2 and NRP1), suggesting subtle differences in the binding of 459 460 VEGF₁₆₅ to its receptors. To this regard, an hypothesis is that the aptamer inhibits VEGFR signaling by providing a steric interference between RBD and the cell-surface receptors, as previously 461 evidenced for some anti-angiogenic HBD-binding proteins [86]. 462

In vitro studies on HUVECs assessed that NX1838 inhibited VEGF₁₆₅ receptor binding and 463 downstream signaling events, including phosphorylation of VEGFR-2, phospholipase Cy activation, 464 calcium mobilization, and cellular proliferation [84]. In this report, the inhibition of VEGF₁₆₅-465 mediated cellular events exerted by NX1838 was comparable to that observed with an anti-VEGF 466 monoclonal antibody; in contrast, this aptamer was ineffective as an inhibitor of VEGF₁₂₁-induced 467 HUVECs proliferation [84]. NX1838 indeed did not bind VEGF₁₂₁ isoform lacking HBD, site for 468 pegaptanib binding. Internal data from Bausch & Lomb [referred by S. Giuffrida] report that 469 pegaptanib binds VEGF₁₈₉ with a lower but significant affinity than VEGF₁₆₅; we cannot exclude 470 that pegaptanib also binds exon 7-containing VEGF₁₈₃ and VEGF₂₀₆, which are expressed at very 471 low level and play a marginal role in angiogenesis. 472

473 Ranibizumab and bevacizumab bind to the RBD sequence [13; 91] which is common to all the 474 VEGF isoforms, thus blocking the binding of all of them to VEGFRs and the related angiogenic 475 signaling; this justifies the more potent effect of these drugs in inhibiting EC migration, 476 proliferation and vascular permeability in comparison to pegaptanib. On the other side, this strength 477 of pan-VEGF drugs may also represent their weakness, since such molecules counteract also the 478 physiologic effects of VEGF₁₂₁ and VEGF₁₆₅b.

479

480 8. Pegaptanib from biological target definition to current therapeutic use

Preclinical in vitro and in vivo studies demonstrated that pegaptanib precursors inhibit two main 481 VEGF₁₆₅-mediated functions, the enhancement of EC proliferation and vascular permeability [82; 482 84], providing the rationale for the therapeutic use of this aptamer for pathologies characterized by 483 angiogenesis and vascular leakage. EYE001 (later on designated pegaptanib) was tested in human 484 485 tumor xenograft mouse model and in various animal models of ocular neovascularization, such as the Miles assay, the rat corneal neoangiogenesis and the mouse retinopathy of prematurity models, 486 487 showing significant attenuation of VEGF₁₆₅-mediated effects in eye diseases [85]. The open-label phase IA safety study on 15 patients with subfoveal choroidal neovascularization (CNV) secondary 488 to AMD revealed no significant safety issues at 0.25-3 mg doses of EYE001/pegaptanib, and that 489 80% of subjects showed stable or improved vision 3 months after a single intravitreous injection 490 [85], opening the way to larger clinical trials on patients with exudative AMD. Pegaptanib was then 491 evaluated in two concurrent, multi-center, prospective, randomized, double-blinded, sham-492 controlled, dose-ranging trials on patients with all types of wet AMD: the VEGF Inhibition Study in 493 Ocular Neovascularization (VISION) trials. These studies showed that pegaptanib treatment every 6 494 weeks reduced vision loss by about 50% in the first year and maintained this benefit stabilizing 495 vision acuity at the second year [92; 93]. In particular, the pooled analysis of these phase III trials 496 showed that 70% of patients treated with pegaptanib 0.3 mg versus 55% of patients receiving the 497 sham injection lost fewer than 3 lines of visual acuity on an ETDRS vision chart. Within the context 498

of the 2-year trial, an exploratory analysis at week 54 of vision outcomes of a subgroup of naïve 499 patients with early CNV secondary to AMD, suggested that pegaptanib treatment is associated with 500 enhanced vision benefits, likely due to increased preservation of photoreceptors and/or RPE [94]. 501 Analogously, the efficacy of pegaptanib as primary therapy for patients with early CNV-AMD was 502 evaluated in a retrospective study with a mean follow-up of about 9 months (range 6-14 months), 503 which showed a 90% rate of improvement or stabilization of vision outcomes for pegaptanib, 504 benefits that exceeded those reported in the VISION trial [95]. A retrospective study of patients 505 with exudative AMD with small lesion size and followed up over 1 year compared the effect of 506 pegaptanib versus ranibizumab, concluding that the visual outcomes of the two drugs were 507 equivalent [96]. However, after the approval of pan-VEGF inhibitors, these latter have been 508 preferred, pegaptanib monotherapy was reconsidered, and this drug was evaluated as a maintenance 509 therapy following non-selective anti-VEGF agents in wet AMD [97]. To this regard, in a small 510 511 number of patients with all types of wet AMD, induction therapy with bevacizumab followed by pegaptanib maintenance produced visual acuity and anatomical improvements at 54-week [97]. The 512 513 efficacy and safety of pegaptanib as a maintenance therapy was then assessed on a larger scale with a phase IV, open-label, uncontrolled exploratory study including patients with subfoveal wet AMD 514 [LEVEL study; 98]. The results showed that pegaptanib was safe and well tolerated, and that the 515 visual acuity and anatomical improvements gained during the induction phase were well preserved 516 at 54-week, with only 50% of patients requiring a booster treatment given after a mean of 5 months 517 post-baseline. Similar results were reported at 54-week for Japanese patients with neovascular 518 AMD enrolled in the multi-center, prospective LEVEL-J study [99], and were further confirmed on 519 a small subgroup of patients after a 3-year follow up [100]. According to these findings, pegaptanib 520 is presently used mainly as a maintenance therapy following pan-VEGF agents in long term 521 treatment of wet AMD. Besides this indication, pegaptanib is used off-label for proliferative DR, 522 DME, BRVO, and myopic choroidal neovascularization [101-107]. 523

525 9. Selective versus non-selective anti-VEGF drugs side effects: an ongoing debate from the 526 bench to the eve of patients

Notably, non-selective anti-VEGF drugs, especially monoclonal antibodies, obtained very good 527 results in numerous clinical trials (such as ANCHOR, CATT, IVAN, MARINA, HORIZON 528 studies) to the extent that they are considered the most effective therapies for neovascular eye 529 diseases, but there is some concern about their potential local and systemic side effects, especially 530 in the long-term period [as reviewed in 34]. Ideally, an effective and safe anti-VEGF therapy should 531 reduce neovascularization without damaging the normal vessels, and also preserving the 532 physiologic functions of the retinal neurons and other cells. As mentioned, pan-VEGF agents exert 533 534 a potent anti-angiogenic action that exceeds the blockade of neoangiogenesis; for example, in vivo studies showed that non-selective VEGF inhibition causes capillary regression, robust and early 535 changes in ECs, pericytes, and basement membrane in the adult normal vasculature [108; 109], all 536 537 effects likely due to the block of the physiological functions of VEGF isoforms, besides the inhibition of the pathological effects, as described in the previous paragraphs. 538

Local side effects are possible for theoretically every drug used in ophthalmology, and include 539 toxicity related to the substance itself or due to the route of administration, such as in this case to 540 single or repeated intraocular injections. Among others, the most common local adverse effects of 541 anti-VEGF treatments comprise endophthalmitis, retinal detachment, intraocular pressure increase, 542 eye inflammation, hyperaemia and hemorrhage, which can occur with major or minor incidence 543 following treatment with any of the anti-VEGF agents [for comprehensive reviews see: 110-112]. 544 Instead, the main occurrence of some adverse effects in non-selective anti-VEGF agents might be 545 explained with their indiscriminate inhibition of all VEGF isoforms, including VEGF₁₂₁ and 546 VEGF₁₆₅b, which indeed seem to play a role mainly in physiological processes at vascular and 547 neuronal level. 548

549

551 9.1. Anti-VEGF drugs and atrophy of the retina

In both MARINA and ANCHOR trials on ranibizumab at 2 years, the increase in RPE 552 abnormalities was one of the most significant characteristic lesions associated to visual acuity loss 553 [as reviewed in 113]. Analogously, the number of ranibizumab injections was significantly 554 associated with the progression of RPE atrophy in wet AMD patients followed for a median of 16 555 months (range 3–36 months) [114]. It has been proposed that pan-VEGF blockade is responsible of 556 increasing geographic atrophy (GA) in AMD patients, a gradual complication characterized, among 557 others, by choriocapillaries and RPE atrophy, photoreceptors death, and leading to a progressive 558 visual loss [115; 116]. Monthly or PRN injection regimen with ranibizumab or bevacizumab led to 559 development of GA by 2 years in 18.3% of wet AMD subjects included in the CATT trials [117]; 560 an update of this study indicates that GA growth rate does not differ between eyes treated monthly 561 or PRN, but it may be accelerated by ranibizumab [118]. Moreover, the multicenter cohort SEVEN 562 563 UP study assessing long-term outcomes 7 to 8 years after initiation of intensive ranibizumab therapy in patients previously enrolled in the MARINA, ANCHOR and HORIZON trials, 564 565 evidenced that, although ranibizumab is efficacious in wet AMD, one third of subjects demonstrated visual benefits and another third had poor outcomes; more alarming, macular atrophy 566 was detected by fundus autofluorescence in 98% of all studied eyes, with the area of atrophy 567 mainly localized in the fovea and significantly correlated with a poor visual outcome [119]. 568 Interestingly, to our knowledge in literature there is only one report of rapid development of foveal 569 GA possibly related to a single injection of pegaptanib in one patient presenting an already 570 imbalanced foveal choroidal circulation due to AMD complicated by chronic serous drusenoid 571 pigment epithelium detachment (PED) [120], suggesting that selectively inhibiting VEGF₁₆₅ and 572 preserving other isoforms may avoid GA occurrence. 573

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- 576

577 9.2. Anti-VEGF drugs, RPE tears and other lesions

578 Other complications possibly occurring in wet AMD patients are fibrosis and formation of scars; in addition, since the approval of intravitreal pharmacotherapy, there has been a huge number of 579 reports of RPE tears developing after anti-VEGF injections, and thus a debate raised whether in 580 these cases anti-VEGF therapy is beneficial, or not, on the anatomical and visual outcomes. To cite 581 only a few of these evidence, RPE tears were diagnosed about 2 months after the first injection with 582 583 an anti-VEGF agent, and observed in 12–15% of all eyes treated for PED in wet AMD [121]. The SEVEN UP study on ranibizumab reported the absence of fibrotic lesions in almost 40% of the 584 examined eyes, although central fibrotic scars were demonstrated in approximately one third of the 585 586 retina, with significant repercussions on visual acuity [119]. Approximately 45% of eyes treated with either ranibizumab or bevacizumab and enrolled in the CATT study developed scar by 2 years 587 [122]. However, the majority of these RPE tear cases identified after ranibizumab, bevacizumab or 588 589 pegaptanib therapy were associated with a pre-existing complication, a baseline vascularized PED, which in most cases evolves into RPE tears [123; 124]. A recent study with an average follow-up of 590 591 42 months in patients with a diagnosis of RPE tear developed spontaneously or after anti-VEGF therapy, stated that the formation of atrophic or fibrotic disciform scars occurred equally in both 592 ranibizumab-treated and discontinued groups, and finally suggested that, in general, continuing anti-593 594 VEGF therapy is beneficial, reduces adverse outcomes and improves prognosis [125]. In support of this conclusion, in vitro and in vivo studies showed that fibroblast proliferation is stimulated by 595 VEGF and inhibited by administration of bevacizumab, which contrasts collagen deposition and 596 improves the outcomes after glaucoma surgery [126]. It has been suggested that VEGF isoforms 597 play distinct roles in scar formation, with VEGF₁₈₉ being mainly involved in fibrosis; for this 598 reason, pan-VEGF blockade may have a better anti-scarring potency than the selective VEGF₁₆₅ 599 inhibitor pegaptanib, which in vitro inhibits fibroblast growth only at the highest doses tested, and 600 whose benefits in post-operatory seem to be mediated mainly by inhibition of angiogenesis, but not 601 reduction of inflammation or collagen deposition [127]. 602

9.3. Anti-VEGF agents and the haemodynamics of eye vessels

604 Studies on the effects of anti-VEGF drugs on retrobulbar and retinal haemodynamics in wet AMD patients indicate that non-selective molecules may induce hypoperfusion. Specifically, 4 weeks 605 606 after a single injection of bevacizumab, the flow of all retrobulbar arteries - in particular the blood flow velocities (BFVs) in the central retinal (CRA), temporal posterior ciliary (TPCA) and 607 ophthalmic arteries (OA) - has been shown to be reduced [128]. A significant vasoconstriction of 608 the retinal arterioles lasting thirty days after each injection of ranibizumab was observed in wet 609 AMD patients [129]. A more recent study showed that ranibizumab leads to an early impairment of 610 the native choroidal and retinal vascular networks, but most of these effects are reversible after its 611 612 discontinuation; however, the study evidenced a significant correlation between the number of injections and percentage of changes in BFVs of CRA at month 6 [130]. The sole study on 613 pegaptanib, specifically on DME and BRVO patients, only shows a significant decrease of blood 614 615 flow velocity in the CRA after the third injection, possibly due to a cumulative effect for repeated treatments; no effects on retinal capillary blood flow, velocity or resistance index in the OA or 616 617 TPCA were evidenced in the small number of subjects examined [131]. However, further and larger studies are needed to confirm the pegaptanib's better profile than other anti-VEGF agents on the 618 haemodynamics of retrobulbar and retinal vessels. 619

620

621 10. Risk of inhibiting VEGF beyond the eye

Since their appearance in clinics, many observational studies, reviews and meta-analyses have been published on the systemic tolerability of ophthalmic anti-VEGF drugs and related adverse drug reactions (ADRs). Indeed, all the intraocular injected anti-VEGF agents can pass through the bloodretinal barrier and enter the systemic circulation, causing a decrease in VEGF plasma levels at various degrees, with several consequences. For instance, it was documented that intravitreally administered bevacizumab rapidly penetrates the rabbit retina before leaking into the blood circulation [132], and that in patients the intraocular injection of bevacizumab strongly decreases

the VEGF serum concentration, to the extent that after 1 month after the antibody treatment blood 629 630 VEGF is still 23% of baseline [133]. The circulating VEGF protects the vascular patency and integrity, and up-regulates NOS, thus a prolonged anti-VEGF treatment potentially increases the 631 risk of hypertension and thromboembolic events [134]. Relevantly, in comparison with healthy age-632 and sex-matched populations, neovascular AMD patients are elderly people presenting an increased 633 prevalence of hypertension, myocardial infarction, stroke, diabetes, and thus they may be more 634 susceptible to cardiovascular and cerebrovascular toxicities and prone to manifest ADRs [135-139]. 635 In particular, the most frequently documented comorbidities with wet AMD are hypertension and 636 other cardiovascular diseases, accounting 57.5% of cases [140]. After a 10-year period, people with 637 638 early-stage AMD have almost a 2-fold cumulative incidence of stroke than controls (4.08% vs. 2.14%) [141]. As well, DR subjects are more likely to have an increased risk for vascular events 639 [139]. 640

641 A recent review of some relevant clinical trials shows that the rates of serious thromboembolic events, such as stroke, heart attack and death, are similar for AMD patients treated with different 642 643 anti-VEGF agents. In particular, the authors state that in these subjects the arterial thrombotic risk appears sufficiently low, when compared with the natural incidence of thromboembolic events in 644 this category of elderly people, to be considered acceptable and counterbalanced by the advantage 645 of a visual improvement [142]; in few words, the risk of thrombotic events is seen as the worthy 646 price for ocular benefits. Similarly, a recent meta-analysis in DME patients evidences no significant 647 difference between anti-VEGF treated subjects and controls for arterial thromboembolic events; 648 however, the authors judge the quality of the evidence on these ADRs as moderate due to an 649 incomplete report of safety data, and the exclusion of high-risk participants (people with previous 650 cardiovascular events) in some studies [106]. Another systematic review of pre- and post-marketing 651 safety data on ranibizumab, pegaptanib and aflibercept, including 7,720 spontaneous reports from 652 the European database EudraVigilance, highlights an increased number of thromboembolic events 653 (0.8%–5%) and mortality (2.8%–4%) with anti-VEGF agents evidenced by post-marketing studies, 654

and suggests the need to properly evaluate the risk for such serious and long-term ADRs with 655 656 further studies [112]. Again, data from real life evidence relevant safety issues for some nonselective anti-VEGF agents; a comparative analysis of ADRs in the WHO database shows an 657 elevated disproportionality for cardiovascular events in patients treated with ranibizumab, in 658 particular myocardial infarction, cardiac failure congestive, and cerebrovascular accidents [111]. 659 This analysis was performed on 3,180 reports of worldwide pharmacovigilance from 2002 to 2012, 660 corresponding to 7,753 drug-reaction pairs concerning ranibizumab (5,130, 66%), bevacizumab 661 (2,069, 27%), and pegaptanib (554, 7%). Interestingly, although the number of reports on 662 pegaptanib were more limited in comparison with other agents, no relevant safety issues were 663 identified for this drug. In agreement, safety data from year 2 and 4 of the VISION trial previously 664 suggested no evidence of an increased risk of systemic adverse events associated with long-term 665 treatment with pegaptanib [143]. As AMD patients, DR population may require long-term anti-666 667 VEGF therapy, thus it is important to consider potential systemic effects in subgroups prone to vascular events when deciding between non-selective and selective agents [144]. In light of this 668 669 consideration, and being pegaptanib potentially safer than non-selective anti-VEGF agents, some authors have suggested to use pegaptanib as an initial therapy for DME, substituted with a pan-670 VEGF blocker in case the desired result is not achieved [145]. 671

VEGF regulates vascular permeability in various districts and it exerts neurotrophic and neuroprotective effects on blood-retinal and blood-brain barriers [146], thus it is conceivable that pan-VEGF suppression induced by intravitreal treatment may be deleterious also at cerebral level. According to the above mentioned spontaneous reports from the WHO database, a potential increased risk of cerebrovascular events associated with ranibizumab, especially with a more intensive treatment, was evidenced also by meta-analyses on five clinical trials with this drug (FOCUS, MARINA, ANCHOR, PIER, and SAILOR) [147; 148].

679 Clinical and experimental findings report that the use of anti-VEGF agents can result in neuronal 680 damage, often leading to pain [149]. Recent evidence have shown that both $VEGF_{165}a$ and VEGF₁₆₅b are neuroprotective, but they have pro- and anti-nociceptive effects, respectively [150]; the authors thus suggest that pain associated with anti-VEGF agents, and especially with molecules non-discriminating among the two 165 isoforms, is not fully attributable to a loss of a neuroprotective effect, but possibly also involves the modulation of nociception by VEGF-A isoforms.

Another organ dependent on VEGF and potentially exposed to injury from systemic absorption of 686 ophthalmic anti-VEGF drugs is the kidney. Hypertension and proteinuria were described soon as 687 adverse effects of systemic treatment with bevacizumab [151]. In general, anti-VEGF agents have 688 common adverse vascular effects attributable directly or indirectly to VEGF blockade, including 689 hypertension and renal vascular injury, usually manifested by proteinuria and thrombotic 690 microangiopathy [152]. The renal toxicity is likely due to the loss of VEGF functions in the 691 developed kidney, and to the close relationships existing among VEGF, NO, endothelin-1 and 692 693 angiotensin-II expression [153]. In particular, $VEGF_{121}$ is fundamental for renal function [154] and it has been shown to protect rats from kidney infarction in thrombotic microangiopathy through 694 695 maintaining NO production and/or preventing EC death [155]. Down-regulation of NO by anti-VEGF has been also implicated among the mechanisms underlying hypertension, besides 696 rarefaction of the microvasculature induced by anti-VEGF agents [156]. 697

698 Further studies are needed to better identify the main thromboembolic events related to the use of 699 anti-VEGF agents, in particular non-selective inhibitors, and their occurrence. However, the risk for some systemic ADRs may be increased in patients treated with anti-VEGF agents due to their 700 intrinsic characteristics; moreover, since most of the patients are aged 65 or older, age-related 701 702 physiologic changes, such as impairment of hepatic and renal function, may increase the odds for ADRs. To this criticism we should add potential comorbidities and polypharmacy, which are 703 common in the elderly and contribute to increase risk factors for cardio- and cerebro-vascular 704 705 events. In light of these evidence, pegaptanib maintenance strategy after a loading phase with pan-VEGF drugs may represent a safer therapeutic option in AMD patients with various comorbidities, 706

offering clinically meaningful benefits with a minimal systemic exposure to non-selective VEGF
 inhibition (and related potential side effects), reduced number of intravitreal injections required for
 treatment, and thus an improved cost/effectiveness profile [157].

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711 11. Tachyphylaxis: a relevant issue in anti-VEGF therapy

Anti-VEGF therapy is efficacious in the majority of patients; however, in the long-term, repeated 712 713 intravitreal injections of these molecules seem to be associated with a reduced bioefficacy. To this regard, still up to one-fourth of all treated patients, defined as no-responders, does not benefit from 714 intravitreal injections and visual acuity deteriorates even under treatment [158]. In general, long-715 716 term efficacy of a drug can be affected by tachyphylaxis, a phenomenon which is often confused with tolerance, both determining a reduced drug efficacy. When drugs are administered repeatedly 717 718 over a short period, tachyphylaxis can develop quite quickly and no response is observed following 719 a dosage increase, although the efficacy can be restored if the compound is suspended for a short while [159]. 720

Keane and collaborators [160] first speculated the possible implication of the tachyphylaxis
phenomenon following administration of ranibizumab for the treatment of neovascular AMD, and a
diminished therapeutic response due to tachyphylaxis was also indicated for intravitreal
bevacizumab [161; 162].

Although some cases of presumed tachyphylaxis may be ascribed to a poor or suboptimal response 725 to treatment [11], several mechanisms have been proposed to contribute to the diminished drug 726 response, such as the alteration of the neovascular membrane including increased fibrosis, chronic 727 changes in the vessel wall and in relevant neighboring structures as photoreceptors or RPE [159]. 728 The attenuated response after repeated administration may be also explained in terms of a raise in 729 other angiogenic signaling pathways which are aimed to compensate the blocked activity of VEGF; 730 for example, macrophages located within the choroidal neovascular tissue may respond to VEGF 731 inhibition by upregulating the production of VEGF itself [164]. As already reported for other 732

therapeutic humanized monoclonal antibodies, the formation of circulating neutralizing antibodies 733 734 may also take part to tachyphylaxis; indeed, neutralizing antibodies have been already documented against ranibizumab [163] and bevacizumab [164]. Clearly, this last aspect is in favor of pegaptanib 735 736 since aptamers are nonimmunogenic and are less likely to cause tolerability issues [86]. Genetic variants of the VEGF gene seem also to alter the response to anti-VEGF treatment [165], therefore, 737 it has been suggested that even minor differences in the binding properties might explain a 738 differential response to the various anti-VEGF therapeutics, offering the possibility of a response 739 even in patients who developed a tolerance to one drug [158]. Within this general context, clinical 740 evidence demonstrated that no-responders to either bevacizumab or ranibizumab benefit from a 741 switch to the other drug [158; 166; 167]. Moreover, a significant improvement in visual and 742 anatomical outcomes was also described after switching therapy to aflibercept in 34 eyes with 743 persistent subfoveal fluid formerly treated with ranibizumab [168]. More recently, switching to 744 745 pegaptanib monotherapy has been also documented to be strongly effective in those AMD patients who did not respond to ranibizumab or ranibizumab combined with photodynamic therapy [169]. 746 747 The authors ascribe the efficacy of pegaptanib mainly to its selectivity towards the VEGF₁₆₅ isoform and to the fact that it is immunologically lenient. Moreover, since according to Pfizer's 748 internal data pegaptanib has a weak binding ability towards PDGF, they also assert that this feature 749 can additionally contribute to pegaptanib activity [169]. To this regard, several lines of evidence 750 751 suggest that the response of blood vessels to anti-VEGF therapy is influenced by vessel maturation which involves pericytes [see 170]. The recruitment of pericytes on endothelial cells is mediated by 752 PDGF-B signaling via PDGF receptor-type β (PDGFR- β). Indeed, transgenic mice lacking PDGF-B 753 754 and PDGFR- β are characterized by abnormal vessel stabilization and maturation [171], and inhibition of PDGF cascade increases EC sensitivity to anti-VEGF agents [172]. Therefore, a 755 combined therapy targeting PDGF-B and VEGF-A may represent a more effective pharmacological 756 approach to face neovascular AMD and possibly to avoid tachyphylaxis challenge. Within this 757 context, pegaptanib itself, although provided with a weak binding activity towards PDGF, may 758

further benefit of this combination therapy strategy not only in inhibiting new vessel growth butalso vessel regression, as documented by Jo and collaborators in mice [170].

761

762 12. Future perspectives in eye neovascularization: oligonucleotide-based interventions to 763 modulate VEGF pathway

Anti-VEGF therapy has certainly represented a breakthrough intervention to counteract pathological angiogenesis, although it should be taken into account that these agents usually help to delay further vision loss rather than improve it. This latter aspect underscores the need to identify novel approaches, also considering that novel VEGF-dependent [173] as well VEGF-independent pathways [174] may be involved, the latter may contribute to better explain the resistance, observed in some patients, to the anti-VEGF treatment itself, as previously mentioned.

Within this general context, as recently reviewed [175], efforts have also been directed to develop 770 771 advanced drug-delivery devices to reduce treatment burden, such as using the encapsulated cell technology (designed to deliver active compounds directly into the vitreous following trans-scleral 772 773 implantation) or utilizing colloidal drug carriers (consisting of suspensions of microparticles/nanoparticles or liposomes), and also taking into account the need to develop eye 774 drop or oral formulations to improve patient compliance. Moreover, considering that, as recently 775 reviewed [175-177], single nucleotide polymorphisms can play a predictive role in AMD 776 progression or, in general, in the response to treatment, pharmacogenomics studies may help in the 777 choice of a more appropriate therapy. 778

The present paragraph specifically deals with novel oligonucleotide-based interventions in the eye mainly targeting, directly or indirectly, VEGF mRNA/protein/receptors or to be used in combination with the currently used anti-VEGF agents (Table 1; Figure 2), leaving the reader to other recent publications for differential pharmacological approaches or biological targets [i.e. 22; 172; 178; 179].

NAME	TYPE OF MOLECULE	BIOLOGICAL TARGET	CLINICAL STAGE	
RANIBIZUMAB	Recombinant humanized monoclonal antibody fragment	ALL VEGF-A		
BEVACIZUMAB	Recombinant humanized monoclonal antibody		CURRENT DRUGS	
AFLIBERCEPT	Fusion protein	ALL VEGF-A & VEGF-B ISOFORMS		
PEGAPTANIB	RNA aptamer	+ PLGF VEGF-A ₁₆₅ ONLY		
BEVASIRANIB	siRNA	VEGF-A mRNA	Phase III	
SIRNA-027	siRNA	VEGFR-1 mRNA	Phase I	
PF-04523655	siRNA	DDIT4 mRNA	Phase II	
ALN-VSP02	Dual -siRNA	VEGF-A and KSP mRNAs	Phase I	
E10030	DNA aptamer	PDGF-BB	Phase III	
ARC1905	RNA aptamer	C5 complement	Phase I	
Table 1. Current drugs and novel oligonucleotide-based molecules to face ocularneovascularization. The drugs currently used (also off-label) in therapy or the oligonucleotide-				

neovascularization. The drugs currently used (also off-label) in therapy or the oligonucleotidebased molecules under clinical trials and potentially useful to counteract ocular neovascularization
in different eye diseases are reported. When not specifically indicated, the biological target is
referred to the protein (for more details see text, paragraph 12). PLGF: Placental Growth Factor.



Figure 2. Oligonucleotide-based interventions targeting VEGF and other molecules involved in neovascularization. Aptamers (in green color), siRNAs (in blue color) and their targets, expressed in endothelial cells and pericytes, are depicted. These under-trials molecules may be used in combination with the currently used anti-VEGF agents (for more details see text, paragraph 12).

796 12.1. Small interfering RNAs (siRNAs)

Sometimes also named short interfering RNAs or silencing RNAs, siRNAs are double-stranded RNA molecules, 20-25 base pairs in length, capable of operating gene silencing at posttranscriptional level. Indeed, their catalytic nature allows for one siRNA to guide the cleavage of thousands of mRNAs, resulting in effective gene silencing with no translation of the related proteins. These molecules hold a great potential since can be easily designed and are characterized by high efficacy and specificity [180].

Bevasiranib is a double-stranded RNA of 21 nucleotides in length directed to VEGF-A mRNA. 803 804 Preclinical studies documented bevasiranib efficacy in inhibiting neovascularization in both mice and nonhuman primate models [181; 182]. Moreover, in rabbits, an extensive uptake into the retina 805 was observed following intravitreal injections of a single dose of either 0.5 mg/eye or 2.0 mg/eye 806 of ³H-bevasiranib [183]. Promising results for the treatment of AMD and DME originated from 807 Phase I and II clinical trials, also showing that bevasiranib effects were not manifest until 6 weeks 808 after treatment. However, in March 2009 OPKO Health Pharmaceuticals decided to terminate its 809 Phase III clinical study for the treatment of wet AMD. 810

Sirna-027 (also named AGN211745) is a chemical-modified siRNA that targets a conserved region 811 of VEGFR-1 mRNA. In mice, it was shown to reduce pathological angiogenesis in a laser-induced 812 CNV model [184], and it was proven to be safe and effective in nonclinical safety studies [185]. 813 Based on its preclinical activity and tolerability, a Phase I study was subsequently conducted 814 815 concluding that a single intravitreal dose of Sirna-027, up to 1600 µg/eye, is well tolerated in patients with CNV resulting from neovascular AMD [185]. However, no additional trials are 816 currently running on this molecule (search up to July 16th 2015 at: ClinicalTrials.gov and EU 817 Clinical Trials Register). 818

PF-04523655 is a 19-nucleotide, O-methyl stabilized, siRNA that specifically targets the DNA-819 damage-inducible transcript 4 (DDIT4) genes, also known as REDD1 or RTP-801, indirectly 820 leading to a decrease in VEGF-A production [22]. RTP801 is a hypoxia-inducible factor 1-821 responsive gene, which displays strong hypoxia-dependent up-regulation both in vitro and in vivo. 822 Indeed, in diabetic rats its expression usually increases by up to 70% in RPE/choroid and it is 823 reduced by the administration of PF-04523655 [186]. Furthermore, RTP801 knockout mice show a 824 significant reduction in retinal neovascularization in a model of retinopathy of prematurity [187]. A 825 Phase I multicentre study has been completed on AMD in 2010, showing that a single intravitreal 826 injection of 50 to 3000 µg of PF-045237655 is generally safe and well tolerated over 24 months 827 [188]. A Phase II interventional clinical trial (the DEGAS study) was subsequently conducted to 828

evaluate the safety and efficacy of three doses of PF-04523655 (0.4mg, 1mg and 3mg) for the 829 treatment of DME in comparison with focal/grid laser. In general, PF-04523655 was proven to be 830 safe and well-tolerated, with few adverse events judged treatment-dependent. All the three dose 831 levels of the siRNA continued to improve visual acuity from baseline through month 12 in patients 832 with DME. Moreover, at month 12, a trend for a greater improvement in visual acuity from baseline 833 was observed in the 3mg PF-04523655 group with respect to the laser photocoagulation one. 834 Unfortunately, the study was terminated early at month 12 based on the high patient discontinuation 835 rate, mainly due to lack of efficacy [189]. Two Phase II studies have been additionally run to 836 investigate the benefits of a combined therapy of this siRNA with ranibizumab in wet AMD and in 837 838 DME, respectively. The first clinical trial, the MONET study, which assessed the efficacy of different dosing paradigms of PF-04523655 versus ranibizumab (0.5mg) showed that, in subjects 839 with neovascular AMD, the combined therapy leads to an average gain in visual acuity that is more 840 841 elevated than with ranibizumab monotherapy, with no safety concerns identified [190]. In relation to the second clinical trial, the dose escalation study and evaluation of PF-04523655 with/without 842 843 ranibizumab (MATISSE study) carried in DME patients, although already completed, no results have been posted thus far (search up to July 16th 2015 at: ClinicalTrials.gov and EU Clinical Trials 844 Register). 845

Although no specific studies have been yet performed on ocular diseases, it is worth to mention 846 ALN-VSP02 since it is a lipid nanoparticle-formulated dual-targets drug candidate. It contains 2 847 different siRNAs, chemically modified (with 2'-O-methyl groups to minimize immunostimulation) 848 in a 1:1 molar ratio, directed to two different pathways: VEGF-A and kinesin spindle protein (KSP). 849 KSP is a member of the kinesin superfamily of microtubule-based motor proteins whose inhibition 850 determines cycle arrest at mitosis, finally leading to cell death [191]. To assess the activity and 851 safety of intravenous ALN-VSP02 in humans, a Phase I trial was initiated in patients with advanced 852 solid tumors with liver involvement. On the whole, ALN-VSP02 was well tolerated, with an 853 adverse event profile favorable in comparison with chemotherapy and with other orally or 854

intravenously targeted therapies administered in oncology. At molecular level, ALN-VSP02
counteracts the translation of both VEGF-A and KSP proteins, which results in growth inhibition of
tumor cells and complete regression of liver metastases in endometrial cancer [192].

No Phase II clinical trials are currently running (search up to July 16th 2015 at: ClinicalTrials.gov
and EU Clinical Trials Register).

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861 **12.2. Aptamers**

Aptamers represent a step forward with respect to siRNAs; indeed, although still oligonucleotidebased molecules, they do not necessarily require transfection and their stability/bioavailability can be largely improved since the *in vitro* production allows to manipulate their kinetic properties. In particular, 2'F or 2'OMe modifications as well as the presence of conformationally restricted nucleotides confer resistance to nucleases, while PEG-conjugation limits renal filtration [178].

As previously said, pegaptanib has been the first aptamer entered in therapy. Besides pegaptanib, later on other aptamers have been synthesized to target VEGF. For example Nonaka and collaborators identified a DNA aptamer (named Vap7) able to bind both VEGF₁₂₁ and VEGF₁₆₅ isoforms through the RBD region. However, they subsequently optimized this aptamer for a diagnostic potential application as biosensor for VEGF detection [193; 194].

As cited earlier, current theories suggest that blocking simultaneously VEGF-A and PDGF results in a more effective inhibition of neovascularization [170; 195]. PDGF is a family of proteins comprising four different polypeptides (PDGF A-D) which can combine either as homodimers or heterodimers. The homodimer PDGF-BB has been involved in pericyte recruitment, maturation and survival through the binding on its specific receptor, namely the PDGFR- β , on pericytes [196]. Indeed, PDGF inhibition causes a loss of pericytes, leaving ECs vulnerable to anti-VEGF therapy, an effect that can also help avoid tachyphylaxis.

On these premises it has been designed E10030, a DNA aptamer specifically targeting PDGF-BB.
This drug candidate is a PEGylated, 2'F- and 2'OMe-modified aptamer of 29 nucleotides. E10030

has been successfully assessed in association with anti-VEGF molecules in inducing neovascular 881 regression [197]. A Phase I trial was performed to determine the combined effect of E10030 and 882 ranibizumab on subjects with subfoveal CNV, showing a significant vascular regression and a 883 superior efficacy in comparison to ranibizumab monotherapy after 12 weeks of treatment [198]. 884 Similar results were obtained in a Phase II trial where a greater efficacy was observed, following 6 885 monthly injections, especially with the higher dose of E10030 in combination with ranimizumab 886 with respect to ranibizumab alone [179]. Phase III studies are currently running/recruiting to 887 evaluate the safety and efficacy of E10030 in combination with ranibizumab or bevacizumab or 888 aflibercept in comparison to the respective anti-VEGF alone (search up to July 16th 2015 at: 889 890 ClinicalTrials.gov and EU Clinical Trials Register).

Still remaining in the context of combined therapies, another promising aptamer to be associated to 891 anti-VEGF agents is ARC1905 which targets C5 complement. C5 is a serum glycoprotein that is 892 893 cleaved in two fragments, C5a (active) and C5b, during complement activation. C5a is chemotactic and plays a key role in stimulating neutrophil-endothelial adhesion [199]. The aptamer antagonizes 894 895 C5 cleavage thus preventing complement activation. A Phase I trial has been completed on the safety, tolerability, and pharmacokinetic profile of multiple doses of ARC1905 in combination with 896 ranibizumab in subjects with subfoveal CNV secondary to AMD, however up to now no results 897 have been posted (search up to July 12th 2015 at: ClinicalTrials.gov and EU Clinical Trials 898 Register). 899

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901 13. Conclusions

The study of anti-VEGF strategies in the treatment of ocular diseases linked to abnormal vascularization raises several questions relevant for both the understanding of the biology of the VEGF system and the rational design of the interventions directed to counteract VEGF. The observations on the role of VEGF in AMD and DR also underscore the need to increase the knowledge of the molecular bases of ocular diseases due to an impaired angiogenesis control.

The discovery of several isoforms of VEGF having different biological activity has revealed a 907 908 previously unforeseen biological complexity which needs to be addressed when studying the clinical activity of currently available anti-VEGF drugs and while exploring new molecules active 909 on this target. In particular, an answer has to be provided to the question whether for a full 910 antiangiogenic activity is better to act against all the existing VEGF isoforms or to selectively block 911 few or one of them. The fact that some of the VEGF isoforms have antiangiogenic and 912 neuroprotective action suggests that the VEGF system is physiologically balanced and that in 913 presence of an angiogenic process it would be preferable in the long run to hit the angiogenic 914 isoforms leaving unaffected those isoforms having a different biological activity, the neutralization 915 916 of which may be responsible for a derangement of vasculature control as well as of tissue reparative processes. When matching the molecular profiles and comparing the clinical activities of the 917 available drugs, in particular of the pan-VEGF antibodies and of the aptamers, such as pegaptanib, 918 919 displaying a preferential affinity toward the VEGF₁₆₅a isoform, one is tempted to speculate that a selective action is sufficient to sustain in the time the antiangiogenic effect, while to quickly 920 921 develop a full block of the angiogenic process an action directed toward all the isoforms is more effective. The preclinical studies with some of the newer and more selective siRNAs will help to 922 clarify this point. In the meantime, the new molecular advances may be used to better tailor the 923 existing therapies and to explain some of the cases of therapy resistant patients and to understand 924 the possible mechanisms underlying their side effects. 925

The discovery that, in addition to VEGF, other factors, such as PDGF, may participate to the control of the angiogenic process, also raises the question whether it is necessary to simultaneously act also on these other molecules, or VEGF inhibition is sufficient for an effective therapy. On the other hand it is possible that different patients, so far included in the same diagnostic category, indeed have different molecular dysfunctions underlying their disease requiring a more refined profiling of the VEGF isoforms and of the other factors regulating angiogenesis in that particular patient/disease.

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936 937	Bibliography				
938 939 940	1.	Bressler NM. Age-related macular degeneration is the leading cause of blindness JAMA 2004;291(15):1900-1.			
941 942	2.	Song SJ, Wong TY. Current concepts in diabetic retinopathy. Diabetes Metab J 2014;38(6):416-25.			
943 944 945	3.	Jager RD, Mieler WF, Miller JW. Age related macular degeneration. N Engl J Med 2008;358: 2606–17.			
946 947	4.	Klein R. Retinopathy in a population-based study. Trans Am Ophthalmol Soc 1992; 90:561–94.			
948 949	5.	Cheung N, Mitchell P, Wong TY. Diabetic retinopathy. Lancet 2010; 376(9735):124-36.			
950 951 952 953	6.	Friedman DS, Wilson MR, Liebmann JM, Fechtner RD, Weinreb RN. An evidence-based assessment of risk factors for the progression of ocular hypertension and glaucoma. Am J Ophthalmol 2004;138(3 Suppl):S19-31.			
954 955 956	7.	Das A, McGuire PG. Retinal and choroidal angiogenesis: pathophysiology and strategies for inhibition. Prog Retin Eye Res 2003;22(6):721-48.			
957 958 959 960	8.	Brylla E, Tscheudschilsuren G, Santos AN, Nieber K, Spanel-Borowski K, Aust G. Differences between retinal and choroidal microvascular endothelial cells (MVECs) under normal and hypoxic conditions. Exp Eye Res 2003;77(5):527-35.			
961 962 963	9.	Kvanta A. Ocular angiogenesis: the role of growth factors. Acta Ophthalmol Scand 2006;84(3):282-8.			
964 965 966	10.	Cheung N, Wong IY, Wong TY. Ocular anti-VEGF therapy for diabetic retinopathy: overview of clinical efficacy and evolving applications. Diabetes Care 2014;37(4):900-5.			
967 968 969 970	11.	Amoaku WM, Chakravarthy U, Gale R, Gavin M, Ghanchi F, Gibson J, Harding S, Johnston RL, Kelly S, Lotery A, Mahmood S, Menon G, Sivaprasad S, Talks J, Tufail A, Yang Y. Defining response to anti-VEGF therapies in neovascular AMD. Eye (Lond) 2015;29(6):721-731.			
971 972 973	12.	Giet MV, Henkel C, Schuchardt M, Tölle M. Anti-VEGF Drugs in Eye Diseases: Local Therapy with Potential Systemic Effects. Curr Pharm Des. 2015 Feb 25.			

974 975 976	13.	Lowe J, Araujo J, Yang J, Reich M, Oldendorp A, Shiu V, Quarmby V, Lowman H, Lien S, Gaudreault J, Maia M. Ranibizumab inhibits multiple forms of biologically active vascular endothelial growth factor in vitro and in vivo. Exp Eye Res 2007;85(4):425-30.
977 978 979	14.	FDA Consum. 2006 Sep-Oct;40(5):6. Treatment for wet macular degeneration.
980 981 982 983 984	15.	Schmidt-Erfurth U, Chong V, Loewenstein A, Larsen M, Souied E, Schlingemann R, Eldem B, Monés J, Richard G, Bandello F; European Society of Retina Specialists. Guidelines for the management of neovascular age-related macular degeneration by the European Society of Retina Specialists (EURETINA). Br J Ophthalmol 2014;98(9):1144-67.
985 986 987	16.	Schmucker C, Antes G, Lelgemann M. Position paper: the need for head-to-head studies comparing Avastin versus Lucentis. Surv Ophthalmol 2009;54(6):705-7.
988 989 990 991 992	17.	Nepomuceno AB, Takaki E, Paes de Almeida FP, Peroni R, Cardillo JA, Siqueira RC, Scott IU, Messias A, Jorge R. A prospective randomized trial of intravitreal bevacizumab versus ranibizumab for the management of diabetic macular edema. Am J Ophthalmol 2013;156(3):502-10.e2.
993 994 995	18.	Silver J. Drugs for macular degeneration, price discrimination, and Medicare's responsibility not to overpay. JAMA 2014;312(1):23-4. Erratum in: JAMA 2014;312(19):2045.
996 997 998	19.	Aflibercept: AVE 0005, AVE 005, AVE0005, VEGF Trap - Regeneron, VEGF Trap (R1R2), VEGF Trap-Eye. Drugs R D. 2008;9(4):261-9.
999 1000 1001	20.	Dixon JA, Oliver SC, Olson JL, Mandava N. VEGF Trap-Eye for the treatment of neovascular age-related macular degeneration. Expert Opin Investig Drugs 2009;18(10):1573-80.
1002 1003 1004 1005	21.	Stefanini FR, Badaró E, Falabella P, Koss M, Farah ME, Maia M. Anti-VEGF for the management of diabetic macular edema. J Immunol Res 2014;632307. doi: 10.1155/2014/632307.
1006 1007 1008	22.	Nowak W, Żarowska A, Szul-Pietrzak E, Misiuk-Hojło M. System and measurement method for binocular pupillometry to study pupil size variability. Biomed Eng Online 2014;13:69.
1009 1010 1011	23.	Ng EW, Shima DT, Calias P, Cunningham ET Jr, Guyer DR, Adamis AP. Pegaptanib, a targeted anti-VEGF aptamer for ocular vascular disease. Nat Rev Drug Discov 2006;5(2):123-32.
1012 1013 1014 1015	24.	Houck KA, Ferrara N, Winer J, Cachianes G, Li B, Leung DW. The vascular endothelial growth factor family: identification of a fourth molecular species and characterization of alternative splicing of RNA. Mol Endocrinol 1991;5(12):1806-14.

- 1016 25. Tischer E, Mitchell R, Hartman T, Silva M, Gospodarowicz D, Fiddes JC, Abraham JA. The
 1017 human gene for vascular endothelial growth factor. Multiple protein forms are encoded through
 1018 alternative exon splicing. J Biol Chem 1991;266(18):11947-54.
- 1019

- 1020 26. Ferrara N, Gerber HP, LeCouter J. The biology of VEGF and its receptors. Nat Med
 1021 2003;9(6):669-76.
- 1023 27. Bates DO, Cui TG, Doughty JM, Winkler M, Sugiono M, Shields JD, Peat D, Gillatt D, Harper
 1024 SJ. VEGF165b, an inhibitory splice variant of vascular endothelial growth factor, is down1025 regulated in renal cell carcinoma. Cancer Res 2002;62(14):4123-31.
- 1026

1031

- 1027 28. Usui T, Ishida S, Yamashiro K, Kaji Y, Poulaki V, Moore J, Moore T, Amano S, Horikawa Y,
 1028 Dartt D, Golding M, Shima DT, Adamis AP. VEGF164(165) as the pathological isoform:
 1029 differential leukocyte and endothelial responses through VEGFR1 and VEGFR2. Invest
 1030 Ophthalmol Vis Sci 2004;45(2):368-74.
- 1032 29. Muthusamy A, Lin CM, Shanmugam S, Lindner HM, Abcouwer SF, Antonetti DA. Ischemia 1033 reperfusion injury induces occludin phosphorylation/ubiquitination and retinal vascular
 1034 permeability in a VEGFR-2-dependent manner. J Cereb Blood Flow Metab 2014;34(3):522-31.
- 30. Soker S, Takashima S, Miao HQ, Neufeld G, Klagsbrun M. Neuropilin-1 is expressed by
 endothelial and tumor cells as an isoform-specific receptor for vascular endothelial growth
 factor. Cell 1998;92(6):735-45.
- 1039

1042

- **31.** Harper SJ, Bates DO. VEGF-A splicing: the key to anti-angiogenic therapeutics? Nat Rev Cancer 2008;8(11):880-7.
- 32. Kawamura H, Li X, Harper SJ, Bates DO, Claesson-Welsh L. Vascular endothelial growth factor
 (VEGF)-A165b is a weak in vitro agonist for VEGF receptor-2 due to lack of coreceptor binding
 and deficient regulation of kinase activity. Cancer Res 2008;68(12):4683-92.
- 1046
 1047 **33.** Djordjevic S, Driscoll PC. Targeting VEGFsignalling via the neuropilin co-receptor. Drug Discov Today 2013;18(9-10):447-55.
 - 1049
 - **34.** Tolentino M. Systemic and ocular safety of intravitreal anti-VEGF therapies for ocular neovascular disease. Surv Ophthalmol 2011; 56: 95–113.
 - 1052
 - 1053 35. Lee S, Chen TT, Barber CL, Jordan MC, Murdock J, Desai S, Ferrara N, Nagy A, Roos KP,
 1054 Iruela-Arispe ML. Autocrine VEGF signaling is required for vascular homeostasis. Cell
 1055 2007;130(4):691-703.
 - 1056
 - 36. Baffert F, Le T, Sennino B, Thurston G, Kuo CJ, Hu-Lowe D, McDonald DM. Cellular changes
 in normal blood capillaries undergoing regression after inhibition of VEGF signaling. Am J
 Physiol Heart Circ Physiol 2006;290(2):H547-59.

- 37. Vempati P, Popel AS, Mac Gabhann F. Extracellular regulation of VEGF: isoforms, proteolysis,
 and vascular patterning. Cytokine Growth Factor Rev 2014;25(1):1-19.
- 1062

1070

1075

1079

1082

1086

1089

- **38.** Carmeliet P, Collen D. Role of vascular endothelial growth factor and vascular endothelial growth factor receptors in vascular development. Curr Top Microbiol Immunol 1999;237:133-58.
- 39. Stalmans I, Ng YS, Rohan R, Fruttiger M, Bouché A, Yuce A, Fujisawa H, Hermans B, Shani
 M, Jansen S, Hicklin D, Anderson DJ, Gardiner T, Hammes HP, Moons L, Dewerchin M, Collen
 D, Carmeliet P, D'Amore PA. Arteriolar and venular patterning in retinas of mice selectively
 expressing VEGF isoforms. J Clin Invest 2002;109(3):327-36.
- 40. Ishida S, Usui T, Yamashiro K, Kaji Y, Amano S, Ogura Y, Hida T, Oguchi Y, Ambati J, Miller
 JW, Gragoudas ES, Ng YS, D'Amore PA, Shima DT, Adamis AP. VEGF164-mediated
 inflammation is required for pathological, but not physiological, ischemia-induced retinal
 neovascularization. J Exp Med 2003;198(3):483-9.
- 41. Ruhrberg C, Gerhardt H, Golding M, Watson R, Ioannidou S, Fujisawa H, Betsholtz C, Shima DT. Spatially restricted patterning cues provided by heparin-binding VEGF-A control blood vessel branching morphogenesis. Genes Dev 2002;16(20):2684-98.
- 42. Ruhrberg C. Growing and shaping the vasculartree: multiple roles for VEGF. Bioessays
 2003;25(11):1052-60.
- 43. Gerhardt H, Golding M, Fruttiger M, Ruhrberg C, Lundkvist A, Abramsson A, Jeltsch M,
 Mitchell C, Alitalo K, Shima D, Betsholtz C. VEGF guides angiogenic sprouting utilizing
 endothelial tip cell filopodia. J Cell Biol 2003;161(6):1163-77.
- 44. Gerhardt H. VEGF and endothelial guidance in angiogenic sprouting. Organogenesis
 2008;4(4):241-6.
- 45. Saint-Geniez M, Maldonado AE, D'Amore PA. VEGF expression and receptor activation in the choroid during development and in the adult. Invest Ophthalmol Vis Sci 2006;47(7):3135-42.
- 46. Saint-Geniez M, Kurihara T, Sekiyama E, Maldonado AE, D'Amore PA. An essential role for
 RPE-derived soluble VEGF in the maintenance of the choriocapillaris. Proc Natl AcadSci U S A
 2009;106(44):18751-6.
- 1096
- 47. Nagineni CN, Kommineni VK, William A, Detrick B, Hooks JJ. Regulation of VEGF expression
 in human retinal cells by cytokines: implications for the role of inflammation in age-related
 macular degeneration. J Cell Physiol 2012;227(1):116-26.
- 1100
- 48. Keyt BA, Berleau LT, Nguyen HV, Chen H, Heinsohn H, Vandlen R, Ferrara N. The carboxylterminal domain (111-165) of vascular endothelial growth factor is critical for its mitogenic
 potency. J Biol Chem. 1996;271(13):7788-9.

49. Whitaker GB, Limberg BJ, Rosenbaum JS. Vascular endothelial growth factor receptor-2 and 1104 neuropilin-1 form a receptor complex that is responsible for the differential signaling potency of 1105 VEGF(165) and VEGF(121). J Biol Chem 2001;276(27):25520-31. 1106 1107 50. Storkebaum E, Lambrechts D, Carmeliet P. VEGF: once regarded as a specific angiogenic 1108 1109 factor, now implicated in neuroprotection. Bioessays 2004;26(9):943-54. 1110 **51.** Greenberg DA, Jin K. From angiogenesis to neuropathology. Nature 2005;438(7070):954-9. 1111 1112 52. Lambrechts D, Carmeliet P. VEGF at the neurovascular interface: therapeutic implications for 1113 motor neuron disease. Biochim Biophys Acta 2006;1762(11-12):1109-21. 1114 1115 53. Jin KL, Mao XO, Greenberg DA. Vascular endothelial growth factor: direct neuroprotective 1116 effect in in vitro ischemia. Proc Natl Acad Sci U S A 2000;97(18):10242-7. 1117 1118 54. Rosenstein JM, Mani N, Khaibullina A, Krum JM. Neurotrophic effects of vascular endothelial 1119 growth factor on organotypic cortical explants and primary cortical neurons. J Neurosci 1120 2003;23(35):11036-44. 1121 1122 1123 55. Zachary I. Neuroprotective role of vascular endothelial growth factor: signalling mechanisms, biological function, and therapeutic potential. Neurosignals 2005;14(5):207-21. 1124 1125 1126 56. Sun FY, Guo X. Molecular and cellular mechanisms of neuroprotection by vascular endothelial 1127 growth factor. J Neurosci Res 2005;79(1-2):180-4. 1128 1129 57. Nicoletti JN, Shah SK, McCloskey DP, Goodman JH, Elkady A, Atassi H, Hylton D, Rudge JS, Scharfman HE, Croll SD. Vascular endothelial growth factor is up-regulated after status 1130 1131 epilepticus and protects against seizure-induced neuronal loss in hippocampus. Neuroscience 2008;151(1):232-41. 1132 1133 58. Silverman WF, Krum JM, Mani N, Rosenstein JM. Vascular, glial and neuronal effects of 1134 1135 vascular endothelial growth factor in mesencephalic explantcultures. Neuroscience 1999;90:1529-1541. 1136 1137 59. Nishijima K, Ng YS, Zhong L, Bradley J, Schubert W, Jo N, Akita J, Samuelsson SJ, Robinson 1138 GS, Adamis AP, Shima DT. Vascular endothelial growth factor-A is a survival factor for retinal 1139 neurons and a critical neuroprotectant during the adaptive response to ischemic injury. Am J 1140 Patholm 2007;171(1):53-67. 1141 1142 1143 60. Singh S, Evans TW. Nitric oxide, the biological mediator of the decade: fact or fiction? Eur 1144 Respir J 1997;10(3):699-707. 1145 61. Martínez-Vila E, Sieira PI. Current status and perspectives of neuroprotection in ischemic stroke 1146 treatment. Cerebrovasc Dis 2001;11 Suppl 1:60-70. 1147

- 62. Foxton RH, Finkelstein A, Vijay S, Dahlmann-Noor A, Khaw PT, Morgan JE, Shima DT, Ng
 YS. VEGF-A is necessary and sufficient for retinal neuroprotection in models of experimental
 glaucoma. Am J Pathol 2013;182(4):1379-90.
- 1151

1159

1162

1166

1169

- 63. Perrin RM, Konopatskaya O, Qiu Y, Harper S, Bates DO, Churchill AJ. Diabetic retinopathy is
 associated with a switch in splicing from anti- to pro-angiogenic isoforms of vascular endothelial
 growth factor. Diabetologia 2005;48(11):2422-7.
- 64. Hua J, Spee C, Kase S, Rennel ES, Magnussen AL, Qiu Y, Varey A, Dhayade S, Churchill AJ,
 Harper SJ, Bates DO, Hinton DR. Recombinant human VEGF165b inhibits experimental
 choroidal neovascularization. Invest Ophthalmol Vis Sci 2010;51(8):4282-8.
- **65.** Zhao Z, Chen Y, Wang J, Sternberg P, Freeman ML, Grossniklaus HE, Cai J. Age-related
 retinopathy in NRF2-deficient mice. PLoS One 2011;6(4):e19456.
- 66. Ehlken C, Rennel ES, Michels D, Grundel B, Pielen A, Junker B, Stahl A, Hansen LL, Feltgen N, Agostini HT, Martin G. Levels of VEGF but not VEGF(165b) are increased in the vitreous of patients with retinal vein occlusion. Am J Ophthalmol 2011;152(2):298-303.e1.
- 67. Baba T, Bikbova G, Kitahashi M, Yokouchi H, Oshitari T, Yamamoto S. Level of vascular
 endothelial growth factor 165b in human aqueous humor. Curr Eye Res 2014;39(8):830-6.
- **68.** Konopatskaya O, Churchill AJ, Harper SJ, Bates DO, Gardiner TA. VEGF165b, an endogenous
 C-terminal splice variant of VEGF, inhibits retinal neovascularization in mice. Mol Vis 2006;12:626-32.
- 1173

- 69. Magnussen AL, Rennel ES, Hua J, Bevan HS, Beazley Long N, Lehrling C, Gammons M,
 FloegeJ, Harper SJ, Agostini HT, Bates DO, Churchill AJ. VEGF-A165b is cytoprotective and
 antiangiogenic in the retina. Invest Ophthalmol Vis Sci 2010;51(8):4273-81.
- 1177
 1178 70. Rennel ES, Waine E, Guan H, Schüler Y, Leenders W, Woolard J, Sugiono M, Gillatt D, Kleinerman E, Bates D, Harper S. The endogenous anti-angiogenic VEGF isoform, VEGF165b
 - Kleinerman E, Bates D, Harper S. The endogenous anti-angiogenic VEGF isoform, VEGF165b
 inhibits human tumour growth in mice. Br J Cancer 2008;98(7):1250-7.
 - 71. Rennel ES, Hamdollah-Zadeh MA, Wheatley ER, Magnussen A, Schüler Y, Kelly SP, Finucane
 C, Ellison D, Cebe-Suarez S, Ballmer-Hofer K, Mather S, Stewart L, Bates DO, Harper SJ.
 Recombinant human VEGF165b protein is an effective anti-cancer agent in mice. Eur J Cancer
 2008;44(13):1883-94.
 - 1187 72. Woolard J, Wang WY, Bevan HS, Qiu Y, Morbidelli L, Pritchard-Jones RO, Cui TG,
 1188 SugionoM, Waine E, Perrin R, Foster R, Digby-Bell J, Shields JD, Whittles CE, Mushens RE,
 1189 Gillatt DA, Ziche M, Harper SJ, Bates DO. VEGF165b, an inhibitory vascular endothelial
 1190 growth factor splice variant: mechanism of action, in vivo effect on angiogenesis and
 1191 endogenous protein expression. Cancer Res 2004;64(21):7822-35.

- 73. Beazley-Long N, Hua J, Jehle T, Hulse RP, Dersch R, Lehrling C, Bevan H, Qiu Y, Lagrèze
 WA, Wynick D, Churchill AJ, Kehoe P, Harper SJ, Bates DO, Donaldson LF. VEGF-A165b is
 an endogenous neuroprotective splice isoform of vascular endothelial growth factor A in vivo
 and in vitro. Am J Pathol 2013;183(3):918-29.
- 1196

1202

1206

1209

1212

- 1197 74. Jellinek D, Green LS, Bell C, Janjić N. Inhibition of receptor binding by high-affinity RNA
 1198 ligands to vascular endothelial growth factor. Biochemistry 1994;33(34):10450-6.
- Tuerk C, Gold L. Systematic evolution of ligands by exponential enrichment: RNA ligands to
 bacteriophage T4 DNA polymerase. Science 1990;249(4968):505-10.
- 76. Senger DR, Galli SJ, Dvorak AM, Perruzzi CA, Harvey VS, Dvorak HF. Tumor cells secrete a
 vascular permeability factor that promotes accumulation of ascitesfluid. Science
 1983;219(4587):983-5.
- 1207 77. Leung DW, Cachianes G, Kuang WJ, Goeddel DV, Ferrara N. Vascular endothelial growth
 1208 factor is a secreted angiogenic mitogen. Science 1989;246(4935):1306-9.
- **78.** Keck PJ, Hauser SD, Krivi G, Sanzo K, Warren T, Feder J, Connolly DT. Vascular permeability
 factor, an endothelial cell mitogen related to PDGF. Science 1989;246(4935):1309-12.
- 1213 79. Kim KJ, Li B, Winer J, Armanini M, Gillett N, Phillips HS, Ferrara N. Inhibition of vascular
 1214 endothelial growth factor-induced angiogenesis suppresses tumour growth in vivo. Nature
 1215 1993;362(6423):841-4.
- 1216

1219

1223

- 1217 80. Kendall RL, Thomas KA. Inhibition of vascular endothelial cell growth factor activity by an
 1218 endogenously encoded soluble receptor. Proc Natl Acad Sci U S A 1993;90(22):10705-9.
- 81. Green LS, Jellinek D, Bell C, Beebe LA, Feistner BD, Gill SC, Jucker FM, Janjić N. Nucleaseresistant nucleic acid ligands to vascular permeability factor/vascular endothelial growth factor.
 Chem Biol 1995;2(10):683-95.
- 82. Ruckman J, Green LS, Beeson J, Waugh S, Gillette WL, Henninger DD, Claesson-Welsh L, Janjić N. 2'-Fluoropyrimidine RNA-based aptamers to the 165-amino acid form of vascular endothelial growth factor (VEGF165). Inhibition of receptor binding and VEGF-induced vascular permeability through interactions requiring the exon 7-encoded domain. J Biol Chem 1998;273(32):20556-67.
- 83. Healy JM, Lewis SD, Kurz M, Boomer RM, Thompson KM, Wilson C, McCauley TG.
 Pharmacokinetics and biodistribution of novel aptamer compositions. Pharm Res 2004;21(12):2234-46.
 - 1233

1229

1234 84. Bell C, Lynam E, Landfair DJ, Janjic N, Wiles ME. Oligonucleotide NX1838 inhibits
 1235 VEGF165-mediated cellular responses in vitro. In Vitro Cell Dev Biol Anim 1999;35(9):533-42.

- 1236 85. Eyetech Study Group. Preclinical and phase 1A clinical evaluation of an anti-VEGF pegylated
 1237 aptamer (EYE001) for the treatment of exudative age-related macular degeneration. Retina
 1238 2002;22(2):143-52.
- 1239

1246

1250

1255

1258

1262

1266

- 86. Lee JH, Canny MD, De Erkenez A, Krilleke D, Ng YS, Shima DT, Pardi A, Jucker F. A
 therapeutic aptamer inhibits angiogenesis by specifically targeting the heparin binding domain of
 VEGF165. Proc Natl Acad Sci U S A 2005;102(52):18902-7.
- 1244 **87.** Hermann T, Patel DJ. Adaptive recognition by nucleic acid aptamers. Science 2000;287(5454):820-5.
- 1247 88. Bates DO, Mavrou A, Qiu Y, Carter JG, Hamdollah-Zadeh M, Barratt S, Gammons MV, Millar
 1248 AB, Salmon AH, Oltean S, Harper SJ. Detection of VEGF-A(xxx)b isoforms in human tissues.
 1249 PLoS One 2013;8(7):e68399.
- 89. Varey AH, Rennel ES, Qiu Y, Bevan HS, Perrin RM, Raffy S, Dixon AR, Paraskeva C, Zaccheo
 O, Hassan AB, Harper SJ, Bates DO. VEGF 165 b, an antiangiogenic VEGF-A isoform, binds
 and inhibits bevacizumab treatment in experimental colorectal carcinoma: balance of pro- and
 antiangiogenic VEGF-A isoforms has implications for therapy. Br J Cancer 2008;98(8):1366-79.
- 90. Klettner A, Roider J. Comparison of bevacizumab, ranibizumab, and pegaptanib in vitro:
 efficiency and possible additional pathways. Invest Ophthalmol Vis Sci 2008;49(10):4523-7.
- 91. Presta LG, Chen H, O'Connor SJ, Chisholm V, Meng YG, Krummen L, Winkler M, Ferrara N.
 Humanization of an anti-vascular endothelial growth factor monoclonal antibody for the therapy of solid tumors and other disorders. Cancer Res 1997;57(20):4593-9.
- 92. Gragoudas ES, Adamis AP, Cunningham ET Jr, Feinsod M, Guyer DR; VEGF Inhibition Study
 in Ocular Neovascularization Clinical Trial Group. Pegaptanib for neovascular age-related
 macular degeneration. N Engl J Med 2004;351(27):2805-16.
- 93. VEGF Inhibition Study in Ocular Neovascularization (V.I.S.I.O.N.) Clinical Trial Group,
 Chakravarthy U, Adamis AP, Cunningham ET Jr, Goldbaum M, Guyer DR, Katz B, Patel M.
 Year 2 efficacy results of 2 randomized controlled clinical trials of pegaptanib for neovascular
 age-related macular degeneration. Ophthalmology 2006;113(9):1508.e1-25.
- 1271
- 94. Gonzales CR; VEGF Inhibition Study in Ocular Neovascularization (V.I.S.I.O.N.) Clinical Trial
 Group. Enhanced efficacy associated with early treatment of neovascular age-related macular
 degeneration with pegaptanib sodium: an exploratory analysis. Retina 2005;25(7):815-27.
- 95. Quiram PA, Hassan TS, Williams GA. Treatment of naïve lesions in neovascular age-related
 macular degeneration with pegaptanib. Retina 2007;27(7):851-6.
- 1278

- 96. Nishimura Y, Taguchi M, Nagai T, Fujihara M, Honda S, Uenishi M. Comparison of the effect
 between pegaptanib and ranibizumab on exudative age-related macular degeneration with small
 lesion size. Clin Ophthalmol 2012;6:365-8.
- 1282
- 97. Hughes MS, Sang DN. Safety and efficacy of intravitreal bevacizumab followed by pegaptanib
 maintenance as a treatment regimen for age-related macular degeneration. Ophthalmic Surg
 Lasers Imaging 2006;37(6):446-54.
- 98. Friberg TR, Tolentino M; LEVEL Study Group, Weber P, Patel S, Campbell S, Goldbaum M.
 Pegaptanib sodium as maintenance therapy in neovascular age-related macular degeneration:
 the LEVEL study. Br J Ophthalmol 2010;94(12):1611-7.
- 1290

1298

1286

- 99. Ishibashi T; LEVEL-J Study Group. Maintenance therapy with pegaptanib sodium for
 neovascular age-related macular degeneration: an exploratory study in Japanese patients
 (LEVEL-J study). Jpn J Ophthalmol 2013;57(5):417-23.
- 100.Inoue M, Kadonosono K, Arakawa A, Yamane S, Ishibashi T. Long-term outcome of intravitreal
 pegaptanib sodium as maintenance therapy in Japanese patients with neovascular age-related
 macular degeneration. Jpn J Ophthalmol 2015;59(3):173-8.
- 101.Adamis AP, Altaweel M, Bressler NM, Cunningham ET Jr, Davis MD, Goldbaum M,
 Gonzales C, Guyer DR, Barrett K, Patel M; Macugen Diabetic Retinopathy Study
 Group.Changes in retinal neovascularization after pegaptanib (Macugen) therapy in diabetic
 individuals. Ophthalmology 2006;113(1):23-8.
- 1303

1306

1309

- 1304 102.Querques G, Bux AV, Martinelli D, Iaculli C, Noci ND. Intravitreal pegaptanib sodium
 1305 (Macugen) for diabetic macular oedema. Acta Ophthalmol 2009;87(6):623-30. (a)
- 1307 103.Salam A, Mathew R, Sivaprasad S.Treatment of proliferative diabetic retinopathy with anti 1308 VEGF agents. Acta Ophthalmol 2011;89(5):405-11.
- 104.Rinaldi M, Chiosi F, Dell'Omo R, Romano MR, Parmeggiani F, Semeraro F, Menzione M,
 Costagliola C. Intravitreal pegaptanib sodium (Macugen) for treatment of myopic choroidal
 neovascularization: a morphologic and functional study. Retina 2013;33(2):397-402.
- 1313
- 105.Martìnez-Zapata MJ, Martí-Carvajal AJ, Solà I, Pijoán JI, Buil-Calvo JA, Cordero JA, Evans
 JR. Anti-vascular endothelial growth factor for proliferative diabetic retinopathy. Cochrane
 Database Syst Rev 2014;11:CD008721.
- 1318 106.Virgili G, Parravano M, Menchini F, Evans JR. Anti-vascular endothelial growth factor for
 1319 diabetic macular oedema. Cochrane Database Syst Rev 2014;10:CD007419.
- 1320

- 1321 107.Braithwaite T, Nanji AA, Lindsley K, Greenberg PB. Anti-vascular endothelial growth factor
 1322 for macular oedema secondary to central retinal vein occlusion. Cochrane Database Syst Rev
 1323 2014;5:CD007325.
- 1324

1334

1337

- 108.Inai T, Mancuso M, Hashizume H, Baffert F, Haskell A, Baluk P, Hu-Lowe DD, Shalinsky
 DR, Thurston G, Yancopoulos GD, McDonald DM. Inhibition of vascular endothelial growth
 factor (VEGF) signaling in cancer causes loss of endothelial fenestrations, regression of tumor
 vessels, and appearance of basement membrane ghosts. Am J Pathol 2004;165(1):35-52.
- 109.Kamba T, Tam BY, Hashizume H, Haskell A, Sennino B, Mancuso MR, Norberg SM, O'Brien
 SM, Davis RB, Gowen LC, Anderson KD, Thurston G, Joho S, Springer ML, Kuo CJ,
 McDonald DM. VEGF-dependent plasticity of fenestrated capillaries in the normal adult
 microvasculature. Am J Physiol Heart Circ Physiol 2006;290(2):H560-76.
- 1335 110.Falavarjani KG, Nguyen QD. Adverse events and complications associated with intravitreal
 1336 injection of anti-VEGF agents: a review of literature. Eye (Lond) 2013;27(7):787-94.
- 111.Biagi C, Conti V, Montanaro N, Melis M, Buccellato E, Donati M, Covezzoli A, Amato R,
 Pazzi L, Venegoni M, Vaccheri A, Motola D. Comparative safety profiles of intravitreal
 bevacizumab, ranibizumab and pegaptanib: the analysis of the WHO database of adverse drug
 reactions. Eur J Clin Pharmacol 2014;70(12):1505-12..
- 1343 112.Penedones A, Mendes D, Alves C, Batel Marques F. Safety monitoring of ophthalmic
 1344 biologics: a systematic review of pre- and postmarketing safety data. J Ocul Pharmacol Ther
 1345 2014;30(9):729-51..
- 1346

1350

1342

- 1347 113.Rosenfeld PJ, Shapiro H, Tuomi L, Webster M, Elledge J, Blodi B; MARINA and ANCHOR
 1348 Study Groups. Characteristics of patients losing vision after 2 years of monthly dosing in the
 1349 phase III ranibizumab clinical trials. Ophthalmology 2011;118(3):523-30.
- 1351 114.Lois N, McBain V, Abdelkader E, Scott NW, Kumari R. Retinal pigment epithelial atrophy in
 patients with exudative age-related macular degeneration undergoing anti-vascular endothelial
 growth factor therapy. Retina 2013;33(1):13-22.
- 1354
- 1355 115.Bhutto I, Lutty G. Understanding age-related macular degeneration (AMD): relationships
 1356 between the photoreceptor/retinal pigment epithelium/Bruch's membrane/choriocapillaris
 1357 complex. Mol Aspects Med 2012;33(4):295-317.
- 1359 116.Holz FG, Strauss EC, Schmitz-Valckenberg S, van Lookeren Campagne M. Geographic
 1360 atrophy: clinical features and potential therapeutic approaches. Ophthalmology
 1361 2014;121(5):1079-91.
- 1362

1358

1363 117.Grunwald JE, Daniel E, Huang J, Ying GS, Maguire MG, Toth CA, Jaffe GJ, Fine SL, Blodi B,
1364 Klein ML, Martin AA, Hagstrom SA, Martin DF; CATT Research Group. Risk of geographic

- 1365 atrophy in the comparison of age-related macular degeneration treatments trials.1366 Ophthalmology 2014;121(1):150-61.
- 1367

1376

1380

1384

- 1368 118.Grunwald JE, Pistilli M, Ying GS, Maguire MG, Daniel E, Martin DF; Comparison of Agerelated Macular Degeneration Treatments Trials Research Group. Growth of geographic
 1370 atrophy in the comparison of age-related macular degeneration treatments trials.
 1371 Ophthalmology 2015;122(4):809-16.
- 1373 119.Rofagha S, Bhisitkul RB, Boyer DS, Sadda SR, Zhang K; SEVEN-UP Study Group. Seven1374 year outcomes in ranibizumab-treated patients in ANCHOR, MARINA, and HORIZON: a
 1375 multicenter cohort study (SEVEN-UP). Ophthalmology 2013;120(11):2292-9.
- 1377 120.Querques G, Bux AV, Delle Noci N. Foveal geographic atrophy following intravitreal
 1378 pegaptanib sodium (Macugen) for drusenoid pigment epithelium detachment. Eur J Ophthalmol
 1379 2009;19(5):890-3. (b)
- 121.Gutfleisch M, Heimes B, Schumacher M, Dietzel M, Lommatzsch A, Bird A, Pauleikhoff D.
 Long-term visual outcome of pigment epithelial tears in association with anti-VEGF therapy of
 pigment epithelial detachment in AMD. Eye (Lond) 2011;25(9):1181-6.
- 1385 122.Daniel E, Toth CA, Grunwald JE, Jaffe GJ, Martin DF, Fine SL, Huang J, Ying GS, Hagstrom
 1386 SA, Winter K, Maguire MG; Comparison of Age-related Macular Degeneration Treatments
 1387 Trials Research Group. Risk of scar in the comparison of age-related macular degeneration
 1388 treatments trials. Ophthalmology 2014;121(3):656-66.
- 1389

1392

- 1390 123.Chang LK, Sarraf D. Tears of the retinal pigment epithelium: an old problem in a new era.
 1391 Retina 2007;27(5):523-34.
- 124.Doguizi S, Ozdek S. Pigment epithelial tears associated with anti-VEGF therapy: incidence,
 long-term visual outcome, and relationship with pigment epithelial detachment in age-related
 macular degeneration. Retina 2014;34(6):1156-1162.
- 1397 125.Sarraf D, Joseph A, Rahimy E. Retinal pigment epithelial tears in the era of intravitreal
 pharmacotherapy: risk factors, pathogenesis, prognosis and treatment (an American
 Ophthalmological Society thesis). Trans Am Ophthalmol Soc 2014;112:142-59.
- 1400
- 126.Li Z, Van Bergen T, Van de Veire S, Van de Vel I, Moreau H, Dewerchin M, Maudgal PC,
 Zeyen T, Spileers W, Moons L, Stalmans I. Inhibition of vascular endothelial growth factor
 reduces scar formation after glaucoma filtration surgery. Invest Ophthalmol Vis Sci
 2009;50(11):5217-25.
- 1405

^{1406 127.}Van Bergen T, Vandewalle E, Van de Veire S, Dewerchin M, Stassen JM, Moons L, Stalmans
1407 I. The role of different VEGF isoforms in scar formation after glaucoma filtration surgery. Exp
1408 Eye Res 2011;93(5):689-99.

- 128.Bonnin P, Pournaras JA, Lazrak Z, Cohen SY, Legargasson JF, Gaudric A, Levy BI, Massin P.
 Ultrasound assessment of short-term ocular vascular effects of intravitreal injection of bevacizumab (Avastin®) in neovascular age-related macular degeneration. Acta Ophthalmol 2010;88(6):641-5.
- 1413
- 1414 129.Papadopoulou DN, Mendrinos E, Mangioris G, Donati G, Pournaras CJ. Intravitreal
 1415 ranibizumab may induce retinal arteriolar vasoconstriction in patients with neovascular age 1416 related macular degeneration. Ophthalmology 2009;116(9):1755-61.
- 1417

1426

1429

- 130.Bonnin P, Pournaras JA, Makowiecka K, Krivosic V, Kedra AW, Le Gargasson JF, Gaudric A,
 Levy BI, Cohen YS, Tadayoni R, Massin P. Ultrasound assessment of ocular vascular effects of
 repeated intravitreal injections of ranibizumab for wet age-related macular degeneration. Acta
 Ophthalmol 2014;92(5):e382-7.
- 1423 131.Hussain RM, Harris A, Siesky B, Yung CW, Ehrlich R, Prall R. The effect of pegaptanib
 (Macugen®) injection on retinal and retrobulbar blood flow in retinal Ischaemic diseases. Acta
 Ophthalmol 2015 Jan 4.
- 1427 132.Kim H, Robinson SB, Csaky KG. FcRn receptor-mediated pharmacokinetics of therapeutic IgG
 1428 in the eye. Mol Vis 2009;15:2803-12.
- 133.Matsuyama K, Ogata N, Matsuoka M, Wada M, Takahashi K, Nishimura T. Plasma levels of
 vascular endothelial growth factor and pigment epithelium-derived factor before and after
 intravitreal injection of bevacizumab. Br J Ophthalmol 2010;94(9):1215-8.
- 1433

1436

134.Hayman SR, Leung N, Grande JP, Garovic VD. VEGF inhibition, hypertension, and renal toxicity. Curr Oncol Rep 2012;14(4):285-94.

- 1437 135.Duan Y, Mo J, Klein R, Scott IU, Lin HM, Caulfield J, Patel M, Liao D. Age-related macular
 1438 degeneration is associated with incident myocardial infarction among elderly Americans.
 1439 Ophthalmology 2007;114(4):732-7.
- 136.Tuñón J, Ruiz-Moreno JM, Martìn-Ventura JL, Blanco-Colio LM, Lorenzo O, Egido J.
 Cardiovascular risk and antiangiogenic therapy for agerelated macular degeneration. Surv
 Ophthalmol 2009;54(3):339-48.
- 1444

1440

- 137.Curtis LH, Hammill BG, Schulman KA, Cousins SW. Risks of mortality, myocardial
 infarction, bleeding, and stroke associated with therapies for age-related macular degeneration.
 Arch Ophthalmol 2010;128(10):1273-9.
- 138.Ikram MK, Mitchell P, Klein R, Sharrett AR, Couper DJ, Wong TY. Age-related macular
 degeneration and long-term risk of stroke subtypes. Stroke 2012;43(6):1681-3.
- 1451

- 139.Mancia et al. ESH/ESC Task Force for the Management of Arterial Hypertension. 2013
 Practice guidelines for the management of arterial hypertension of the European Society of
 Hypertension (ESH) and the European Society of Cardiology (ESC): ESH/ESC Task Force for
 the Management of Arterial Hypertension. J Hypertens 2013;31(10):1925-38.
- 1456

- 1457 140.Krause L, Yousif T, Pohl K; CAPTAIN study group. An epidemiological study of neovascular
 1458 age-related macular degeneration in Germany. Curr Med Res Opin 2013;29(10):1391-7.
- 1460 141.Wong TY, Klein R, Sun C, Mitchell P, Couper DJ, Lai H, Hubbard LD, Sharrett AR;
 1461 Atherosclerosis Risk in Communities Study. Age-related macular degeneration and risk of
 1462 stroke. Ann Intern Med 2006;145(2):98-107.
- 1463
- 1464 142.Semeraro F, Morescalchi F, Duse S, Gambicorti E, Romano MR, Costagliola C. Systemic
 1465 thromboembolic adverse events in patients treated with intravitreal anti-VEGF drugs for
 1466 neovascular age-related macular degeneration: an overview. Expert Opin Drug Saf.
 1467 2014;13(6):785-802.
- 1469 143.Singerman LJ, Masonson H, Patel M, Adamis AP, Buggage R, Cunningham E, Goldbaum M,
 1470 Katz B, Guyer D. Pegaptanib sodium for neovascular age-related macular degeneration: third1471 year safety results of the VEGF Inhibition Study in Ocular Neovascularisation (VISION) trial.
 1472 Br J Ophthalmol 2008;92:1606-11.
- 1473

1468

- 1474 144.Chong V. Biological, preclinical and clinical characteristics of inhibitors of vascular
 1475 endothelial growth factors. Ophthalmologica 2012;227 Suppl 1:2-10.
- 1476

1479

1483

- 1477 145.Morjaria R, Chong NV. Pharmacokinetic evaluation of pegaptanib octasodium for the
 1478 treatment of diabetic edema. Expert Opin Drug Metab Toxicol 2014;10(8):1185-92.
- 1480 146.Zachary AA, Montgomery RA, Leffell MS. Factors associated with and predictive of
 persistence of donor-specific antibody after treatment with plasmapheresis and intravenous
 immunoglobulin. Hum Immunol 2005;66(4):364-70.
- 1484 147.Bressler NM, Boyer DS, Williams DF, Butler S, Francom SF, Brown B, Di Nucci F, Cramm T, Tuomi
 1485 LL, Ianchulev T, Rubio RG. Cerebrovascular accidents in patients treated for choroidal
 1486 neovascularization with ranibizumab in randomized controlled trials. Retina 2012;32(9):1821-8.
- 1487

- 1488 148.Ueta T, Noda Y, Toyama T, Yamaguchi T, Amano S. Systemic vascular safety of ranibizumab
 1489 for age-related macular degeneration: systematic review and meta-analysis of randomized
 1490 trials. Ophthalmology 2014;121(11):2193-203.e1-7.
- 1492 149.Verheyen A, Peeraer E, Nuydens R, Dhondt J, Poesen K, Pintelon I, Daniels A, Timmermans
 1493 JP, Meert T, Carmeliet P, Lambrechts D. Systemic anti-vascular endothelial growth factor
 1494 therapies induce a painful sensory neuropathy. Brain 212;135:2629–41.
- 1495

- 1496 150.Hulse RP, Beazley-Long N, Hua J, Kennedy H, Prager J, Bevan H, Qiu Y, Fernandes ES,
 1497 Gammons MV, Ballmer-Hofer K, Gittenberger de Groot AC, Churchill AJ, Harper SJ, Brain SD,
 1498 Bates DO, Donaldson LF. Regulation of alternative VEGF-A mRNA splicing is a therapeutic
 1499 target for analgesia. Neurobiol Dis 2014;71:245-59.
- 1501 151.Zhu X, Wu S, Dahut WL, Parikh CR. Risks of proteinuria and hypertension with bevacizumab,
 1502 an antibody against vascular endothelial growth factor: systematic review and meta-analysis. Am
 1503 J Kidney Dis 2007;49:186–193.
- 1505 152.Hayman SR, Leung N, Grande JP, Garovic VD. VEGF inhibition, hypertension, and renal toxicity. Curr Oncol Rep 2012;14(4):285-94.
- 1508 153.Kitamoto Y, Tokunaga H, Miyamoto K, Tomita K. VEGF is an essential molecule for
 1509 glomerular structuring. Nephrol Dial Transplant 2002;17 Suppl 9:25-7
- 1511 154.Eremina V, Quaggin SE. The role of VEGF-A in glomerular development and function. Curr
 1512 Opin Nephrol Hypertens 2004;13(1):9-15.
- 1514 155.Suga S, Kim YG, Joly A, Puchacz E, Kang DH, Jefferson JA, Abraham JA, Hughes J, Johnson
 RJ, Schreiner GF. Vascular endothelial growth factor (VEGF121) protects rats from renal
 infarction in thrombotic microangiopathy. Kidney Int 2001;60(4):1297-308.
- 1518 156.Feihl F, Liaudet L, Waeber B, Levy BI. Hypertension: a disease of the microcirculation?
 1519 Hypertension 2006;48:1012–1017.
- 1520

1527

1517

1500

1504

1507

1510

1513

- 1521 157.Neri P, Mariotti C, Arapi I, Bambini E, Giovannini A. Anti vascular endothelial growth factor
 1522 sequential therapy for neovascular age-related macular degeneration: is this the new deal? Curr
 1523 Med Res Opin 2012;28(3):395-400.
- 1525 158.Ehlken C, Jungmann S, Böhringer D, Agostini HT, Junker B, Pielen A. Switch of anti-VEGF
 agents is an option for nonresponders in the treatment of AMD. Eye (Lond) 2014;28(5):538-45.
- 1528 159.Binder S. Loss of reactivity in intravitreal anti-VEGF therapy: tachyphylaxis or tolerance? Br J
 1529 Ophthalmol 2012;96(1):1-2.
- 1530
- 160.Keane PA, Liakopoulos S, Ongchin SC, Heussen FM, Msutta S, Chang KT, Walsh AC, Sadda
 SR. Quantitative subanalysis of optical coherence tomography after treatment with ranibizumab
 for neovascular age-related macular degeneration. Invest Ophthalmol Vis Sci 2008;
 49(7):3115-20.
- 1536 161.Schaal S, Kaplan HJ, Tezel TH. Is there tachyphylaxis to intravitreal anti-vascular endothelial
 1537 growth factor pharmacotherapy in age-related macular degeneration? Ophthalmology 2008;
 1538 115(12):2199-205.
- 1539

- 1540 162.Forooghian F, Cukras C, Meyerle CB, Chew EY, Wong WT. Tachyphylaxis after intravitreal
 1541 bevacizumab for exudative age-related macular degeneration. Retina 2009;29(6):723-31.
- 1542

1549

1553

1558

1561

1566

1570

- 1543 163.Rosenfeld PJ, Brown DM, Heier JS, Boyer DS, Kaiser PK, Chung CY, Kim RY. Ranibizumab
 1544 for neovascular agerelated macular degeneration. N Engl J Med 2006;355:1419-31.
- 1546 164.Forooghian F, Chew EY, Meyerle CB, Cukras C, Wong WT. Investigation of the role of
 neutralizing antibodies against bevacizumab as mediators of tachyphylaxis. Acta Ophthalmol
 1548 2011;89(2):e206-7.
- 165.Abedi F, Wickremasinghe S, Richardson AJ, Makalic E, Schmidt DF, Sandhu SS, Baird PN,
 Guymer RH. Variants in the VEGFA gene and treatment outcome after anti-VEGF treatment
 for neovascular age-related macular degeneration. Ophthalmology 2013;120(1):115-21.
- 166.Stepien KE, Rosenfeld PJ, Puliafito CA, Feuer W, Shi W, Al-Attar L, Dubovy SR, Murray TG,
 Davis JL, Lee WH, Schwartz SG, Smiddy WE, Berrocal AM, Flynn HW Jr. Comparison of
 intravitreal bevacizumab followed by ranibizumab for the treatment of neovascular age-related
 macular degeneration. Retina 2009;29(8):1067-73.
- 1559 167.Almony A, Mansouri A, Shah GK, Blinder KJ. Efficacy of intravitreal bevacizumab after
 unresponsive treatment with intravitreal ranibizumab. Can J Ophthalmol 2011;46(2):182-5.
- 168.Kumar N, Marsiglia M, Mrejen S, Fung AT, Slakter J, Sorenson J, Freund KB. Visual and
 anatomical outcomes of intravitreal aflibercept in eyes with persistent subfoveal fluid despite
 previous treatments with ranibizumab in patients with neovascular age-related macular
 degeneration. Retina 2013; 33(8):1605-12.
- 169.Shiragami C, Ono A, Kobayashi M, Manabe S, Yamashita A, Shiraga F. Effect of switching
 therapy to pegaptanib in eyes with the persistent cases of exudative age-related macular
 degeneration. Medicine (Baltimore) 2014;93(18):e116.
- 170.Jo N, Mailhos C, Ju M, Cheung E, Bradley J, Nishijima K, Robinson GS, Adamis AP, Shima DT. Inhibition of platelet-derived growth factor B signaling enhances the efficacy of anti-vascular endothelial growth factor therapy in multiple models of ocular neovascularization. Am J Pathol 2006;168(6):2036-53.
- 1575

1578

- 1576 171.Lindahl P, Johansson BR, Leveen P, Betsholtz C. Pericyte loss and microaneurysm formation
 1577 in PDGF-B-deficient mice. Science 1997;277:242-5.
- 172.Ishikawa M, Jin D, Sawada Y, Abe S, Yoshitomi T. Future therapies of wet age-related
 macular degeneration. J Ophthalmol 2015; 2015:138070.
- 1582 173.Amadio M., Bucolo C., Leggio G.M., Drago F., Govoni S., Pascale A. The
 1583 PKCbeta/HuR/VEGF pathway in diabetic retinopathy. Biochem Pharmacol. 80, 2010.

- 174.Viiri J, Amadio M, Marchesi N, Hyttinen JM, Kivinen N, Sironen R, Rilla K, Akhtar S,
 Provenzani A, D'Agostino VG, Govoni S, Pascale A, Agostini H, Petrovski G, Salminen A,
 Kaarniranta K. Autophagy Activation Clears ELAVL1/HuR-Mediated Accumulation of
 SQSTM1/p62 during Proteasomal Inhibition in Human Retinal Pigment Epithelial Cells. PLoS
 One; 8, 2013.
- 175.Agarwal A, Rhoades WR, Hanout M, Soliman MK, Sarwar S, Sadiq MA, Sepah YJ, Do DV,
 Nguyen QD. Management of neovascular age-related macular degeneration: current state-of the-art care for optimizing visual outcomes and therapies in development. Clin Ophthalmol
 2015; 9:1001-15.
- 1594

1600

1589

- 1595 176.Horie-Inoue K, Inoue S. Genomic aspects of age-related macular degeneration. Biochem
 1596 Biophys Res Commun 2014;452(2):263-75.
- 1598 177.Hu Z, Xie P, Ding Y, Yuan D, Liu Q. Association between variants A69S in ARMS2 gene and
 1599 response to treatment of exudative AMD: a meta-analysis. Br J Ophthalmol 2015;99(5):593-8.
- 1601 178.Kanwar JR, Shankaranarayanan JS, Gurudevan S, Kanwar RK. Aptamer-based therapeutics of
 1602 the past, present and future: from the perspective of eye-related diseases. Drug Discov Today
 1603 2014; 19(9):1309-21.
- 1604

1607

1610

- 1605 179.Tolentino MJ, Dennrick A, John E, Tolentino MS. Drugs in Phase II clinical trials for the
 treatment of age-related macular degeneration. Expert Opin Investig Drugs 2015;24(2):183-99.
- 1608 180.Chen S, Feng J, Ma L, Liu Z, Yuan W. RNA interference technology for anti-VEGF treatment.
 1609 Expert Opin Drug Deliv 2014; 11(9):1471-80.
- 1611 181.Reich SJ, Fosnot J, Kuroki A, Tang W, Yang X, Maguire AM, Bennett J, Tolentino MJ. Small
 1612 interfering RNA (siRNA) targeting VEGF effectively inhibits ocular neovascularization in a
 1613 mouse model. Mol Vis 2003; 9:210-6.
- 1614

1619

- 182.Tolentino MJ, Brucker AJ, Fosnot J, Ying GS, Wu IH, Malik G, Wan S, Reich SJ. Intravitreal
 injection of vascular endothelial growth factor small interfering RNA inhibits growth and
 leakage in a nonhuman primate, laser-induced model of choroidal neovascularization. Retina
 2004; 24(1):132-8.
- 1620 183.Dejneka NS, Wan S, Bond OS, Kornbrust DJ, Reich SJ. Ocular biodistribution of bevasiranib
 1621 following a single intravitreal injection to rabbit eyes. Mol Vis 2008; 14:997-1005.
- 184.Shen J, Samul R, Silva RL, Akiyama H, Liu H, Saishin Y, Hackett SF, Zinnen S, Kossen K,
 Fosnaugh K, Vargeese C, Gomez A, Bouhana K, Aitchison R, Pavco P, Campochiaro PA.
 Suppression of ocular neovascularization with siRNA targeting VEGF receptor 1. Gene Ther
 2006; 13(3):225-34.
- 1627

- 185.Kaiser PK, Symons RC, Shah SM, Quinlan EJ, Tabandeh H, Do DV, Reisen G, Lockridge JA,
 Short B, Guerciolini R, Nguyen QD; Sirna-027 Study Investigators. RNAi-based treatment for
 neovascular age-related macular degeneration by Sirna-027. Am J Ophthalmol 2010;
 150(1):33-9.
- 1632

1641

1646

- 186.Rittenhouse KD, Johnson TR, Vicini P, Hirakawa B, Kalabat D, Yang AH, Huang W, Basile
 AS. RTP801 gene expression is differentially upregulated in retinopathy and is silenced by PF04523655, a 19-Mer siRNA directed against RTP801. Invest Ophthalmol Vis Sci 2014;
 55(3):1232-40.
- 1638 187.Brafman A, Mett I, Shafir M, Gottlieb H, Damari G, Gozlan-Kelner S, Vishnevskia-Dai V,
 1639 Skaliter R, Einat P, Faerman A, Feinstein E, Shoshani T. Inhibition of oxygen-induced
 1640 retinopathy in RTP801-deficient mice. Invest Ophthalmol Vis Sci 2004; 45(10):3796-805.
- 188.Nguyen QD, Schachar RA, Nduaka CI, Sperling M, Basile AS, Klamerus KJ, Chi-Burris K,
 Yan E, Paggiarino DA, Rosenblatt I, Khan A, Aitchison R, Erlich SS; PF-04523655 Study
 Group. Phase 1 dose-escalation study of a siRNA targeting the RTP801 gene in age-related
 macular degeneration patients. Eye (Lond) 2012; 26(8):1099-105. (a)
- 1647 189.Nguyen QD, Schachar RA, Nduaka CI, Sperling M, Basile AS, Klamerus KJ, Chi-Burris K,
 1648 Yan E, Paggiarino DA, Rosenblatt I, Aitchison R, Erlich SS; DEGAS Clinical Study Group.
 1649 Dose-ranging evaluation of intravitreal siRNA PF-04523655 for diabetic macular edema (the
 1650 DEGAS study). Invest Ophthalmol Vis Sci 2012; 53(12):7666-74. (b)
- 1651

1656

1659

- 190.Nguyen QD, Schachar RA, Nduaka CI, Sperling M, Klamerus KJ, Chi-Burris K, Yan E,
 Paggiarino DA, Rosenblatt I, Aitchison R, Erlich SS; MONET Clinical Study Group.
 Evaluation of the siRNA PF-04523655 versus ranibizumab for the treatment of neovascular
 age-related macular degeneration (MONET Study). Ophthalmology 2012; 119(9):1867-73. (c)
- 1657 191.Sarli V, Giannis A. Targeting the kinesin spindle protein: basic principles and clinical
 1658 implications. Clin Cancer Res 2008; 14(23):7583-7.
- 192. Tabernero J, Shapiro GI, LoRusso PM, Cervantes A, Schwartz GK, Weiss GJ, Paz-Ares L, Cho
 DC, Infante JR, Alsina M, Gounder MM, Falzone R, Harrop J, White AC, Toudjarska I,
 Bumcrot D, Meyers RE, Hinkle G, Svrzikapa N, Hutabarat RM, Clausen VA, Cehelsky J,
 Nochur SV, Gamba-Vitalo C, Vaishnaw AK, Sah DW, Gollob JA, Burris HA 3rd. First-inhumans trial of an RNA interference therapeutic targeting VEGF and KSP in cancer patients
 with liver involvement. Cancer Discov 2013; 3(4):406-17.
- 1667 193.Nonaka Y, Sode K, Ikebukuro K. Screening and improvement of an anti-VEGF DNA aptamer.
 1668 Molecules 2010;15(1):215-25.
- 1669

- 1670 194.Nonaka Y, Yoshida W, Abe K, Ferri S, Schulze H, Bachmann TT, Ikebukuro K. Affinity
 1671 improvement of a VEGF aptamer by in silico maturation for a sensitive VEGF-detection
 1672 system. Anal Chem 2013;85(2):1132-7.
- 1673

- 1674 195.Benjamin LE, Hemo I, Keshet E. A plasticity window for blood vessel remodelling is defined
 by pericyte coverage of the preformed endothelial network and is regulated by PDGF-B and
 VEGF. Development 1998; 125(9):1591-8.
- 1678 196.Choudhury P, Chen W, Hunt RC. Production of platelet-derived growth factor by interleukin-1
 beta and transforming growth factor-beta-stimulated retinal pigment epithelial cells leads to
 contraction of collagen gels. Invest Ophthalmol Vis Sci 1997;38(5):824-33.
- 1682 197.Bradley J, Ju M, Robinson GS. Combination therapy for the treatment of ocular
 1683 neovascularization. Angiogenesis 2007;10(2):141-8.
- 1684

1687

- 1685 198.Zarbin MA, Rosenfeld PJ. Pathway-based therapies for age-related macular degeneration: an
 1686 integrated survey of emerging treatment alternatives. Retina 2010;30(9):1350-67.
- 1688 199.Mulligan MS, Schmid E, Till GO, Hugli TE, Friedl HP, Roth RA, Ward PA. C5a-dependent
 up-regulation in vivo of lung vascular P-selectin. J Immunol 1997;158(4):1857-61.